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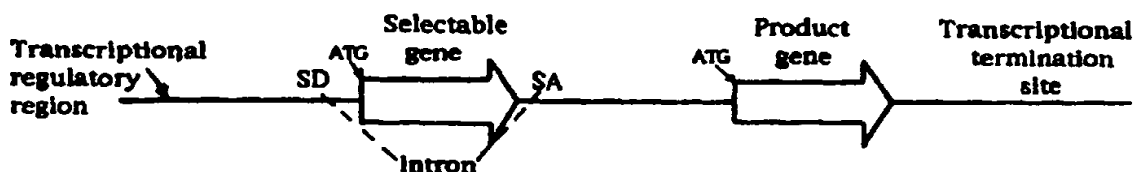
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(54) Title: METHOD FOR SELECTING HIGH-EXPRESSING HOST CELLS

**(57) Abstract**

A method for selecting recombinant host cells expressing high levels of a desired protein is described. This method utilizes eukaryotic host cells harboring a DNA construct comprising a selectable gene (preferably an amplifiable gene) and a product gene provided 3' to the selectable gene. The selectable gene is positioned within an intron defined by a splice donor site and a splice acceptor site and the selectable gene and product gene are under the transcriptional control of a single transcriptional regulatory region. The splice donor site is generally an efficient splice donor site and thereby regulates expression of the product gene using the transcriptional regulatory region. The transfected cells are cultured so as to express the gene encoding the product in a selective medium comprising an amplifying agent for sufficient time to allow amplification to occur, whereupon either the desired product is recovered or cells having multiple copies of the product gene are identified.

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METHOD FOR SELECTING HIGH-EXPRESSING HOST CELLSBACKGROUND OF THE INVENTIONField of the Invention

This invention relates to a method of selecting for high-expressing host cells, a method of producing a protein of interest in high yields and a method of producing eukaryotic cells having multiple copies of a sequence encoding a protein of interest.

Description of Background and Related Art

The discovery of methods for introducing DNA into living host cells in a functional form has provided the key to understanding many fundamental biological processes, and has made possible the production of important proteins and other molecules in commercially useful quantities.

Despite the general success of such gene transfer methods, several common problems exist that may limit the efficiency with which a gene encoding a desired protein can be introduced into and expressed in a host cell. One problem is knowing when the gene has been successfully transferred into recipient cells. A second problem is distinguishing between those cells that contain the gene and those that have survived the transfer procedures but do not contain the gene. A third problem is identifying and isolating those cells that contain the gene and that are expressing high levels of the protein encoded by the gene.

In general, the known methods for introducing genes into eukaryotic cells tend to be highly inefficient. Of the cells in a given culture, only a small proportion take up and express exogenously added DNA, and an even smaller proportion stably maintain that DNA.

Identification of those cells that have incorporated a product gene encoding a desired protein typically is achieved by introducing into the same cells another gene, commonly referred to as a selectable gene, that encodes a selectable marker. A selectable marker is a protein that is necessary for the growth or survival of a host cell under the particular culture conditions chosen, such as an enzyme that confers resistance to an antibiotic or other drug, or an enzyme that compensates for a metabolic or catabolic defect in the host cell. For example, selectable genes commonly used with eukaryotic cells include the genes for aminoglycoside phosphotransferase (APH), hygromycin phosphotransferase (hyg), dihydrofolate reductase (DHFR), thymidine kinase (tk), neomycin, puromycin, glutamine synthetase, and asparagine synthetase.

The method of identifying a host cell that has incorporated one gene on the basis of expression by the host cell of a second incorporated gene encoding a selectable marker is referred to as cotransfection (or cotransfection). In that method, a gene encoding a desired polypeptide and a selection gene typically are introduced into the host cell simultaneously, although they may be introduced sequentially. In the case of simultaneous cotransfection, the gene encoding the desired polypeptide

and the selectable gene may be present on a single DNA molecule or on separate DNA molecules prior to being introduced into the host cells. Wigler et al., Cell, 16:777 (1979). Cells that have incorporated the gene encoding the desired polypeptide then are identified or isolated by
5 culturing the cells under conditions that preferentially allow for the growth or survival of those cells that synthesize the selectable marker encoded by the selectable gene.

The level of expression of a gene introduced into a eukaryotic host cell depends on multiple factors, including gene copy number, efficiency
10 of transcription, messenger RNA (mRNA) processing, stability, and translation efficiency. Accordingly, high level expression of a desired polypeptide typically will involve optimizing one or more of those factors.

For example, the level of protein production may be increased by covalently joining the coding sequence of the gene to a "strong" promoter
15 or enhancer that will give high levels of transcription. Promoters and enhancers are nucleotide sequences that interact specifically with proteins in a host cell that are involved in transcription. Kriegler, Meth. Enzymol., 185:512 (1990); Maniatis et al., Science, 236:1237 (1987). Promoters are located upstream of the coding sequence of a gene and
20 facilitate transcription of the gene by RNA polymerase. Among the eukaryotic promoters that have been identified as strong promoters for high-level expression are the SV40 early promoter, adenovirus major late promoter, mouse metallothionein-I promoter, Rous sarcoma virus long terminal repeat, and human cytomegalovirus immediate early promoter (CMV).

Enhancers stimulate transcription from a linked promoter. Unlike
25 promoters, enhancers are active when placed downstream from the transcription initiation site or at considerable distances from the promoter, although in practice enhancers may overlap physically and functionally with promoters. For example, all of the strong promoters
30 listed above also contain strong enhancers. Bendig, Genetic Engineering, 7:91 (Academic Press, 1988).

The level of protein production also may be increased by increasing the gene copy number in the host cell. One method for obtaining high gene copy number is to directly introduce into the host cell multiple copies of
35 the gene, for example, by using a large molar excess of the product gene relative to the selectable gene during cotransfection. Kaufman, Meth. Enzymol., 185:537 (1990). With this method, however, only a small proportion of the cotransfected cells will contain the product gene at high copy number. Furthermore, because no generally applicable, convenient
40 method exists for distinguishing such cells from the majority of cells that contain fewer copies of the product gene, laborious and time-consuming screening methods typically are required to identify the desired high-copy number transfectants.

Another method for obtaining high gene copy number involves cloning
45 the gene in a vector that is capable of replicating autonomously in the host cell. Examples of such vectors include mammalian expression vectors

derived from Epstein-Barr virus or bovine papilloma virus, and yeast 2-micron plasmid vectors. Stephens & Hentschel, Biochem. J., 248:1 (1987); Yates et al., Nature, 313:812 (1985); Beggs, Genetic Engineering, 2:175 (Academic Press, 1981).

5 Yet another method for obtaining high gene copy number involves gene amplification in the host cell. Gene amplification occurs naturally in eukaryotic cells at a relatively low frequency. Schimke, J. Biol. Chem., 263:5989 (1988). However, gene amplification also may be induced, or at least selected for, by exposing host cells to appropriate selective
10 pressure. For example, in many cases it is possible to introduce a product gene together with an amplifiable gene into a host cell and subsequently select for amplification of the marker gene by exposing the cotransfected cells to sequentially increasing concentrations of a selective agent. Typically the product gene will be coamplified with the marker gene under
15 such conditions.

The most widely used amplifiable gene for that purpose is a DHFR gene, which encodes a dihydrofolate reductase enzyme. The selection agent used in conjunction with a DHFR gene is methotrexate (Mtx). A host cell is cotransfected with a product gene encoding a desired protein and a DHFR
20 gene, and transfectants are identified by first culturing the cells in culture medium that contains Mtx. A suitable host cell when a wild-type DHFR gene is used is the Chinese Hamster Ovary (CHO) cell line deficient in DHFR activity, prepared and propagated as described by Urlaub & Chasin, Proc. Nat. Acad. Sci. USA, 77:4216 (1980). The transfected cells then are
25 exposed to successively higher amounts of Mtx. This leads to the synthesis of multiple copies of the DHFR gene, and concomitantly, multiple copies of the product gene. Schimke, J. Biol. Chem., 263:5989 (1988); Axel et al., U.S. Patent No. 4,399,216; Axel et al., U.S. Patent No. 4,634,665. Other references directed to co-transfection of a gene together with a genetic
30 marker that allows for selection and subsequent amplification include Kaufman in Genetic Engineering, ed. J. Setlow (Plenum Press, New York), Vol. 9 (1987); Kaufman and Sharp, J. Mol. Biol., 159:601 (1982); Ringold et al., J. Mol. Appl. Genet., 1:165-175 (1981); Kaufman et al., Mol. Cell Biol., 5:1750-1759 (1985); Kaetzel and Nilson, J. Biol. Chem., 263:6244-
35 6251 (1988); Hung et al., Proc. Natl. Acad. Sci. USA, 83:261-264 (1986); Kaufman et al., EMBO J., 6:87-93 (1987); Johnston and Kucey, Science, 242:1551-1554 (1988); Urlaub et al., Cell, 33:405-412 (1983).

To extend the DHFR amplification method to other cell types, a mutant DHFR gene that encodes a protein with reduced sensitivity to methotrexate
40 may be used in conjunction with host cells that contain normal numbers of an endogenous wild-type DHFR gene. Simonsen and Levinson, Proc. Natl. Acad. Sci. USA, 80:2495 (1983); Wigler et al., Proc. Natl. Acad. Sci. USA, 77:3567-3570 (1980); Haber and Schimke, Somatic Cell Genetics, 8:499-508 (1982).

45 Alternatively, host cells may be co-transfected with the product gene, a DHFR gene, and a dominant selectable gene, such as a *neo^r* gene. Kim

and Wold, Cell, 42:129 (1985); Capon et al., U.S. Pat. No. 4,965,199. Transfectants are identified by first culturing the cells in culture medium containing neomycin (or the related drug G418), and the transfectants so identified then are selected for amplification of the DHFR gene and the product gene by exposure to successively increasing amounts of Mtx.

As will be appreciated from this discussion, the selection of recombinant host cells that express high levels of a desired protein generally is a multi-step process. In the first step, initial transfectants are selected that have incorporated the product gene and the selectable gene. In subsequent steps, the initial transfectants are subject to further selection for high-level expression of the selectable gene and then random screening for high-level expression of the product gene. To identify cells expressing high levels of the desired protein, typically one must screen large numbers of transfectants. The majority of transfectants produce less than maximal levels of the desired protein. Further, Mtx resistance in DHFR transformants is at least partially conferred by varying degrees of gene amplification. Schimke, Cell, 37:705-713 (1984). The inadequacies of co-expression of the non-selected gene have been reported by Wold et al., Proc. Natl. Acad. Sci. USA, 76:5684-5688 (1979). Instability of the amplified DNA is reported by Kaufman and Schimke, Mol. Cell Biol., 1:1069-1076 (1981); Haber and Schimke, Cell, 26:355-362 (1981); and Fedespiel et al., J. Biol. Chem., 259:9127-9140 (1984).

Several methods have been described for directly selecting such recombinant host cells in a single step. One strategy involves co-transfecting host cells with a product gene and a DHFR gene, and selecting those cells that express high levels of DHFR by directly culturing in medium containing a high concentration of Mtx. Many of the cells selected in that manner also express the co-transfected product gene at high levels. Page and Sydenham, Bio/Technology, 9:64 (1991). This method for single-step selection suffers from certain drawbacks that limit its usefulness. High-expressing cells obtained by direct culturing in medium containing a high level of a selection agent may have poor growth and stability characteristics, thus limiting their usefulness for long-term production processes. Page and Snyderman, Bio/Technology, 9:64 (1991). Single-step selection for high-level resistance to Mtx may produce cells with an altered, Mtx-resistant DHFR enzyme, or cells that have altered Mtx transport properties, rather than cells containing amplified genes. Haber et al., J. Biol. Chem., 256:9501 (1981); Assaraf and Schimke, Proc. Natl. Acad. Sci. USA, 84:7154 (1987).

Another method involves the use of polycistronic mRNA expression vectors containing a product gene at the 5' end of the transcribed region and a selectable gene at the 3' end. Because translation of the selectable gene at the 3' end of the polycistronic mRNA is inefficient, such vectors exhibit preferential translation of the product gene and require high levels of polycistronic mRNA to survive selection. Kaufman, Meth.

Enzymol., 185:487 (1990); Kaufman, Meth. Enzymol., 185:537 (1990); Kaufman et al., EMBO J., 6:187 (1987). Accordingly, cells expressing high levels of the desired protein product may be obtained in a single step by culturing the initial transfectants in medium containing a selection agent appropriate for use with the particular selectable gene. However, the utility of these vectors is variable because of the unpredictable influence of the upstream product reading frame on selectable marker translation and because the upstream reading frame sometimes becomes deleted during methotrexate amplification (Kaufman et al., J. Mol. Biol., 159:601-621 [1982]; Levinson, Methods in Enzymology, San Diego: Academic Press, Inc. [1990]). Later vectors incorporated an internal translation initiation site derived from members of the picornavirus family which is positioned between the product gene and the selectable gene (Pelletier et al., Nature, 334:320 [1988]; Jang et al., J. Virol., 63:1651 [1989]).

A third method for single-step selection involves use of a DNA construct with a selectable gene containing an intron within which is located a gene encoding the protein of interest. See U.S. Patent No. 5,043,270 and Abrams et al., J. Biol. Chem., 264(24): 14016-14021 (1989). In yet another single-step selection method, host cells are co-transfected with an intron-modified selectable gene and a gene encoding the protein of interest. See WO 92/17566, published October 15, 1992. The intron-modified gene is prepared by inserting into the transcribed region of a selectable gene an intron of such length that the intron is correctly spliced from the corresponding mRNA precursor at low efficiency, so that the amount of selectable marker produced from the intron-modified selectable gene is substantially less than that produced from the starting selectable gene. These vectors help to insure the integrity of the integrated DNA construct, but transcriptional linkage is not achieved as selectable gene and the protein gene are driven by separate promoters.

Other mammalian expression vectors that have single transcription units have been described. Retroviral vectors have been constructed (Cepko et al., Cell, 37:1053-1062 [1984]) in which a cDNA is inserted between the endogenous Moloney murine leukemia virus (M-MuLV) splice donor and splice acceptor sites which are followed by a neomycin resistance gene. This vector has been used to express a variety of gene products following retroviral infection of several cell types.

With the above drawbacks in mind, it is one object of the present invention to increase the level of homogeneity with regard to expression levels of stable clones transfected with a product gene of interest, by expressing a selectable marker (DHFR) and the protein of interest from a single promoter.

It is another object to provide a method for selecting stable, recombinant host cells that express high levels of a desired protein product, which method is rapid and convenient to perform, and reduces the numbers of transfected cells which need to be screened. Furthermore, it is

an object to allow high levels of single and two unit polypeptides to be rapidly generated from clones or pools of stable host cell transfectants.

It is an additional object to provide expression vectors which bias for active integration events (i.e. have an increased tendency to generate transformants wherein the DNA construct is inserted into a region of the genome of the host cell which results in high level expression of the product gene) and can accommodate a variety of product genes without the need for modification.

10

SUMMARY OF THE INVENTION

Accordingly, the present invention is directed to a DNA construct (DNA molecule) alternative terminology comprising a 5' transcriptional initiation site and a 3' transcriptional termination site, a selectable gene (preferably an amplifiable gene) and a product gene provided 3' to the selectable gene, a transcriptional regulatory region regulating transcription of both the selectable gene and the product gene, the selectable gene positioned within an intron defined by a splice donor site and a splice acceptor site. The splice donor site preferably comprises an effective splice donor sequence as herein defined and thereby regulates expression of the product gene using the transcriptional regulatory region.

In another embodiment, the invention provides a method for producing a product of interest comprising culturing a eukaryotic cell which has been transfected with the DNA construct described above, so as to express the product gene and recovering the product.

In a further embodiment, the invention provides a method for producing eukaryotic cells having multiple copies of the product gene comprising transfecting eukaryotic cells with the DNA construct described above (where the selectable gene is an amplifiable gene), growing the cells in a selective medium comprising an amplifying agent for a sufficient time for amplification to occur, and selecting cells having multiple copies of the product gene. Preferably transfection of the cells is achieved using electroporation.

After transfection of the host cells, most of the transfectants fail to exhibit the selectable phenotype characteristic of the protein encoded by the selectable gene, but surprisingly a small proportion of the transfectants do exhibit the selectable phenotype, and among those transfectants, the majority are found to express high levels of the desired product encoded by the product gene. Thus, the invention provides an improved method for the selection of recombinant host cells expressing high levels of a desired product, which method is useful with a wide variety of eukaryotic host cells and avoids the problems inherent in existing cell selection technology.

BRIEF DESCRIPTION OF THE DRAWINGS

Figures 1A-1D illustrate schematically various DNA constructs encompassed by the instant invention. The large arrows represent the selectable gene and the product gene, the V formed by the dashed lines shows the region of the precursor RNA internal to the 5' splice donor site (SD) and 3' splice acceptor site (SA) that is excised from vectors that contain a functional SD. The transcriptional regulatory region, selectable gene, product gene and transcriptional termination site are depicted in Figure 1A. Figure 1B depicts the DNA constructs of Example 1. The various splice donor sequences are depicted, i.e., wild type ras splice donor sequence (WT ras), mutant ras splice donor sequence (MUTANT ras) and non-functional splice donor sequence (Δ GT). The probes used for Northern blot analysis in Example 1 are shown in Figure 1B. Figure 1C depicts the DNA constructs of Example 2 and Figure 1D depicts the DNA construct of Example 3 used for expression of anti-IgE V_H.

Figure 2 depicts schematically the control DNA construct used in Example 1.

Figures 3A-Q depict the nucleotide sequence (SEQ ID NO: 1) of the DHFR/intron- (WT ras SD) -tPA expression vector of Example 1.

Figure 4 is a bar graph which shows the number of colonies that form in selective medium after electroporation of linearized duplicate miniprep DNA's prepared in parallel from the three vectors shown in Figure 1B (i.e. with wild type ras splice donor sequence [WT ras], mutant ras splice donor sequence [MUTANT ras] and non-functional splice donor sequence [Δ GT]) and from the control vector that has DHFR under control of SV40 promoter and tPA under control of CMV promoter (see Figure 2). Cells were selected in nucleoside free medium and counted with an automated colony counter.

Figures 5A-C are bar graphs depicting expression of tPA from stable pools and clones generated from the vectors shown in Figure 1B. In Figure 5A greater than 100 clones from each vector transfection were mixed, plated in 24 well plates, and assayed by tPA ELISA at "saturation". In Figure 5B, twenty clones chosen at random derived from each of the vectors were assayed by tPA ELISA at "saturation". In Figure 5C, the pools mentioned in Figure 5A (except the Δ GT pool) were exposed to 200nM Mtx to select for DHFR amplification and then pooled and assayed for tPA expression.

Figures 6A-P depict the nucleotide sequence (SEQ ID NO: 2) of the DHFR/intron- (WT ras SD) -TNFr-IgG expression vector of Example 2.

Figures 7A-B are bar graphs depicting expression of TNFr-IgG using dicistronic or control vectors (see Example 2). Vectors containing TNFr-IgG (but otherwise identical to those described for tPA expression in Example 1) were constructed (see Figure 1C), introduced into dp12.CHO cells by electroporation, pooled, and assayed for product expression before (Figure 7A) and after (Figure 7B) being subjected to amplification in 200nM Mtx.

Figure 8 depicts schematically the DNA construct used for expression of the V_L of anti-IgE in Example 3.

Figures 9A-O depict the nucleotide sequence (SEQ ID NO: 3) of the anti-IgE V_H expression vector of Example 3.

Figures 10A-Q depict the nucleotide sequence (SEQ ID NO: 4) of the anti-IgE V_L expression vector of Example 3.

5 Figure 11 is a bar graph depicting anti-IgE expression in Example 3. Heavy (V_H) and light (V_L) chain expression vectors were constructed, co-electroporated into CHO cells, clones were selected and assayed for antibody expression. Additionally, pools were established and assessed with regard to expression before and after Mtx selection at 200nM and 1μM.

10 DESCRIPTION OF THE PREFERRED EMBODIMENTS

Definitions:

The "DNA construct" disclosed herein comprises a non-naturally occurring DNA molecule which can either be provided as an isolate or integrated in another DNA molecule e.g. in an expression vector or the
15 chromosome of an eukaryotic host cell.

The term "selectable gene" as used herein refers to a DNA that encodes a selectable marker necessary for the growth or survival of a host cell under the particular cell culture conditions chosen. Accordingly, a host cell that is transformed with a selectable gene will be capable of
20 growth or survival under certain cell culture conditions wherein a non-transfected host cell is not capable of growth or survival. Typically, a selectable gene will confer resistance to a drug or compensate for a metabolic or catabolic defect in the host cell. Examples of selectable genes are provided in the following table. See also Kaufman, Methods in
25 Enzymology, 185: 537-566 (1990), for a review of these.

TABLE 1
Selectable Genes and their Selection Agents

Selection Agent	Selectable Gene
Methotrexate	Dihydrofolate reductase
30 Cadmium	Metallothionein
PALA	CAD
Xyl-A-or adenosine and 2'-deoxycoformycin	Adenosine deaminase
35 Adenine, azaserine, and coformycin	Adenylate deaminase
6-Azaauridine, pyrazofuran	UMP Synthetase
Mycophenolic acid	IMP 5'-dehydrogenase

	Mycophenolic acid with limiting xanthine	Xanthine-guanine phosphoribosyltransferase
	Hypoxanthine, aminopterin, and thymidine (HAT)	Mutant HGPRTase or mutant thymidine kinase
5	5-Fluorodeoxyuridine	Thymidylate synthetase
	Multiple drugs <i>e.g.</i> adriamycin, vincristine or colchicine	P-glycoprotein 170
	Aphidicolin	Ribonucleotide reductase
10	Methionine sulfoximine	Glutamine synthetase
	β -Aspartyl hydroxamate or Albizziin	Asparagine synthetase
	Canavanine	Arginosuccinate synthetase
	α -Difluoromethylornithine	Ornithine decarboxylase
15	Compactin	HMG-CoA reductase
	Tunicamycin	N-Acetylglucosaminyl transferase
	Borrelidin	Threonyl-tRNA synthetase
	Ouabain	Na ⁺ K ⁺ -ATPase

The preferred selectable gene is an amplifiable gene. As used herein, the term "amplifiable gene" refers to a gene which is amplified (*i.e.* additional copies of the gene are generated which survive in intrachromosomal or extrachromosomal form) under certain conditions. The amplifiable gene usually encodes an enzyme (*i.e.* an amplifiable marker) which is required for growth of eukaryotic cells under those conditions. For example, the gene may encode DHFR which is amplified when a host cell transformed therewith is grown in Mtx. According to Kaufman, the selectable genes in Table 1 above can also be considered amplifiable genes. An example of a selectable gene which is generally not considered to be an amplifiable gene is the neomycin resistance gene (Cepko *et al.*, *supra*).

As used herein, "selective medium" refers to nutrient solution used for growing eukaryotic cells which have the selectable gene and therefore includes a "selection agent". Commercially available media such as Ham's F10 (Sigma), Minimal Essential Medium ([MEM], Sigma), RPMI-1640 (Sigma), and Dulbecco's Modified Eagle's Medium ([DMEM], Sigma) are exemplary nutrient solutions. In addition, any of the media described in Ham and Wallace, Meth. Enz., 58:44 (1979), Barnes and Sato, Anal. Biochem., 102:255

(1980), U.S. Patent Nos. 4,767,704; 4,657,866; 4,927,762; or 4,560,655; WO 90/03430; WO 87/00195; U.S. Patent Re. 30,985; or U.S. Patent No. 5,122,469, the disclosures of all of which are incorporated herein by reference, may be used as culture media. Any of these media may be
5 supplemented as necessary with hormones and/or other growth factors (such as insulin, transferrin, or epidermal growth factor), salts (such as sodium chloride, calcium, magnesium, and phosphate), buffers (such as HEPES), nucleosides (such as adenosine and thymidine), antibiotics (such as Gentamycin™ drug), trace elements (defined as inorganic compounds usually
10 present at final concentrations in the micromolar range), and glucose or an equivalent energy source. Any other necessary supplements may also be included at appropriate concentrations that would be known to those skilled in the art. The preferred nutrient solution comprises fetal bovine serum.

The term "selection agent" refers to a substance that interferes with
15 the growth or survival of a host cell that is deficient in a particular selectable gene. Examples of selection agents are presented in Table 1 above. The selection agent preferably comprises an "amplifying agent" which is defined for purposes herein as an agent for amplifying copies of the amplifiable gene, such as Mtx if the amplifiable gene is DHFR. See Table
20 1 for examples of amplifying agents.

As used herein, the term "transcriptional initiation site" refers to the nucleic acid in the DNA construct corresponding to the first nucleic acid incorporated into the primary transcript, i.e., the mRNA precursor, which site is generally provided at, or adjacent to, the 5' end of the DNA
25 construct.

The term "transcriptional termination site" refers to a sequence of DNA, normally represented at the 3' end of the DNA construct, that causes RNA polymerase to terminate transcription.

As used herein, "transcriptional regulatory region" refers to a
30 region of the DNA construct that regulates transcription of the selectable gene and the product gene. The transcriptional regulatory region normally refers to a promoter sequence (i.e. a region of DNA involved in binding of RNA polymerase to initiate transcription) which can be constitutive or inducible and, optionally, an enhancer (i.e. a cis-acting DNA element,
35 usually from about 10-300 bp, that acts on a promoter to increase its transcription).

As used herein, "product gene" refers to DNA that encodes a desired protein or polypeptide product. Any product gene that is capable of expression in a host cell may be used, although the methods of the
40 invention are particularly suited for obtaining high-level expression of a product gene that is not also a selectable or amplifiable gene. Accordingly, the protein or polypeptide encoded by a product gene typically will be one that is not necessary for the growth or survival of a host cell under the particular cell culture conditions chosen. For example, product
45 genes suitably encode a peptide, or may encode a polypeptide sequence of

amino acids for which the chain length is sufficient to produce higher levels of tertiary and/or quaternary structure.

Examples of bacterial polypeptides or proteins include, e.g., alkaline phosphatase and β -lactamase. Examples of mammalian polypeptides or proteins include molecules such as renin; a growth hormone, including human growth hormone, and bovine growth hormone; growth hormone releasing factor; parathyroid hormone; thyroid stimulating hormone; lipoproteins; alpha-1-antitrypsin; insulin A-chain; insulin B-chain; proinsulin; follicle stimulating hormone; calcitonin; luteinizing hormone; glucagon; clotting factors such as factor VIIIC, factor IX, tissue factor, and von Willebrands factor; anti-clotting factors such as Protein C; atrial natriuretic factor; lung surfactant; a plasminogen activator, such as urokinase or human urine or tissue-type plasminogen activator (t-PA); bombesin; thrombin; hemopoietic growth factor; tumor necrosis factor-alpha and -beta; enkephalinase; RANTES (regulated on activation normally T-cell expressed and secreted); human macrophage inflammatory protein (MIP-1-alpha); a serum albumin such as human serum albumin; mullerian-inhibiting substance; relaxin A-chain; relaxin B-chain; prorelaxin; mouse gonadotropin-associated peptide; a microbial protein, such as beta-lactamase; DNase; inhibin; activin; vascular endothelial growth factor (VEGF); receptors for hormones or growth factors; integrin; protein A or D; rheumatoid factors; a neurotrophic factor such as bone-derived neurotrophic factor (BDNF), neurotrophin-3, -4, -5, or -6 (NT-3, NT-4, NT-5, or NT-6), or a nerve growth factor such as NGF- β ; platelet-derived growth factor (PDGF); fibroblast growth factor such as aFGF and bFGF; epidermal growth factor (EGF); transforming growth factor (TGF) such as TGF-alpha and TGF-beta, including TGF- β 1, TGF- β 2, TGF- β 3, TGF- β 4, or TGF- β 5; insulin-like growth factor-I and -II (IGF-I and IGF-II); des(1-3)-IGF-I (brain IGF-I), insulin-like growth factor binding proteins; CD proteins such as CD-3, CD-4, CD-8, and CD-19; erythropoietin; osteoinductive factors; immunotoxins; a bone morphogenetic protein (BMP); an interferon such as interferon-alpha, -beta, and -gamma; colony stimulating factors (CSFs), e.g., M-CSF, GM-CSF, and G-CSF; interleukins (ILs), e.g., IL-1 to IL-10; superoxide dismutase; T-cell receptors; surface membrane proteins; decay accelerating factor; viral antigen such as, for example, a portion of the AIDS envelope; transport proteins; homing receptors; addressins; regulatory proteins; antibodies; chimeric proteins such as immunoadhesins and fragments of any of the above-listed polypeptides.

The product gene preferably does not consist of an anti-sense sequence for inhibiting the expression of a gene present in the host. Preferred proteins herein are therapeutic proteins such as TGF- β , TGF- α , PDGF, EGF, FGF, IGF-I, DNase, plasminogen activators such as t-PA, clotting factors such as tissue factor and factor VIII, hormones such as relaxin and insulin, cytokines such as IFN- γ , chimeric proteins such as TNF receptor IgG immunoadhesin (TNFr-IgG) or antibodies such as anti-IgE.

The term "intron" as used herein refers to a nucleotide sequence present within the transcribed region of a gene or within a messenger RNA precursor, which nucleotide sequence is capable of being excised, or spliced, from the messenger RNA precursor by a host cell prior to translation. Introns suitable for use in the present invention are suitably prepared by any of several methods that are well known in the art, such as purification from a naturally occurring nucleic acid or *de novo* synthesis. The introns present in many naturally occurring eukaryotic genes have been identified and characterized. Mount, Nuc. Acids Res., 10:459 (1982). Artificial introns comprising functional splice sites also have been described. Winey et al., Mol. Cell Biol., 9:329 (1989); Gattermann et al., Mol. Cell Biol., 9:1526 (1989). Introns may be obtained from naturally occurring nucleic acids, for example, by digestion of a naturally occurring nucleic acid with a suitable restriction endonuclease, or by PCR cloning using primers complementary to sequences at the 5' and 3' ends of the intron. Alternatively, introns of defined sequence and length may be prepared synthetically using various methods in organic chemistry. Narang et al., Meth. Enzymol., 68:90 (1979); Caruthers et al., Meth. Enzymol., 154:287 (1985); Froehler et al., Nuc. Acids Res., 14:5399 (1986).

As used herein "splice donor site" or "SD" refers to the DNA sequence immediately surrounding the exon-intron boundary at the 5' end of the intron, where the "exon" comprises the nucleic acid 5' to the intron. Many splice donor sites have been characterized and Ohshima et al., J. Mol. Biol., 195:247-259 (1987) provides a review of these. An "efficient splice donor sequence" refers to a nucleic acid sequence encoding a splice donor site wherein the efficiency of splicing of messenger RNA precursors having the splice donor sequence is between about 80 to 99% and preferably 90 to 95% as determined by quantitative PCR. Examples of efficient splice donor sequences include the wild type (WT) ras splice donor sequence and the GAC:GTAAGT sequence of Example 3. Other efficient splice donor sequences can be readily selected using the techniques for measuring the efficiency of splicing disclosed herein.

The terms "PCR" and "polymerase chain reaction" as used herein refer to the *in vitro* amplification method described in US Patent No. 4,683,195 (issued July 28, 1987). In general, the PCR method involves repeated cycles of primer extension synthesis, using two DNA primers capable of hybridizing preferentially to a template nucleic acid comprising the nucleotide sequence to be amplified. The PCR method can be used to clone specific DNA sequences from total genomic DNA, cDNA transcribed from cellular RNA, viral or plasmid DNAs. Wang & Mark, in PCR Protocols, pp. 70-75 (Academic Press, 1990); Scharf, in PCR Protocols, pp. 84-98; Kawasaki & Wang, in PCR Technology, pp. 89-97 (Stockton Press, 1989). Reverse transcription-polymerase chain reaction (RT-PCR) can be used to analyze RNA samples containing mixtures of spliced and unspliced mRNA transcripts. Fluorescently tagged primers designed to span the intron are used to

amplify both spliced and unspliced targets. The resultant amplification products are then separated by gel electrophoresis and quantitated by measuring the fluorescent emission of the appropriate band(s). A comparison is made to determine the amount of spliced and unspliced transcripts present in the RNA sample.

One preferred splice donor sequence is a "consensus splice donor sequence". The nucleotide sequences surrounding intron splice sites, which sequences are evolutionarily highly conserved, are referred to as "consensus splice donor sequences". In the mRNAs of higher eukaryotes, the 5' splice site occurs within the consensus sequence AG:GUAAGU (wherein the colon denotes the site of cleavage and ligation). In the mRNAs of yeast, the 5' splice site is bounded by the consensus sequence :GUAUGU. Padgett, et al., Ann. Rev. Biochem., 55:1119 (1986).

The expression "splice acceptor site" or "SA" refers to the sequence immediately surrounding the intron-exon boundary at the 3' end of the intron, where the "exon" comprises the nucleic acid 3' to the intron. Many splice acceptor sites have been characterized and Ohshima et al., J. Mol. Biol., 195:247-259 (1987) provides a review of these. The preferred splice acceptor site is an efficient splice acceptor site which refers to a nucleic acid sequence encoding a splice acceptor site wherein the efficiency of splicing of messenger RNA precursors having the splice acceptor site is between about 80 to 99% and preferably 90 to 95% as determined by quantitative PCR. The splice acceptor site may comprise a consensus sequence. In the mRNAs of higher eukaryotes, the 3' splice acceptor site occurs within the consensus sequence (U/C)₁₁NCAG:G. In the mRNAs of yeast, the 3' acceptor splice site is bounded by the consensus sequence (C/U)AG:. Padgett, et al., *supra*.

As used herein "culturing for sufficient time to allow amplification to occur" refers to the act of physically culturing the eukaryotic host cells which have been transformed with the DNA construct in cell culture media containing the amplifying agent, until the copy number of the amplifiable gene (and preferably also the copy number of the product gene) in the host cells has increased relative to the transformed cells prior to this culturing.

The term "expression" as used herein refers to transcription or translation occurring within a host cell. The level of expression of a product gene in a host cell may be determined on the basis of either the amount of corresponding mRNA that is present in the cell or the amount of the protein encoded by the product gene that is produced by the cell. For example, mRNA transcribed from a product gene is desirably quantitated by northern hybridization. Sambrook, et al., Molecular Cloning: A Laboratory Manual, pp. 7.3-7.57 (Cold Spring Harbor Laboratory Press, 1989). Protein encoded by a product gene can be quantitated either by assaying for the biological activity of the protein or by employing assays that are independent of such activity, such as western blotting or radioimmunoassay using antibodies that are capable of reacting with the protein. Sambrook,

et al., Molecular Cloning: A Laboratory Manual, pp. 18.1-18.88 (Cold Spring Harbor Laboratory Press, 1989).

Modes for Carrying Out the Invention

Methods and compositions are provided for enhancing the stability and/or copy number of a transcribed sequence in order to allow for elevated levels of a RNA sequence of interest. In general, the methods of the present invention involve transfecting a eukaryotic host cell with an expression vector comprising both a product gene encoding a desired polypeptide and a selectable gene (preferably an amplifiable gene).

Selectable genes and product genes may be obtained from genomic DNA, cDNA transcribed from cellular RNA, or by *in vitro* synthesis. For example, libraries are screened with probes (such as antibodies or oligonucleotides of about 20-80 bases) designed to identify the selectable gene or the product gene (or the protein(s) encoded thereby). Screening the cDNA or genomic library with the selected probe may be conducted using standard procedures as described in chapters 10-12 of Sambrook et al., Molecular Cloning: A Laboratory Manual (New York: Cold Spring Harbor Laboratory Press, 1989). An alternative means to isolate the selectable gene or product gene is to use PCR methodology as described in section 14 of Sambrook et al., *supra*.

A preferred method of practicing this invention is to use carefully selected oligonucleotide sequences to screen cDNA libraries from various tissues known to contain the selectable gene or product gene. The oligonucleotide sequences selected as probes should be of sufficient length and sufficiently unambiguous that false positives are minimized.

The oligonucleotide generally is labeled such that it can be detected upon hybridization to DNA in the library being screened. The preferred method of labeling is to use ³²P- labeled ATP with polynucleotide kinase, as is well known in the art, to radiolabel the oligonucleotide. However, other methods may be used to label the oligonucleotide, including, but not limited to, biotinylation or enzyme labeling.

Sometimes, the DNA encoding the selectable gene and product gene is preceded by DNA encoding a signal sequence having a specific cleavage site at the N-terminus of the mature protein or polypeptide. In general, the signal sequence may be a component of the expression vector, or it may be a part of the selectable gene or product gene that is inserted into the expression vector. If a heterologous signal sequence is used, it preferably is one that is recognized and processed (*i.e.*, cleaved by a signal peptidase) by the host cell. For yeast secretion the native signal sequence may be substituted by, *e.g.*, the yeast invertase leader, alpha factor leader (including *Saccharomyces* and *Kluyveromyces* α -factor leaders, the latter described in U.S. Pat. No. 5,010,182 issued 23 April 1991), or acid phosphatase leader, the *C. albicans* glucoamylase leader (EP 362,179 published 4 April 1990), or the signal described in WO 90/13646 published 15 November 1990. In mammalian cell expression the native signal sequence

of the protein of interest is satisfactory, although other mammalian signal sequences may be suitable, such as signal sequences from secreted polypeptides of the same or related species, as well as viral secretory leaders, for example, the herpes simplex gD signal. The DNA for such precursor region is ligated in reading frame to the selectable gene or product gene.

As shown in Figure 1A, the selectable gene is generally provided at the 5' end of the DNA construct and this selectable gene is followed by the product gene. Therefore, the full length (non-spliced) message will contain DHFR as the first open reading frame and will therefore generate DHFR protein to allow selection of stable transfectants. The full length message is not expected to generate appreciable amounts of the protein of interest as the second AUG in a dicistronic message is an inefficient initiator of translation in mammalian cells (Kozak, J. Cell Biol., 115: 887-903 [1991]).

The selectable gene is positioned within an intron. Introns are noncoding nucleotide sequences, normally present within many eukaryotic genes, which are removed from newly transcribed mRNA precursors in a multiple-step process collectively referred to as splicing.

A single mechanism is thought to be responsible for the splicing of mRNA precursors in mammalian, plant, and yeast cells. In general, the process of splicing requires that the 5' and 3' ends of the intron be correctly cleaved and the resulting ends of the mRNA be accurately joined, such that a mature mRNA having the proper reading frame for protein synthesis is produced. Analysis of a variety of naturally occurring and synthetically constructed mutant genes has shown that nucleotide changes at many of the positions within the consensus sequences at the 5' and 3' splice sites have the effect of reducing or abolishing the synthesis of mature mRNA. Sharp, Science, 235:766 (1987); Padgett, et al., Ann. Rev. Biochem., 55:1119 (1986); Green, Ann. Rev. Genet., 20:671 (1986). Mutational studies also have shown that RNA secondary structures involving splicing sites can affect the efficiency of splicing. Solnick, Cell, 43:667 (1985); Konarska, et al., Cell, 42:165 (1985).

The length of the intron may also affect the efficiency of splicing. By making deletion mutations of different sizes within the large intron of the rabbit beta-globin gene, Wieringa, et al. determined that the minimum intron length necessary for correct splicing is about 69 nucleotides. Cell, 37:915 (1984). Similar studies of the intron of the adenovirus E1A region have shown that an intron length of about 78 nucleotides allows correct splicing to occur, but at reduced efficiency. Increasing the length of the intron to 91 nucleotides restores normal splicing efficiency, whereas truncating the intron to 63 nucleotides abolishes correct splicing. Ulfendahl, et al., Nuc. Acids Res., 13:6299 (1985).

To be useful in the invention, the intron must have a length such that splicing of the intron from the mRNA is efficient. The preparation of introns of differing lengths is a routine matter, involving methods well known in the art, such as *de novo* synthesis or *in vitro* deletion

mutagenesis of an existing intron. Typically, the intron will have a length of at least about 150 nucleotides, since introns which are shorter than this tend to be spliced less efficiently. The upper limit for the length of the intron can be up to 30 kB or more. However, as a general
5 proposition, the intron is generally less than about 10 kB in length.

The intron is modified to contain the selectable gene not normally present within the intron using any of the various known methods for modifying a nucleic acid *in vitro*. Typically, a selectable gene will be introduced into an intron by first cleaving the intron with a restriction
10 endonuclease, and then covalently joining the resulting restriction fragments to the selectable gene in the correct orientation for host cell expression, for example by ligation with a DNA ligase enzyme.

The DNA construct is dicistronic, i.e. the selectable gene and product gene are both under the transcriptional control of a single
15 transcriptional regulatory region. As mentioned above, the transcriptional regulatory region comprises a promoter. Suitable promoting sequences for use with yeast hosts include the promoters for 3-phosphoglycerate kinase (Hitzeman et al., J. Biol. Chem., 255:2073 [1980]) or other glycolytic enzymes (Hess et al., J. Adv. Enzyme Req., 7:149 [1968]; and Holland,
20 Biochemistry, 17:4900 [1978]), such as enolase, glyceraldehyde-3-phosphate dehydrogenase, hexokinase, pyruvate decarboxylase, phosphofructokinase, glucose-6-phosphate isomerase, 3-phosphoglycerate mutase, pyruvate kinase, triosephosphate isomerase, phosphoglucose isomerase, and glucokinase.

Other yeast promoters, which are inducible promoters having the
25 additional advantage of transcription controlled by growth conditions, are the promoter regions for alcohol dehydrogenase 2, isocytochrome C, acid phosphatase, degradative enzymes associated with nitrogen metabolism, metallothionein, glyceraldehyde-3-phosphate dehydrogenase, and enzymes responsible for maltose and galactose utilization. Suitable vectors and
30 promoters for use in yeast expression are further described in Hitzeman et al., EP 73,657A. Yeast enhancers also are advantageously used with yeast promoters.

Expression control sequences are known for eukaryotes. Virtually all eukaryotic genes have an AT-rich region located approximately 25 to 30
35 bases upstream from the site where transcription is initiated. Another sequence found 70 to 80 bases upstream from the start of transcription of many genes is a CXCAAT region where X may be any nucleotide.

Product gene transcription from vectors in mammalian host cells is controlled by promoters obtained from the genomes of viruses such as
40 polyoma virus, fowlpox virus (UK 2,211,504 published 5 July 1989), adenovirus (such as Adenovirus 2), bovine papilloma virus, avian sarcoma virus, cytomegalovirus, a retrovirus, hepatitis-B virus and most preferably Simian Virus 40 (SV40), from heterologous mammalian promoters, e.g. the actin promoter or an immunoglobulin promoter, from heat-shock promoters,
45 and from the promoter normally associated with the product gene, provided such promoters are compatible with the host cell systems.

The early and late promoters of the SV40 virus are conveniently obtained as an SV40 restriction fragment that also contains the SV40 viral origin of replication. Fiers et al., Nature, 273:113 (1978); Mulligan and Berg, Science, 209:1422-1427 (1980); Pavlakis et al., Proc. Natl. Acad. Sci. USA, 78:7398-7402 (1981). The immediate early promoter of the human cytomegalovirus (CMV) is conveniently obtained as a HindIII E restriction fragment. Greenaway et al., Gene, 18:355-360 (1982). A system for expressing DNA in mammalian hosts using the bovine papilloma virus as a vector is disclosed in U.S. 4,419,446. A modification of this system is described in U.S. 4,601,978. See also Gray et al., Nature, 295:503-508 (1982) on expressing cDNA encoding immune interferon in monkey cells; , Reyes et al., Nature, 297:598-601 (1982) on expression of human β -interferon cDNA in mouse cells under the control of a thymidine kinase promoter from herpes simplex virus, Canaani and Berg, Proc. Natl. Acad. Sci. USA, 79:5166-5170 (1982) on expression of the human interferon β 1 gene in cultured mouse and rabbit cells, and Gorman et al., Proc. Natl. Acad. Sci. USA, 79:6777-6781 (1982) on expression of bacterial CAT sequences in CV-1 monkey kidney cells, chicken embryo fibroblasts, Chinese hamster ovary cells, HeLa cells, and mouse NIH-3T3 cells using the Rous sarcoma virus long terminal repeat as a promoter.

Preferably the transcriptional regulatory region in higher eukaryotes comprises an enhancer sequence. Enhancers are relatively orientation and position independent having been found 5' (Lainins et al., Proc. Natl. Acad. Sci. USA, 78:993 [1981]) and 3' (Lusky et al., Mol. Cell Bio., 3:1108 [1983]) to the transcription unit, within an intron (Banerji et al., Cell, 33:729 [1983]) as well as within the coding sequence itself (Osborne et al., Mol. Cell Bio., 4:1293 [1984]). Many enhancer sequences are now known from mammalian genes (globin, elastase, albumin, α -fetoprotein and insulin). Typically, however, one will use an enhancer from a eukaryotic cell virus. Examples include the SV40 enhancer on the late side of the replication origin (bp 100-270), the cytomegalovirus early promoter enhancer (CMV), the polyoma enhancer on the late side of the replication origin, and adenovirus enhancers. See also Yaniv, Nature, 297:17-18 (1982) on enhancing elements for activation of eukaryotic promoters. The enhancer may be spliced into the vector at a position 5' or 3' to the product gene, but is preferably located at a site 5' from the promoter.

The DNA construct has a transcriptional initiation site following the transcriptional regulatory region and a transcriptional termination region following the product gene (see Figure 1A). These sequences are provided in the DNA construct using techniques which are well known in the art.

The DNA construct normally forms part of an expression vector which may have other components such as an origin of replication (i.e., a nucleic acid sequence that enables the vector to replicate in one or more selected host cells) and, if desired, one or more additional selectable gene(s). Construction of suitable vectors containing the desired coding and control sequences employs standard ligation techniques. Isolated plasmids or DNA

fragments are cleaved, tailored, and religated in the form desired to generate the plasmids required.

Generally, in cloning vectors the origin of replication is one that enables the vector to replicate independently of the host chromosomal DNA, and includes origins of replication or autonomously replicating sequences. Such sequences are well known. The 2 μ plasmid origin of replication is suitable for yeast, and various viral origins (SV40, polyoma, adenovirus, VSV or BPV) are useful for cloning vectors in mammalian cells. Generally, the origin of replication component is not needed for mammalian expression vectors (the SV40 origin may typically be used only because it contains the early promoter).

Most expression vectors are "shuttle" vectors, i.e., they are capable of replication in at least one class of organisms but can be transfected into another organism for expression. For example, a vector is cloned in *E. coli* and then the same vector is transfected into yeast or mammalian cells for expression even though it is not capable of replicating independently of the host cell chromosome.

For analysis to confirm correct sequences in plasmids constructed, plasmids from the transformants are prepared, analyzed by restriction, and/or sequenced by the method of Messing et al., Nucleic Acids Res., 9:309 (1981) or by the method of Maxam et al., Methods in Enzymology, 65:499 (1980).

The expression vector having the DNA construct prepared as discussed above is transformed into a eukaryotic host cell. Suitable host cells for cloning or expressing the vectors herein are yeast or higher eukaryote cells.

Eukaryotic microbes such as filamentous fungi or yeast are suitable hosts for vectors containing the product gene. *Saccharomyces cerevisiae*, or common baker's yeast, is the most commonly used among lower eukaryotic host microorganisms. However, a number of other genera, species, and strains are commonly available and useful herein, such as *S. pombe* [Beach and Nurse, Nature, 290:140 (1981)], *Kluyveromyces lactis* [Louvencourt et al., J. Bacteriol., 737 (1983)], *Yarrowia* [EP 402,226], *Pichia pastoris* [EP 183,070], *Trichoderma reesia* [EP 244,234], *Neurospora crassa* [Case et al., Proc. Natl. Acad. Sci. USA, 76:5259-5263 (1979)], and *Aspergillus* hosts such as *A. nidulans* [Ballance et al., Biochem. Biophys. Res. Commun., 112:284-289 (1983); Tilburn et al., Gene, 26:205-221 (1983); Yelton et al., Proc. Natl. Acad. Sci. USA, 81:1470-1474 (1984)] and *A. niger* [Kelly and Hynes, EMBO J., 4:475-479 (1985)].

Suitable host cells for the expression of the product gene are derived from multicellular organisms. Such host cells are capable of complex processing and glycosylation activities. In principle, any higher eukaryotic cell culture is workable, whether from vertebrate or invertebrate culture. Examples of invertebrate cells include plant and insect cells. Numerous baculoviral strains and variants and corresponding permissive insect host cells from hosts such as *Spodoptera frugiperda*

(caterpillar), *Aedes aegypti* (mosquito), *Aedes albopictus* (mosquito), *Drosophila melanogaster* (fruitfly), and *Bombyx mori* host cells have been identified. See, e.g., Luckow et al., Bio/Technology, 6:47-55 (1988); Miller et al., in Genetic Engineering, Setlow, J.K. et al., eds., Vol. 8
5 (Plenum Publishing, 1986), pp. 277-279; and Maeda et al., Nature, 315:592-594 (1985). A variety of such viral strains are publicly available, e.g., the L-1 variant of *Autographa californica* NPV and the Bm-5 strain of *Bombyx mori* NPV, and such viruses may be used as the virus herein according to the present invention, particularly for transfection of *Spodoptera frugiperda*
10 cells.

Plant cell cultures of cotton, corn, potato, soybean, petunia, tomato, and tobacco can be utilized as hosts. Typically, plant cells are transfected by incubation with certain strains of the bacterium *Agrobacterium tumefaciens*, which has been previously manipulated to contain
15 the product gene. During incubation of the plant cell culture with *A. tumefaciens*, the product gene is transferred to the plant cell host such that it is transfected, and will, under appropriate conditions, express the product gene. In addition, regulatory and signal sequences compatible with plant cells are available, such as the nopaline synthase promoter and polyadenylation signal sequences. Depicker et al., J. Mol. Appl. Gen.,
20 1:561 (1982). In addition, DNA segments isolated from the upstream region of the T-DNA 780 gene are capable of activating or increasing transcription levels of plant-expressible genes in recombinant DNA-containing plant tissue. EP 321,196 published 21 June 1989.

25 However, interest has been greatest in vertebrate cells, and propagation of vertebrate cells in culture (tissue culture) has become a routine procedure in recent years [Tissue Culture, Academic Press, Kruse and Patterson, editors (1973)]. Examples of useful mammalian host cell lines are monkey kidney CV1 line transformed by SV40 (COS-7, ATCC CRL
30 1651); human embryonic kidney line (293 or 293 cells subcloned for growth in suspension culture, Graham et al., J. Gen Virol., 36:59 [1977]); baby hamster kidney cells (BHK, ATCC CCL 10); Chinese hamster ovary cells/-DHFR (CHO, Urlaub and Chasin, Proc. Natl. Acad. Sci. USA, 77:4216 [1980]); dp12.CHO cells (EP 307,247 published 15 March 1989); mouse sertoli cells
35 (TM4, Mather, Biol. Reprod., 23:243-251 [1980]); monkey kidney cells (CV1 ATCC CCL 70); African green monkey kidney cells (VERO-76, ATCC CRL-1587); human cervical carcinoma cells (HELA, ATCC CCL 2); canine kidney cells (MDCK, ATCC CCL 34); buffalo rat liver cells (BRL 3A, ATCC CRL 1442); human lung cells (W138, ATCC CCL 75); human liver cells (Hep G2, HB 8065); mouse
40 mammary tumor (MMT 060562, ATCC CCL51); TRI cells (Mather et al., Annals N.Y. Acad. Sci., 383:44-68 [1982]); MRC 5 cells; FS4 cells; and a human hepatoma line (Hep G2).

Host cells are transformed with the above-described expression or cloning vectors of this invention and cultured in conventional nutrient
45 media modified as appropriate for inducing promoters, selecting transformants, or amplifying the genes encoding the desired sequences.

Infection with *Agrobacterium tumefaciens* is used for transformation of certain plant cells, as described by Shaw et al., Gene, 23:315 (1983) and WO 89/05859 published 29 June 1989. For mammalian cells without such cell walls, the calcium phosphate precipitation method of Graham and van der Eb, Virology, 52:456-457 (1978) may be used. General aspects of mammalian cell host system transformations have been described by Axel in U.S. 4,399,216 issued 16 August 1983. Transformations into yeast are typically carried out according to the method of Van Solingen et al., J. Bact., 130:946 (1977) and Hsiao et al., Proc. Natl. Acad. Sci. (USA), 76:3829 (1979). However, other methods for introducing DNA into cells such as by nuclear injection or by protoplast fusion may also be used.

In the preferred embodiment the DNA is introduced into the host cells using electroporation. See Andreason, J. Tiss. Cult. Meth., 15:56-62 (1993), for a review of electroporation techniques useful for practicing the instantly claimed invention. It was discovered that electroporation techniques for introducing the DNA construct into the host cells were preferable over calcium phosphate precipitation techniques insofar as the latter could cause the DNA to break up and forming concatemers.

The mammalian host cells used to express the product gene herein may be cultured in a variety of media as discussed in the definitions section above. The media contains the selection agent used for selecting transformed host cells which have taken up the DNA construct (either as an intra- or extra-chromosomal element). To achieve selection of the transformed eukaryotic cells, the host cells may be grown in cell culture plates and individual colonies expressing the selectable gene (and thus the product gene) can be isolated and grown in growth medium until the nutrients are depleted. The host cells are then analyzed for transcription and/or transformation as discussed below. The culture conditions, such as temperature, pH, and the like, are those previously used with the host cell selected for expression, and will be apparent to the ordinarily skilled artisan.

Gene amplification and/or expression may be measured in a sample directly, for example, by conventional Southern blotting, Northern blotting to quantitate the transcription of mRNA (Thomas, Proc. Natl. Acad. Sci. USA, 77:5201-5205 [1980]), dot blotting (DNA analysis), or in situ hybridization, using an appropriately labeled probe, based on the sequences provided herein. Various labels may be employed, most commonly radioisotopes, particularly ³²P. However, other techniques may also be employed, such as using biotin-modified nucleotides for introduction into a polynucleotide. The biotin then serves as the site for binding to avidin or antibodies, which may be labeled with a wide variety of labels, such as radionuclides, fluorescents, enzymes, or the like. Alternatively, antibodies may be employed that can recognize specific duplexes, including DNA duplexes, RNA duplexes, and DNA-RNA hybrid duplexes or DNA-protein duplexes. The antibodies in turn may be labeled and the assay may be carried out where the duplex is bound to a surface, so that upon the

formation of duplex on the surface, the presence of antibody bound to the duplex can be detected.

Gene expression, alternatively, may be measured by immunological methods, such as immunohistochemical staining of tissue sections and assay
5 of cell culture or body fluids, to quantitate directly the expression of gene product. With immunohistochemical staining techniques, a cell sample is prepared, typically by dehydration and fixation, followed by reaction with labeled antibodies specific for the gene product coupled, where the labels are usually visually detectable, such as enzymatic labels,
10 fluorescent labels, luminescent labels, and the like. A particularly sensitive staining technique suitable for use in the present invention is described by Hsu et al., Am. J. Clin. Path., 75:734-738 (1980).

In the preferred embodiment, the mRNA is analyzed by quantitative PCR (to determine the efficiency of splicing) and protein expression is
15 measured using ELISA as described in Example 1 herein.

The product of interest preferably is recovered from the culture medium as a secreted polypeptide, although it also may be recovered from host cell lysates when directly expressed without a secretory signal. When the product gene is expressed in a recombinant cell other than one of human
20 origin, the product of interest is completely free of proteins or polypeptides of human origin. However, it is necessary to purify the product of interest from recombinant cell proteins or polypeptides to obtain preparations that are substantially homogeneous as to the product of interest. As a first step, the culture medium or lysate is centrifuged
25 to remove particulate cell debris. The product of interest thereafter is purified from contaminant soluble proteins and polypeptides, for example, by fractionation on immunoaffinity or ion-exchange columns; ethanol precipitation; reverse phase HPLC; chromatography on silica or on a cation exchange resin such as DEAE; chromatofocusing; SDS-PAGE; ammonium sulfate
30 precipitation; gel electrophoresis using, for example, Sephadex G-75; chromatography on plasminogen columns to bind the product of interest and protein A Sepharose columns to remove contaminants such as IgG.

The following examples are offered by way of illustration only and are not intended to limit the invention in any manner. All patent and
35 literature references cited herein are expressly incorporated by reference.

EXAMPLE 1

tPA production using the dicistronic expression vectors

It was sought to increase the level of homogeneity with regard to expression levels of stable clones by expressing a selectable marker (such
40 as DHFR) and the protein of interest from a single promoter. These vectors divert most of the transcript to product expression while linking it at a fixed ratio to DHFR expression via differential splicing.

Vectors were constructed which were derived from the vector pRK (Suva et al., Science, 237:893-896 [1987]) which contains an intron between the
45 cytomegalovirus immediate early promoter (CMV) and the cDNA that encodes

the polypeptide of interest. The intron of pRK is 139 nucleotides in length, has a splice donor site derived from cytomegalovirus immediate early gene (CMVIE), and a splice acceptor site from an IgG heavy chain variable region (V_H) gene (Eaton et al., Biochem., 25:8343 [1986]).

5 DHFR/intron vectors were constructed by inserting an EcoRV linker into the BSTX1 site present in the intron of pRK7. An 830 base-pair fragment containing a mouse DHFR coding fragment was inserted to obtain DHFR intron expression vectors which differ only in the sequence that
10 comprises the splice donor site. Those sequences were altered by overlapping PCR mutagenesis to obtain sequences that match splice donor sites found between exons 3 and 4 of normal and mutant Ras genes. PCR was also used to destroy the splice donor site.

A mouse DHFR cDNA fragment (Simonsen et al., Proc. Natl. Acad. Sci. USA, 80:2495-2499 [1983]) was inserted into the intron of this vector 59
15 nucleotides downstream of the splice donor site. The splice donor site of this vector was altered by mutagenesis to change the ratio of spliced to non-spliced message in transfected cells. It has previously been shown that a single nucleotide change (G to A) converted a relatively efficient splice donor site found in the normal ras gene into an inefficient splice
20 site (Cohen et al., Nature, 334:119-124 [1988]). This effect has been demonstrated in the context of the ras gene and confirmed when these sequences were transferred to human growth hormone constructs (Cohen et al., Cell, 58:461-472 [1989]). Additionally, a non functional 5' splice site (GT to CA) was constructed as a control (Δ GT). A polylinker was
25 inserted 35 nucleotides downstream of the 3' splice site to accept the cDNA of interest. A vector containing tPA (Pennica et al., Nature, 301:214-221 [1983]) was linearized downstream of the polyadenylation site before it was introduced into CHO cells (Potter et al., Proc. Natl. Acad. Sci. USA, 81:7161 [1984]).

30 Plasmid DNA's that contained DHFR/intron, tPA and (a) wild type ras (WT ras), i.e. Figure 3 (SEQ ID NO: 1), (b) mutant ras, or (c) non-functional splice donor site (Δ GT) were introduced into CHO DHFR minus cells by electroporation. The intron vectors were each linearized downstream of the polyadenylation site by restriction endonuclease
35 treatment. The control vector was linearized downstream of the second polyadenylation site. The DNA's were ethanol precipitated after phenol/chloroform extraction and were resuspended in 20 μ l 1/10 Tris EDTA. Then, 10 μ g of DNA was incubated with 10⁷ CHO.dpl2 cells (EP 307,247 published 15 March 1989) in 1 ml of PBS on ice for 10 min. before
40 electroporation at 400 volts and 330 μ f using a BRL Cell Porator.

Cells were returned to ice for 10 min. before being plated into non-selective medium. After 24 hours cells were fed nucleoside-free medium to select for stable DHFR+ clones which were pooled. The pooled DHFR+ clones were lysed and mRNA's were prepared.

45 To prepare the mRNA, RNA was extracted from 5 x 10⁷ cells which were grown from pools of more than 200 clones derived from the stable

transfection of the three vectors, the essential construction of which is shown in Figure 1B and from non-transfected CHO cells. RNA was purified over oligo-DT cellulase (Collaborative Biomedical Products). 10µg of mRNA was then subjected to Northern blotting which involved running the mRNA on
5 a 1.2% agarose, 6.6% formaldehyde gel, and transferring it to a nylon filter (Stratagene Duralon-UV membrane), prehybridized, probed and washed according to the manufacturer's instructions.

The filter was probed sequentially using probes (shown in Figure 1B) that would detect (a) the full length message, (b) both full length and
10 spliced message, or (c) beta actin. Probing with the long probe showed that the vector that contains the efficient splice donor site (i.e. WT ras) generates predominately a mRNA of the size predicted for the spliced product while the other two vectors gave rise primarily to a mRNA that corresponds in size to non-spliced message. The DHFR probe detected only
15 full length message and demonstrated that the WT ras splice donor derived vector generates very little full length message with which to confer a DHFR positive phenotype.

Figure 4 shows the number of DHFR positive colonies obtained after duplicate electroporations with the three intron vectors described above and from a conventional vector that has a CMV promoter driving tPA and a
20 SV40 promoter driving DHFR (see Figure 2). The increase in colony number parallels the increase in full length message that accumulates with the modification of the splice donor sites. The conventional vector efficiently generates colonies and does not vary significantly from the ΔGT
25 construct.

The level of tPA expression was determined by seeding cells in 1 ml of F12:DMEM (50:50, with 5% FBS) in 24 well dishes to near confluency. Growth of the cells continued until the media was exhausted. Media was then assayed by ELISA for tPA production. Briefly, anti-tPA antibody was
30 coated onto the wells of an ELISA microtiter plate, media samples were added to the wells followed by washing. Binding of the antigen (tPA) was then quantified using horse radish peroxidase (HRPO) labelled anti-tPA antibody.

Figure 5A depicts the titers of secreted tPA protein after pooling
35 the clones of each group shown in Figure 4. While the number of colonies increased with a weakening of splice donor function, the inverse was seen with respect to tPA expression. The expression levels are consistent with the RNA products that are observed; as more of the dicistronic message is spliced an increased amount of message will contain tPA as the first open
40 reading frame resulting in increased tPA expression. A mutation of GT to CA in the splice donor site results in an abundance of DHFR positive colonies which express undetectable levels of tPA, possibly resulting from inefficient utilization of the second AUG. Importantly, Figure 5A also shows that expression levels obtained from one of the dicistronic vectors
45 (with WT ras SD) was about threefold higher than that obtained with the control vector containing a CMV promoter/enhancer driving tPA, SV40

promoter/enhancer controlling DHFR and SV40 polyadenylation signals controlling the expression of tPA and DHFR.

Additionally, the homogeneity of expression in the pools was investigated. Figure 5B shows that all 20 clones generated by the WT ras splice donor site derived dicistronic vectors express detectable levels of tPA while only 4 of 20 clones generated by the control vector express tPA. None of the clones transfected with the non-splicing (Δ GT) vector expressed tPA levels detectable by ELISA. This finding is consistent with previous observations that relatively few clones generated by conventional vectors make useful levels of protein.

Expression of tPA was increased following methotrexate amplification of pools. Figure 5C shows that 2 of the dicistronic vector derived pools (i.e. with WT ras and MUTANT ras SD sites) increased in expression markedly (8.4 and 7.7 fold), while the pool generated by the conventional vector increased only slightly (2.8 fold) when each was subjected to 200 nM Mtx. An overall increase of 9 fold was obtained using the best dicistronic (WT ras SD) versus the conventional vector following amplification. Growth of the highest expressing amplified pool in nutrient rich production medium yielded titers of 4.2 μ g/ml tPA.

It was shown that manipulation of the splice donor sequence alters the ratio of spliced to full length message and the number of colonies that form in selective medium. It was also shown that dicistronic expression vectors generate clones that express high levels of recombinant proteins. Surprisingly, it was possible to isolate high expressors which had the efficient WT ras splice donor site by selection for DHFR^r cells despite the efficiency with which the DHFR gene was spliced from the RNA precursors formed in these cells.

EXAMPLE 2

TNFr-IgG production using the dicistronic expression vectors

To prove the general applicability of this approach, a second product was evaluated in the dicistronic vector system containing, as the DNA of interest, an immunoadhesin (TNFr-IgG) capable of binding tumor necrosis factor (TNF) (Ashkenazi et al., Proc. Natl. Acad. Sci. USA, 88:10535-10539 [1991]). The experiments described in Example 1 above were essentially repeated except that the product gene encoded the immunoadhesin TNFr-IgG. Plasmid DNA's that contained a TNFr-IgG cDNA and (a) WT ras, i.e. Figure 6 (SEQ ID NO: 2), (b) mutant ras or (c) nonfunctional splice donor site (Δ GT) were introduced into the dp12.CHO cells as discussed for Example 1. See Figure 1C for an illustration of the DNA constructs.

It was discovered that the number of DHFR positive colonies generated by three of these vectors was similar to that seen with the tPA constructs. Expression of TNFr-IgG also paralleled that seen with the tPA constructs (Figure 7A). Amplification of pools from two of the constructs showed a marked increase in expression of immunoadhesin (9.6 and 6.8 fold) (Figure

7B). The best of these amplified pools expressed 9.5 $\mu\text{g/ml}$ when grown in nutrient rich production medium.

Thus, it was again shown that dicistronic expression vectors generate clones that express high levels of recombinant proteins. Furthermore, contrary to expectations, it was discovered that isolation of high product expressing host DHFR⁺ cells was possible using an efficient splice donor site (i.e. the WT ras splice donor site).

EXAMPLE 3

Antibody production using a dicistronic expression vector

The usefulness of this system for antibody expression was evaluated by testing production of an antibody directed against IgE (Presta et al., Journal of Immunology, 151:2623-2632 [1993]). Further, the flexibility of the system with regard to transcription initiation was tested by replacing the CMV promoter/enhancer present in the previous vectors with the promoter/ enhancer derived from the early region of SV40 virus (Griffin, B., Structure and Genomic Organization of SV40 and Polyoma Virus, In J. Tooze [Ed] DNA Tumor Viruses, Cold Spring Harbor Laboratory, Cold Spring Harbor, New York). The heavy chain of the antibody was inserted downstream of DHFR as described in the earlier tPA and TNF α -IgG constructs. Additionally, a new splice donor site sequence (GAC:GTAAGT) was engineered into the vector which matches the consensus splice donor site more closely than did the splice donor sites present in the vectors tested in Examples 1 and 2. The resultant expression vector is shown in Figures 1D and 9.

It was discovered that this vector produced fewer colonies than the vectors previously tested, and produced predominantly a spliced RNA product. A second vector was constructed to have the light chain of the antibody under control of the SV40 promoter/enhancer and poly-A and the hygromycin B resistance gene under control of the CMV promoter/enhancer and SV40 poly-A. These vectors were linearized at unique HpaI sites downstream of the poly-A signal, mixed at a ratio of light chain vector to heavy chain vector of 10:3 and electroporated into CHO cells using an optimized protocol (as discussed in Examples 1 and 2).

Figure 11 shows the levels of antibody expressed by clones and pools after selection in hygromycin B followed by selection for DHFR expression. All 20 of the clones analyzed expressed high levels of antibody when grown in rich medium and varied from one another by only a factor of four. A pool of antibody producing clones was generated and assayed shortly after it was established. That pool was grown continuously for 6 weeks without a significant decrease in productivity demonstrating that its stability was sufficient to generate gram quantities of protein from its large scale culture.

The pool was subjected to methotrexate amplification at 200nM and 1 μM and achieved a greater than 2 fold increase in antibody titer. The 1 μM Mtx resistant pool achieved a titer of 41 mg/L when grown under optimal conditions in suspension culture.

The structure of the expressed antibody was examined. Proteins expressed by the 200nM methotrexate resistant pool and by a well characterized expression clone generated by conventional vectors (Presta et al. [1993], supra) were metabolically labeled with S^{35} cysteine and methionine. In particular, confluent 35mm plates of cells were metabolically labeled with 50 μ Ci each S-35 methionine and S-35 cysteine (Amersham) in serum free cysteine and methionine free F12:DMEM. After one hour, nutrient rich production media was added and labeled proteins were allowed to "chase" into the medium for six more hours. Proteins were run on a 12% SDS/PAGE gel (NOVEX) non-reduced or following reduction with B-mercaptoethanol. Dried gels were exposed to film for 16 hours. CHO control cells were also labeled.

The majority of the antibody protein is secreted with a molecular weight of about 155 kilodaltons, consistent with a properly disulfide-linked antibody molecule with 2 light and 2 heavy chains. Upon reduction the molecular weight shifts to 2 approximately equally abundant proteins of 22.5 and 55 kilodaltons. The protein generated from the pool is indistinguishable from the antibody produced by the well characterized expression clone, with no apparent increase of free heavy or light chain expressed by the pool.

CONCLUSION

The efficient expression system described herein utilizes vectors consisting of promoter/enhancer elements followed by an intron containing the selectable marker coding sequence, followed by the cDNA of interest and a polyadenylation signal.

Several splice donor site sequences were tested for their effect on colony number and expression of the cDNA of interest. A non-functional splice donor site, splice donor sites found in an intron between exons 3 and 4 of mutant (mutant ras) and normal (WT ras) forms of the Harvey Ras gene and another efficient SD site (see Example 3) were used. The vectors were designed to direct expression of dicistronic primary transcripts. Within a transfected cell some of the transcripts remain full length while the remainder are spliced to excise the DHFR coding sequence. When the splice donor site is weakened or destroyed an increase in colony number is observed.

Expression levels show the inverse pattern, with the most efficient splice donor sites generating the highest levels of tPA, TNFr immunoadhesin or anti-IgE V_H .

The homogeneity of expression of clones generated by the ras splice donor site intron DHFR vectors was compared to clones generated from a conventional vector with a separate promoter/enhancer and polyadenylation signal for each DHFR and tPA. The DHFR intron vector gives rise to colonies that are much more homogeneous with regard to expression than those generated by the conventional vector. Non-expressing clones derived from the conventional vector may be the result of breaks in the tPA or

TNFr-IgG domain of the plasmid during integration into the genome or the result of methylation of promoter elements (Busslinger et al., Cell, 34:197-206 [1983]; Watt et al., Genes and Development, 2:1136-1143 [1988]) driving tPA or TNFr-IgG expression. Promoter silencing by methylation or
5 breaks in the DHFR-intron vectors would very likely render them incapable of conferring a DHFR positive phenotype.

It was found that pools generated by the DHFR-intron vectors could be amplified in methotrexate and would increase in expression by a factor of 8.4 (tPA), or 9.8 (TNFr-IgG). Pools from conventional vectors increased
10 by only 2.8 and 3.0 fold for tPA and TNFr-IgG when amplified similarly. Amplified pools resulted in 9 fold higher tPA levels and 15 fold higher TNFr-IgG levels when compared to the conventional vector amplified pools.

Without being limited to any theory, the increase in expression of methotrexate resistant pools derived from the dicistronic vectors is likely
15 due to the transcriptional linkage of DHFR and the product; when cells are selected for increased DHFR expression they consistently over-express product. Conventional approaches lack selectable marker and cDNA expression linkage and therefore methotrexate amplification often generates DHFR overexpression without the concomitant increase in product expression.

20 A further increase of 4 and 6.3 fold in expression were obtained when amplified tPA and TNFr-IgG pools were transferred from the media used for the selections and amplifications to a nutrient rich production medium.

In Example 3, the expression vector had a splice donor site that more closely matches the consensus splice donor sequence and had the heavy chain
25 of a humanized anti-IgE antibody inserted downstream. This vector was linearized and co-electroporated with a second linearized vector that expresses the hygromycin resistance gene and the light chain of the antibody each under the control of its own promoter/enhancer and poly-A signals. An excess of light chain expression vector over the heavy chain
30 dicistronic expression vector was used to bias in favor of light chain expression. Clones and a pool were generated after hygromycin B and DHFR selections. The clones were found to express relatively consistent, high levels of antibody, as did the pool. The 1 μ M pool achieved a titer of 41mg/L when grown under optimal conditions in suspension culture.

35 The anti-IgE antibody was assessed by metabolic labeling followed by SDS/PAGE under reducing and non reducing conditions and found to be indistinguishable from the protein expressed by a highly characterized clonal cell line. Of particular importance is the finding that no free light chain is observed in the pool relative to the clone.

40 A stable expression system for CHO cells has been developed that produces high levels of recombinant proteins rapidly and with less effort than that required by other expression systems. The vector system generates stable clones that express consistently high levels thereby reducing the number of clones that must be screened to obtain a highly
45 productive clonal line. Alternatively, pools have been used to conveniently generate moderate to high levels of protein. This approach

may be particularly useful when a number of related proteins are to be expressed and compared.

Without being limited to this theory, it is possible the vectors that have very efficient splice donor sites generate very productive clones because so little transcript remains non spliced that only integration events that lead to the generation of high levels of RNA produce enough DHFR protein to give rise to colonies in selective medium. The high level of spliced message from such clones is then translated into abundant amounts of the protein of interest. Pools of clones made concurrently by introducing conventional vectors expressed lower levels of protein, and were unstable with regard to long term expression, and expression could not be appreciably increased when the cells were subjected to methotrexate amplification.

The system developed herein is versatile in that it allows high levels of single and multiple subunit polypeptides to be rapidly generated from clones or pools of stable transfectants. This expression system combines the advantages of transient expression systems (rapid and labor non intensive generation of research amounts of protein) with the concurrent development of highly productive stable production cell lines.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

- 5 (i) APPLICANT: GENENTECH, INC.
- (ii) TITLE OF INVENTION: METHOD FOR SELECTING HIGH-EXPRESSING HOST CELLS
- 10 (iii) NUMBER OF SEQUENCES: 4
- (iv) CORRESPONDENCE ADDRESS:
- (A) ADDRESSEE: Genentech, Inc.
 - (B) STREET: 460 Point San Bruno Blvd
 - (C) CITY: South San Francisco
 - 15 (D) STATE: California
 - (E) COUNTRY: USA
 - (F) ZIP: 94080
- (v) COMPUTER READABLE FORM:
- 20 (A) MEDIUM TYPE: 5.25 inch, 360 Kb floppy disk
 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: patin (Genentech)
- 25 (vi) CURRENT APPLICATION DATA:
- (A) APPLICATION NUMBER:
 - (B) FILING DATE:
 - (C) CLASSIFICATION:
- 30 (vii) PRIOR APPLICATION DATA:
- (A) APPLICATION NUMBER: 08/286740
 - (B) FILING DATE: 05-AUG-1994
- (viii) ATTORNEY/AGENT INFORMATION:
- 35 (A) NAME: Lee, Wendy M.
 - (B) REGISTRATION NUMBER: 00,000
 - (C) REFERENCE/DOCKET NUMBER: 798PCT
- (ix) TELECOMMUNICATION INFORMATION:
- 40 (A) TELEPHONE: 415/225-1994
 - (B) TELEFAX: 415/952-9881
 - (C) TELEX: 910/371-7168

(2) INFORMATION FOR SEQ ID NO:1:

- 45 (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 7360 bases
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - 50 (D) TOPOLOGY: linear
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

55 TTCGAGCTCG CCCGACATTG ATTATTGACT AGTTATTAAT AGTAATCAAT 50

TACGGGGTCA TTAGTTCATA GCCCATATAT GGAGTTCCGC GTTACATAAC 100

60 TTACGGTAAA TGGCCCGCCT GGCTGACCGC CCAACGACCC CCGCCCATTG 150

ACGTCAATAA TGACGTATGT TCCCATAGTA ACGCCAATAG GGACTTTCCA 200

65 TTGACGTCAA TGGGTGGAGT ATTTACGGTA AACTGCCCAC TTGGCAGTAC 250

ATCAAGTGTA TCATATGCCA AGTACGCCCC CTATTGACGT CAATGACGGT 300
5 AAATGGCCCG CCTGGCATTG TGCCAGTAC ATGACCTTAT GGGACTTTCC 350
TACTTGGCAG TACATCTACG TATTAGTCAT CGCTATTACC ATGGTGATGC 400
10 GGTTTTGGCA GTACATCAAT GGGCGTGGAT AGCGGTTTGA CTCACGGGGA 450
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15 AAATCAACGG GACTTTCCAA AATGTCGTAA CAACTCCGCC CCATTGACGC 550
AAATGGGCGG TAGGCGTGTA CGGTGGGAGG TCTATATAAG CAGAGCTCGT 600
20 TTAGTGAACC GTCAGATCGC CTGGAGACGC CATCCACGCT GTTTTGACCT 650
CCATAGAAGA CACCGGGACC GATCCAGCCT CCGCGGCCGG GAACGGTGCA 700
25 TTGGAACGCG GATTCCCCGT GCCAAGAGTG CTGTAAGTAC CGCCTATAGA 750
30 GCGATAAGAG GATTTTATCC CCGCTGCCAT CATGGTTCGA CCATTGAACT 800
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5 GGCTACAATT AATACATAAC CTTATGTATC ATACACATAG ATTTAGGTGA 1500

CACTATAGAA TAACATCCAC TTTGCCTTTC TCTCCACAGG TGTCACTCCA 1550

10 GGTCAACTGC ACCTCGGTTC TAAGCTTGGG CTGCAGGTCG CCGTGAATTT 1600

AAGGGACGCT GTGAAGCAAT CATGGATGCA ATGAAGAGAG GGCTCTGCTG 1650

15 TGTGCTGCTG CTGTGTGGAG CAGTCTTCGT TTCGCCCAGC CAGGAAATCC 1700

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30 GGGGGCACCT GCCAGCAGGC CCTGTACTTC TCAGATTTCTG TGTGCCAGTG 1950

CCCGAAGGA TTTGCTGGGA AGTGCTGTGA AATAGATACC AGGGCCACGT 2000

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40 AGTGGCGCCG AGTGCACCAA CTGGAACAGC AGCGCGTTGG CCCAGAAGCC 2100

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65 CCCTCCTGCT CCACCTGCGG CCTGAGACAG TACAGCCAGC CTCAGTTTCG 2550

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25 TGTGAGCTCT CCGGCTACGG CAAGCATGAG GCCTTGTCTC CTTTCTATTC 3000

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30 CATCACAACA TTTACTTAAC AGAACAGTCA CCGACAACAT GCTGTGTGCT 3100

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50 TCTCCAGACC CACCACACCG CAGAAGCGGG ACGAGACCCT ACAGGAGAGG 3450

55 GAAGAGTGCA TTTTCCCAGA TACTTCCCAT TTTGGAAGTT TTCAGGACTT 3500

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GATGGCCGCC ATGGCCCAAC TTGTTTATTG CAGCTTATAA TGGTTACAAA 3650

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25 CATTCAAATA TGTATCCGCT CATGAGACAA TAACCCTGAT AAATGCTTCA 5300

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30 TTATTCCCTT TTTTGCGGCA TTTTGCCCTT CTGTTTTTGC TCACCCAGAA 5400

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35 TTACATCGAA CTGGATCTCA ACAGCGGTAA GATCCTTGAG AGTTTTCGCC 5500

CCGAAGAACG TTTTCCAATG ATGAGCACTT TTAAAGTTCT GCTATGTGGC 5550

GCGGTATTAT CCCGTGATGA CGCCGGGCAA GAGCAACTCG GTCGCCGCAT 5600

45 ACACTATTCT CAGAATGACT TGGTTGAGTA CTCACCAGTC ACAGAAAAGC 5650

ATCTTACGGA TGGCATGACA GTAAGAGAAT TATGCAGTGC TGCCATAACC 5700

50 ATGAGTGATA ACACTGCGGC CAACTTACTT CTGACAACGA TCGGAGGACC 5750

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60 GACACCACGA TGCCAGCAGC AATGGCAACA ACGTTGCGCA AACTATTAAC 5900

TGGCGAACTA CTTACTCTAG CTTCCCGGCA ACAATTAATA GACTGGATGG 5950

65 AGGCGGATAA AGTTGCAGGA CCACTTCTGC GCTCGGCCCT TCCGGCTGGC 6000

TGGTTTATTG CTGATAAATC TGGAGCCGGT GAGCGTGGGT CTCGCGGTAT 6050

5 CATTGCAGCA CTGGGGCCAG ATGGTAAGCC CTCCCGTATC GTAGTTATCT 6100

ACACGACGGG GAGTCAGGCA ACTATGGATG AACGAAATAG ACAGATCGCT 6150

10 GAGATAGGTG CCTCACTGAT TAAGCATTGG TAACTGTCAG ACCAAGTTTA 6200

CTCATATATA CTTTAGATTG ATTTAAAACT TCATTTTAA TTTAAAAGGA 6250

15 TCTAGGTGAA GATCCTTTTT GATAATCTCA TGACCAAAT CCCTTAACGT 6300

GAGTTTTCGT TCCACTGAGC GTCAGACCCC GTAGAAAAGA TCAAAGGATC 6350

20 TTCTTGAGAT CCTTTTTTTC TGCGCGTAAT CTGCTGCTTG CAAACAAAAA 6400

25 AACCACCGCT ACCAGCGGTG GTTTGTTTGC CGGATCAAGA GCTACCAACT 6450

CTTTTTCCGA AGGTAAGTGG CTTTCAAGCA GCGCAGATAC CAAATACTGT 6500

30 CCTTCTAGTG TAGCCGTAGT TAGGCCACCA CTTCAAGAAC TCTGTAGCAC 6550

CGCCTACATA CCTCGCTCTG CTAATCCTGT TACCAGTGGC TGCTGCCAGT 6600

35 GCGGATAAGT CGTGTCTTAC CGGGTTGGAC TCAAGACGAT AGTTACCGGA 6650

40 TAAGGCGCAG CGGTCGGGCT GAACGGGGGG TTCGTGCACA CAGCCCAGCT 6700

TGGAGCGAAC GACCTACACC GAACTGAGAT ACCTACAGCG TGAGCATTGA 6750

45 GAAAGCGCCA CGCTTCCCGA AGGGAGAAAG GCGGACAGGT ATCCGGTAAG 6800

CGGCAGGGTC GGAACAGGAG AGCGCACGAG GGAGCTTCCA GGGGGAAACG 6850

50 CCTGGTATCT TTATAGTCCT GTCGGGTTTC GCCACCTCTG ACTTGAGCGT 6900

55 CGATTTTTGT GATGCTCGTC AGGGGGGCGG AGCCTATGGA AAAACGCCAG 6950

CAACGCGGCC TTTTACGGT TCCTGGCCTT TTGCTGGCCT TTTGCTCACA 7000

60 TGTTCTTTCC TGCGTTATCC CCTGATTCTG TGGATAACCG TATTACCGCC 7050

TTTGAGTGAG CTGATACCGC TCGCCGAGC CGAACGACCG AGCGCAGCGA 7100

65 GTCAGTGAGC GAGGAAGCGG AAGAGCGCCC AATACGCAA CCGCCTCTCC 7150

CCGCGCGTTG GCCGATTCAT TAATCCAGCT GGCACGACAG GTTTCCTCGAC 7200
5 TGGAAAGCGG GCAGTGAGCG CAACGCAATT AATGTGAGTT ACCTCACTCA 7250
TTAGGCACCC CAGGCTTTAC ACTTTATGCT TCCGGCTCGT ATGTTGTGTG 7300
10 GAATTGTGAG CGGATAACAA TTTCACACAG GAAACAGCTA TGACCATGAT 7350
TACGAATTAA 7360

15

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:
20 (A) LENGTH: 6889 bases
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

25 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

TTCGAGCTCG CCCGACATTG ATTATTGACT AGTTATTAAT AGTAATCAAT 50
30 TACGGGGTCA TTAGTTCATA GCCCATATAT GGAGTTCGCG GTTACATAAC 100
TTACGGTAAA TGGCCCGCCT GGCTGACCGC CCAACGACCC CCGCCCATTG 150
35 ACGTCAATAA TGACGTATGT TCCCATAGTA ACGCCAATAG GGACTTTCCA 200
TTGACGTCAA TGGGTGGAGT ATTTACGGTA AACTGCCCCAC TTGGCAGTAC 250
ATCAAGTGTA TCATATGCCA AGTACGCCCC CTATTGACGT CAATGACGGT 300
45 AAATGGCCCG CCTGGCATTG TGCCCAGTAC ATGACCTTAT GGGACTTTCC 350
TACTTGGCAG TACATCTACG TATTAGTCAT CGCTATTACC ATGGTGATGC 400
50 GGTTTTGGCA GTACATCAAT GGGCGTGGAT AGCGGTTTGA CTCACGGGGA 450
TTTCCAAGTC TCCACCCCAT TGACGTCAAT GGGAGTTTGT TTTGGCACCA 500
AAATCAACGG GACTTTCCAA AATGTCGTAA CAACTCCGCC CCATTGACGC 550
60 AAATGGGCGG TAGGCGTGTA CGGTGGGAGG TCTATATAAG CAGAGCTCGT 600
TTAGTGAACC GTCAGATCGC CTGGAGACGC CATCCACGCT GTTTTGACCT 650
65 CCATAGAAGA CACCGGGACC GATCCAGCCT CCGCGGCCCG GAACGGTGCA 700

TTGGAACGCG GATTCCCCGT GCCAAGAGTG CTGTAAGTAC CGCCTATAGA 750

5 GCGATAAGAG GATTTTATCC CCGCTGCCAT CATGGTTCGA CCATTGAACT 800

GCATCGTCGC CGTGTCCCAA AATATGGGGA TTGGCAAGAA CGGAGACCTA 850

10 CCCTGCCCTC CGCTCAGGAA CGCGTTCAAG TACTTCCAAA GAATGACCAC 900

AACCTCTTCA GTGGAAGGTA AACAGAATCT GGTGATTATG GGTAGGAAAA 950

15 CCTGGTTCTC CATTCCTGAG AAGAATCGAC CTTTAAAGGA CAGAATTAAT 1000

ATAGTTCTCA GTAGAGAACT CAAAGAACCA CCACGAGGAG CTCATTTTCT 1050

20 TGCCAAAAGT TTGGATGATG CCTTAAGACT TATTGAACAA CCGGAATTGG 1100

25 CAAGTAAAGT AGACATGGTT TGGATAGTCG GAGGCAGTTC TGTTTACCAG 1150

GAAGCCATGA ATCAACCAGG CCACCTTAGA CTCTTTGTGA CAAGGATCAT 1200

30 GCAGGAATTT GAAAGTGACA CGTTTTTCCC AGAAATTGAT TTGGGGAAAT 1250

ATAAACCTCT CCCAGAATAC CCAGGCGTCC TCTCTGAGGT CCAGGAGGAA 1300

35 AAAGGCATCA AGTATAAGTT TGAAGTCTAC GAGAAGAAAG ACTAACAGGA 1350

40 AGATGCTTTC AAGTTCTCTG CTCCCCTCCT AAAGCTATGC ATTTTTATAA 1400

GACCATGGGA CTTTGTCTGG CTTTAGACCC CTTGGCTTC GTTAGAACGC 1450

45 GGCTACAATT AATACATAAC CTTATGTATC ATACACATAG ATTTAGGTGA 1500

CACTATAGAA TAACATCCAC TTTGCCTTTC TCTCCACAGG TGTCACCTCA 1550

50 GGTCAACTGC ACCTCGGTTC TATCGATTGA ATTCCCCGGC CATAGCTGTC 1600

55 TGGCATGGGC CTCTCCACCG TGCCTGACCT GCTGCTGCCG CTGGTGCTCC 1650

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60 CACCTAGGGG ACAGGGAGAA GAGAGATAGT GTGTGTCCCC AAGGAAAATA 1750

TATCCACCCT CAAAATAATT CGATTTGCTG TACCAAGTGC CACAAAGGAA 1800

65 CCTACTTGTA CAATGACTGT CCAGGCCCGG GGCAGGATAC GGAAGTGCAGG 1850

GAGTGTGAGA GCGGCTCCTT CACCGCTTCA GAAAACCACC TCAGACACTG 1900
5 CCTCAGCTGC TCCAAATGCC GAAAGGAAAT GGGTCAGGTG GAGATCTCTT 1950
CTTGCACAGT GGACCGGGAC ACCGTGTGTG GCTGCAGGAA GAACCAGTAC 2000
10 CGGCATTATT GGAGTGAAAA CCTTTTCCAG TGCTTCAATT GCAGCCTCTG 2050
CCTCAATGGG ACCGTGCACC TCTCCTGCCA GGAGAAACAG AACACCGTGT 2100
15 GCACCTGCCA TGCAGGTTTC TTTCTAAGAG AAAACGAGTG TGTCTCCTGT 2150
AGTAACTGTA AGAAAAGCCT GGAGTGCACG AAGTTGTGCC TACCCAGAT 2200
20 TGAGAATGTT AAGGGCACTG AGGACTCAGG CACCACAGAC AAGAGAGTTG 2250
25 AGCTCAAAAC CCCACTTGGT GACACAAC TC ACACATGCCC ACGGTGCCCA 2300
GAGCCCAAAT CTTGTGACAC ACCTCCCCCG TGCCACGGT GCCCAGAGCC 2350
30 CAAATCTTGT GACACACCTC CCCCATGCCC ACGGTGCCCA GAGCCCAAAT 2400
CTTGTGACAC ACCTCCCCCA TGCCACGGT GCCCAGCACC TGAATCCTG 2450
35 GGAGGACCGT CAGTCTTCCT CTTCCCCCA AAACCAAGG ATACCCTTAT 2500
40 GATTTCCCGG ACCCCTGAGG TCACGTGCGT GGTGGTGGAC GTGAGCCACG 2550
AAGACCCCGA GGTCCAGTTC AAGTGGTACG TGGACGGCGT GGAGGTGCAT 2600
45 AATGCCAAGA CAAAGCCGCG GGAGGAGCAG TTCAACAGCA CGTTCCGTGT 2650
GGTCAGCGTC CTCACCGTCC TGCACCAGGA CTGGCTGAAC GGCAAGGAGT 2700
50 ACAAGTGCAA GGTCTCCAAC AAAGCCCTCC CAGCCCCCAT CGAGAAAACC 2750
55 ATCTCCAAAA CCAAAGGACA GCCCCGAGAA CCACAGGTGT ACACCCTGCC 2800
CCCATCCCGG GAGGAGATGA CCAAGAACCA GGTCAGCCTG ACCTGCCTGG 2850
60 TCAAAGGCTT CTACCCAGC GACATCGCCG TGGAGTGGGA GAGCAGCGGG 2900
CAGCCGGAGA ACAACTACAA CACCACGCCT CCCATGCTGG ACTCCGACGG 2950
65 CTCCTTCTTC CTCTACAGCA AGCTCACCGT GGACAAGAGC AGGTGGCAGC 3000

AGGGGAACAT CTTCTCATGC TCCGTGATGC ATGAGGCTCT GCACAACCGC 3050

5 TTCACGCAGA AGAGCCTCTC CCTGTCTCCG GGTAATGAG TGCACGCGCC 3100

GGGGATCCTC TAGAGTCGAC CTGCAGAAGC TTGGCCGCCA TGGCCCAACT 3150

10 TGTTTATTGC AGCTTATAAT GGTTACAAAT AAAGCAATAG CATCACAAAT 3200

TTCACAAATA AAGCATTTTT TCACTGTCAT TCTAGTTGTG GTTTGTCCAA 3250

15 ACTCATCAAT GSTATCTTATC ATGTCTGGAT CGATCGGGAA TTAATTCGGC 3300

GCAGCACCAT GGCCTGAAAT AACCTCTGAA AGAGGAACTT GGTTAGGTAC 3350

20 CTTCTGAGGC GGAAAGAACC AGCTGTGGAA TGTGTGTCAG TTAGGGTGTG 3400

25 GAAAGTCCCC AGGCTCCCCA GCAGGCAGAA GTATGCAAAG CATGCATCTC 3450

AATTAGTCAG CAACCAGGTG TGGAAAGTCC CCAGGCTCCC CAGCAGGCAG 3500

30 AAGTATGCAA AGCATGCATC TCAATTAGTC AGCAACCATA GTCCCGCCCC 3550

TAACTCCGCC CATCCCGCCC CTAAGTCCGC CCAGTTCCGC CCATTCTCCG 3600

35 CCCCATGGCT GACTAATTTT TTTTATTTAT GCAGAGGCCG AGGCCGCCTC 3650

40 GGCCTCTGAG CTATTCCAGA AGTAGTGAGG AGGCTTTTTT GGAGGCCTAG 3700

GCTTTTGCAA AAAGCTGTTA ACAGCTTGGC ACTGGCCGTC GTTTTACAAC 3750

45 GTCGTGACTG GGAAAACCTT GGCCTTACCC AACTTAATCG CTTGTCAGCA 3800

CATCCCCCCT TCGCCAGCTG GCGTAATAGC GAAGAGGCCC GCACCGATCG 3850

50 CCCTTCCCAA CAGTTGCGTA GCCTGAATGG CGAATGGCGC CTGATGCGGT 3900

55 ATTTTCTCCT TACGCATCTG TCGGGTATTT CACACCGCAT ACGTCAAAGC 3950

AACCATAGTA CGCGCCCTGT AGCGGCGCAT TAAGCGCGGC GGGTGTGGTG 4000

60 GTTACGCGCA GCGTGACCGC TACACTTGCC AGCGCCCTAG CGCCCGCTCC 4050

TTTCGCTTTC TTCCCTTCCT TTCTCGCCAC GTTCGCCGGC TTTCCCGTC 4100

65 AAGCTCTAAA TCGGGGGCTC CCTTTAGGGT TCCGATTAG TGCTTTACGG 4150

CACCTCGACC CCAAAAAACT TGATTGGGT GATGGTTCAC GTAGTGGGCC 4200

5 ATCGCCCTGA TAGACGGTTT TCGCCCTT GACGTTGGAG TCCACGTTCT 4250

TTAATAGTGG ACTCTTGTT CAACTGGAA CAACACTCAA CCCTATCTCG 4300

10 GGCTATTCTT TTGATTATA AGGGATTTG CCGATTTCGG CCTATTGGTT 4350

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15 TAACGTTTAC AATTTTATGG TGCACCTCA GTACAATCTG CTCTGATGCC 4450

GCATAGTTAA GCCAACTCCG CTATCGCTAC GTGACTGGGT CATGGCTGCG 4500

20 CCCCACACC CGCCAACACC CGCTGACGCG CCCTGACGGG CTTGTCTGCT 4550

25 CCCGGCATCC GCTTACAGAC AAGCTGTGAC CGTCTCCGGG AGCTGCATGT 4600

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30 AGACGAAAGG GCCTCGTGAT ACGCCTATTT TTATAGGTTA ATGTCATGAT 4700

AATAATGGTT TCTTAGACGT CAGGTGGCAC TTTTCGGGGA AATGTGCGCG 4750

35 GAACCCCTAT TTGTTTATTT TTCTAAATAC ATTCAAATAT GTATCCGCTC 4800

40 ATGAGACAAT AACCTGATA AATGCTTCAA TAATATTGAA AAAGGAAGAG 4850

TATGAGTATT CAACATTTCC GTGTCGCCCT TATTCCCTTT TTTGCGGCAT 4900

45 TTTGCCTTCC TGTTTTTGCT CACCCAGAAA CGCTGGTGAA AGTAAAAGAT 4950

50 GCTGAAGATC AGTTGGGTGC ACGAGTGGGT TACATCGAAC TGGATCTCAA 5000

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55 TGAGCACTTT TAAAGTTCTG CTATGTGGCG CGGTATTATC CCGTGATGAC 5100

GCCGGGCAAG AGCAACTCGG TCGCCGCATA CACTATTCTC AGAATGACTT 5150

60 GGTTGAGTAC TCACCACTCA CAGAAAAGCA TCTTACGGAT GGCATGACAG 5200

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65 AACTTACTTC TGACAACGAT CGGAGGACCG AAGGAGCTAA CCGCTTTTTT 5300

GCACAACATG GGGGATCATG TAACTCGCCT TGATCGTTGG GAACCGGAGC 5350
TGAATGAAGC CATAACAAAC GACGAGCGTG ACACCACGAT GCCAGCAGCA 5400
5 ATGGCAACAA CGTTGCGCAA ACTATTAACT GGCGAACCTAC TTACTCTAGC 5450
TTCCCGGCAA CAATTAATAG ACTGGATGGA GGCGGATAAA GTTGCAGGAC 5500
CACTTCTGCG CTCGGCCCTT CCGGCTGGCT GGTTTATTGC TGATAAATCT 5550
15 GGAGCCGGTG AGCGTGGGTC TCGCGGTATC ATTGCAGCAC TGGGGCCAGA 5600
TGGTAAGCCC TCCCGTATCG TAGTTATCTA CACGACGGGG AGTCAGGCAA 5650
20 CTATGGATGA ACGAAATAGA CAGATCGCTG AGATAGGTGC CTCACTGATT 5700
AAGCATTGGT AACTGTCAGA CCAAGTTTAC TCATATATAC TTTAGATTGA 5750
TTTAAAACTT CATTTTAAAT TTAAAAGGAT CTAGGTGAAG ATCCTTTTTTG 5800
30 ATAATCTCAT GACCAAATC CCTTAACGTG AGTTTTCGTT CCACTGAGCG 5850
TCAGACCCCG TAGAAAAGAT CAAAGGATCT TCTTGAGATC CTTTTTTTCT 5900
35 GCGCGTAATC TGCTGCTTGC AAACAAAAAA ACCACCGCTA CCAGCGGTGG 5950
TTTGTTTGCC GGATCAAGAG CTACCAACTC TTTTCCGAA GGTAAGTGGC 6000
TTCAGCAGAG CGCAGATACC AAATACTGTC CTTCTAGTGT AGCCGTAGTT 6050
45 AGGCCACCAC TTCAAGAACT CTGTAGCACC GCCTACATAC CTCGCTCTGC 6100
TAATCCTGTT ACCAGTGGCT GCTGCCAGTG GCGATAAGTC GTGTCTTACC 6150
50 GGGTTGGACT CAAGACGATA GTTACCGGAT AAGGCGCAGC GGTCGGGCTG 6200
AACGGGGGGT TCGTGCACAC AGCCCAGCTT GGAGCGAACG ACCTACACCG 6250
AACTGAGATA CCTACAGCGT GAGCATTGAG AAAGCGCCAC GCTTCCCGAA 6300
60 GGGAGAAAGG CGGACAGGTA TCCGGTAAGC GGCAGGGTCG GAACAGGAGA 6350
GCGCACGAGG GAGCTTCCAG GGGGAAACGC CTGGTATCTT TATAGTCCTG 6400
65 TCGGGTTTCG CCACCTCTGA CTTGAGCGTC GATTTTTGTG ATGCTCGTCA 6450

GGGGGGCGGA GCCTATGGAA AAACGCCAGC AACGCGGCCT TTTTACGGTT 6500
5 CCTGGCCTTT TGCTGGCCTT TTGCTCACAT GTTCTTTCCT GCGTTATCCC 6550
CTGATTCTGT GGATAACCGT ATTACCGCCT TTGAGTGAGC TGATAACCGCT 6600
10 CGCCGCAGCC GAACGACCGA GCGCAGCGAG TCAGTGAGCG AGGAAGCGGA 6650
AGAGCGCCCA ATACGCAAAC CGCCTCTCCC CGCGCGTTGG CCGATTTCATT 6700
15 AATCCAGCTG GCACGACAGG TTTCCCGACT GGAAAGCGGG CAGTGAGCGC 6750
20 AACGCAATTA ATGTGAGTTA CCTCACTCAT TAGGCACCCC AGGCTTTACA 6800
CTTTATGCTT CCGGCTCGTA TGTGTGTGG AATTGTGAGC GGATAACAAT 6850
25 TTCACACAGG AACAGCTAT GACCATGATT ACGAATTAA 6889

30 (2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:

- 35 (A) LENGTH: 6557 bases
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

40 TTCGAGCTCG CCCGACATTG ATTATTGACT AGAGTCGATC GACAGCTGTG 50
GAATGTGTGT CAGTTAGGGT GTGGAAAGTC CCCAGGCTCC CCAGCAGGCA 100
45 GAAGTATGCA AAGCATGCAT CTCAATTAGT CAGCAACCAG GTGTGGAAAG 150
TCCCCAGGCT CCCCAGCAGG CAGAAGTATG CAAAGCATGC ATCTCAATTA 200
50 GTCAGCAACC ATAGTCCCGC CCCTAACTCC GCCCATCCCG CCCCTAACTC 250
55 CGCCCAGTTC CGCCCATTCT CCGCCCCATG GCTGACTAAT TTTTTTTATT 300
TATGCAGAGG CCGAGGCCGC CTCGGCCTCT GAGCTATTCC AGAAGTAGTG 350
60 AGGAGGCTTT TTTGGAGGCC TAGGCTTTTG CAAAAAGCTA GCTTATCCGG 400
CCGGGAACGG TGCATTGGAA CGCGGATTCC CCGTGCCAAG AGTGACGTAA 450
65 GTACCGCCTA TAGAGCGATA AGAGGATTTT ATCCCCGCTG CCATCATGGT 500

TCGACCATTG AACTGCATCG TCGCCGTGTC CCAAAATATG GGGATTGGCA 550
5 AGAACGGAGA CCTACCCCTGG CCTCCGCTCA GGAACGAGTT CAAGTACTTC 600
CAAAGAATGA CCACAACCTC TTCAGTGGAA GGTAAACAGA ATCTGGTGAT 650
10 TATGGGTAGG AAAACCTGGT TCTCCATTCC TGAGAAGAAT CGACCTTTAA 700
AGGACAGAAT TAATATAGTT CTCAGTAGAG AACTCAAAGA ACCACCACGA 750
15 GGAGCTCATT TTCTTGCCAA AAGTTTGGAT GATGCCTTAA GACTTATTGA 800
ACAACCGGAA TTGGCAAGTA AAGTAGACAT GGTTTGGATA GTCGGAGGCA 850
20 GTTCTGTTTA CCAGGAAGCC ATGAATCAAC CAGGCCACCT TAGACTCTTT 900
25 GTGACAAGGA TCATGCAGGA ATTTGAAAGT GACACGTTTT TCCCAGAAAT 950
TGATTTGGGG AAATATAAAC CTCTCCCAGA ATACCCAGGC GTCCTCTCTG 1000
30 AGGTCCAGGA GGAAAAAGGC ATCAAGTATA AGTTTGAAGT CTACGAGAAG 1050
AAAGACTAAC AGGAAGATGC TTTCAAGTTC TCTGCTCCCC TCCTAAAGCT 1100
35 ATGCATTTTT ATAAGACCAT GGGACTTTTG CTGGCTTTAG ATCCCCTTGG 1150
40 CTTCGTTAGA ACGCAGCTAC AATTAATACA TAACCTTATG TATCATACAC 1200
ATACGATTTA GGTGACACTA TAGATAACAT CCACTTTGCC TTTCTCTCCA 1250
45 CAGGTGTCCA CTCCCAGGTC CAACTGCACC TCGGTTCTAT CGATTGAATT 1300
CCACCATGGG ATGGTCATGT ATCATCCTTT TTCTAGTAGC AACTGCAACT 1350
50 GGAGTACATT CAGAAGTTCA GCTGGTGGAG TCTGGCGGTG GCCTGGTGCA 1400
55 GCCAGGGGGC TCACTCCGTT TGTCTGTGC AGTTTCTGGC TACTCCATCA 1450
CCTCCGGATA TAGCTGGAAC TGGATCCGTC AGGCCCCGGG TAAGGGCCTG 1500
60 GAATGGGTTG CATCGATTAC GTATGCCGGA TCGACTAACT ATAACCCTAG 1550
CGTCAAGGGC CGTATCACTA TAAGTCGCGA CGATTCCAAA AACACATTCT 1600
65 ACCTGCAGAT GAACAGCCTG CGTGCTGAGG AACTGCCGT CTATTATTGT 1650

GCTCGAGGCA GCCACTATTT CGGCGCCTGG CACTTCGCCG TGTGGGGTCA 1700
5 AGGAACCCTG GTCACCGTCT CCTCGGCCTC CACCAAGGGC CCATCGGTCT 1750
TCCCCCTGGC ACCCTCCTCC AAGAGCACCT CTGGGGGCAC AGCGGCCCTG 1800
10 GGCTGCCTGG TCAAGGACTA CTTCCCCGAA CCGGTGACGG TGTCTGGAA 1850
CTCAGGCGCC CTGACCAGCG GCGTGACAC CTTCCCGGCT GTCCTACAGT 1900
15 CCTCAGGACT CTACTCCCTC AGCAGCGTGG TGA CTGTGCC CTCTAGCAGC 1950
TTGGGCACCC AGACCTACAT CTGCAACGTG AATCACAAGC CCAGCAACAC 2000
20 CAAGGTGGAC AAGAAAGTTG AGCCCAAATC TTGTGACAAA ACTCACACAT 2050
GCCACCGTG CCCAGCACCT GAACTCCTGG GGGGACCGTC AGTCTTCCTC 2100
TTCCCCCAA AACCAAGGA CACCCTCATG ATCTCCCGGA CCCCTGAGGT 2150
30 CACATGCGTG GTGGTGGACG TGAGCCACGA AGACCCTGAG GTCAAGTTCA 2200
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35 GAGGAGCAGT ACAACAGCAC GTACCGTGTG GTCAGCGTCC TCACCGTCCT 2300
GCACCAGGAC TGGCTGAATG GCAAGGAGTA CAAGTGCAAG GTCTCCAACA 2350
AAGCCCTCCC AGCCCCATC GAGAAAACCA TCTCCAAAGC CAAAGGGCAG 2400
45 CCCCAGAAC CACAGGTGTA CACCCTGCCC CCATCCCGGG AAGAGATGAC 2450
CAAGAACCAG GTCAGCCTGA CCTGCCTGGT CAAAGGCTTC TATCCAGCG 2500
50 ACATCGCCGT GGAGTGGGAG AGCAATGGGC AGCCGGAGAA CAACTACAAG 2550
ACCACGCCTC CCGTGCTGGA CTCCGACGGC TCCTTCTTCC TCTACAGCAA 2600
GCTCACCGTG GACAAGAGCA GGTGGCAGCA GGGGAACGTC TTCTCATGCT 2650
60 CCGTGATGCA TGAGGCTCTG CACAACCACT ACACGCAGAA GAGCCTCTCC 2700
CTGTCTCCGG GTAAATGAGT GCGACGGCCC TAGAGTCGAC CTGCAGAAGC 2750
65 TTGGCCGCCA TGGCCCAACT TGTATTATGC AGCTTATAAT GGTTACAAAT 2800

AAAGCAATAG CATCACAAAT TTCACAAATA AAGCATTTTT TCACTGCAT 2850
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5 CGATCGGGAA TTAATTCGGC GCAGCACCAT GGCCTGAAAT AACCTCTGAA 2950
AGAGGAACTT GGTTAGGTAC CTTCTGAGGC GGAAAGAACC AGCTGTGGAA 3000
TGTGTGTCAG TTAGGGTGTG GAAAGTCCCC AGGCTCCCCA GCAGGCAGAA 3050
15 GTATGCAAAG CATGCATCTC AATTAGTCAG CAACCAGGTG TGGAAAGTCC 3100
CCAGGCTCCC CAGCAGGCAG AAGTATGCAA AGCATGCATC TCAATTAGTC 3150
20 AGCAACCATA GTCCCGCCCC TAACTCCGCC CATCCCGCCC CTAACCTCCG 3200
CCAGTCCGC CCATTCTCCG CCCCATGGCT GACTAATTTT TTTTATTTAT 3250
GCAGAGGCCG AGGCCGCCTC GGCCTCTGAG CTATTCCAGA AGTAGTGAGG 3300
30 AGGCTTTTTT GGAGGCCTAG GCTTTTGCAA AAAGCTGTTA CCTCGAGCGG 3350
CCGCTTAATT AAGGCGCGCC ATTTAAATCC TGCAGGTAAC AGCTTGGCAC 3400
35 TGGCCGTCGT TTTACAACGT CGTGACTGGG AAAACCCTGG CGTTACCCAA 3450
CTTAATCGCC TTGCAGCACA TCCCCCCTTC GCCAGCTGGC GTAATAGCGA 3500
AGAGGCCCGC ACCGATCGCC CTTCCCAACA GTTGCGTAGC CTGAATGGCG 3550
45 AATGGCGCCT GATGCGGTAT TTTCTCCTTA CGCATCTGTG CGGTATTTC 3600
CACCGCATAC GTCAAAGCAA CCATAGTACG CGCCCTGTAG CGGCGCATT 3650
50 AGCGCGGCGG GTGTGGTGGT TACGCGCAGC GTGACCGCTA CACTTGCCAG 3700
CGCCCTAGCG CCCGCTCCTT TCGCTTTCTT CCCTTCCTTT CTCGCCACGT 3750
TCGCCGGCTT TCCCCGTCAA GCTCTAAATC GGGGGCTCCC TTTAGGGTTC 3800
60 CGATTTAGTG CTTTACGGCA CCTCGACCCC AAAAACTTG ATTTGGGTGA 3850
TGTTTCACGT AGTGGGCCAT CGCCCTGATA GACGGTTTTT CGCCCTTTGA 3900
65 CGTTGGAGTC CACGTTCTTT AATAGTGGAC TCTTGTTCCA AACTGGAACA 3950

ACACTCAACC CTATCTCGGG CTATTCTTTT GATTTATAAG GGATTTTGCC 4000
5 GATTTTCGGCC TATTGGTTAA AAAATGAGCT GATTTAACAA AAATTTAACG 4050
CGAATTTTAA CAAAATATTA ACGTTTACAA TTTTATGGTG CACTCTCAGT 4100
10 ACAATCTGCT CTGATGCCGC ATAGTTAAGC CAACTCCGCT ATCGCTACGT 4150
GACTGGGTCA TGGCTGCGCC CCGACACCCG CCAACACCCG CTGACGCGCC 4200
15 CTGACGGGCT TGTCTGCTCC CGGCATCCGC TTACAGACAA GCTGTGACCG 4250
TCTCCGGGAG CTGCATGTGT CAGAGGTTTT CACCGTCATC ACCGAAACGC 4300
20 GCGAGGCAGT ATTCTTGAAG ACGAAAGGGC CTCGTGATAC GCCTATTTTT 4350
ATAGGTTAAT GTCATGATAA TAATGGTTTC TTAGACGTCA GGTGGCACTT 4400
TTCGGGGAAA TGTGCGCGGA ACCCCTATTT GTTTATTTTT CTAAATACAT 4450
30 TCAAATATGT ATCCGCTCAT GAGACAATAA CCCTGATAAA TGCTTCAATA 4500
ATATTGAAAA AGGAAGAGTA TGAGTATTCA ACATTCCGT GTCGCCCTTA 4550
35 TTCCCTTTTT TCGGCATTT TGCCTTCCTG TTTTGGCTCA CCCAGAAACG 4600
CTGGTGAAAG TAAAAGATGC TGAAGATCAG TTGGGTGCAC GAGTGGGTTA 4650
CATCGAACTG GATCTCAACA GCGGTAAGAT CCTTGAGAGT TTTGCCCCG 4700
45 AAGAACGTTT TCCAATGATG AGCACTTTTA AAGTTCTGCT ATGTGGCGCG 4750
GTATTATCCC GTGATGACGC CGGGCAAGAG CAACTCGGTC GCCGCATACA 4800
50 CTATTCTCAG AATGACTTGG TTGAGTACTC ACCAGTCACA GAAAAGCATC 4850
TTACGGATGG CATGACAGTA AGAGAATTAT GCAGTGCTGC CATAACCATG 4900
AGTGATAACA CTGCGGCCAA CTTACTTCTG ACAACGATCG GAGGACCGAA 4950
60 GGAGCTAACC GCTTTTTTGC ACAACATGGG GGATCATGTA ACTCGCCTTG 5000
ATCGTTGGGA ACCGGAGCTG AATGAAGCCA TACCAAACGA CGAGCGTGAC 5050
65 ACCACGATGC CAGCAGCAAT GGCAACAACG TTGCGCAAAC TATTAAGTGG 5100

CGAACTACTT ACTCTAGCTT CCCGGCAACA ATTAATAGAC TGGATGGAGG 5150

5 CGGATAAAGT TGCAGGACCA CTTCTGCGCT CGGCCCTTCC GGCTGGCTGG 5200

TTTATTGCTG ATAAATCTGG AGCCGGTGAG CGTGGGTCTC GCGGTATCAT 5250

10 TGCAGCACTG GGGCCAGATG GTAAGCCCTC CCGTATCGTA GTTATCTACA 5300

CGACGGGGAG TCAGGCAACT ATGGATGAAC GAAATAGACA GATCGCTGAG 5350

15 ATAGGTGCCT CACTGATTAA GCATTGGTAA CTGTCAGACC AAGTTTACTC 5400

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20 AGGTGAAGAT CCTTTTTGAT AATCTCATGA CCAAATCCC TTAACGTGAG 5500

25 TTTTCGTTCC ACTGAGCGTC AGACCCCGTA GAAAAGATCA AAGGATCTTC 5550

TTGAGATCCT TTTTTTCTGC GCGTAATCTG CTGCTTGCAA ACAAAAAAAC 5600

30 CACCGCTACC AGCGGTGGTT TGTTTGCCGG ATCAAGAGCT ACCAACTCTT 5650

TTTCCGAAGG TAACTGGCTT CAGCAGAGCG CAGATACCAA ATACTGTCCT 5700

35 TCTAGTGTAG CCGTAGTTAG GCCACCACTT CAAGAACTCT GTAGCACCGC 5750

40 CTACATACCT CGCTCTGCTA ATCCTGTTAC CAGTGGCTGC TGCCAGTGGC 5800

GATAAGTCGT GTCTTACCGG GTTGGACTCA AGACGATAGT TACCGGATAA 5850

45 GGCGCAGCGG TCGGGCTGAA CGGGGGGTTC GTGCACACAG CCCAGCTTGG 5900

AGCGAACGAC CTACACCGAA CTGAGATACC TACAGCGTGA GCATTGAGAA 5950

50 AGCGCCACGC TTCCCGAAGG GAGAAAGGCG GACAGGTATC CGGTAAGCGG 6000

55 CAGGGTCGGA ACAGGAGAGC GCACGAGGGA GCTTCCAGGG GGAAACGCCT 6050

GGTATCTTTA TAGTCCTGTC GGGTTTCGCC ACCTCTGACT TGAGCGTCGA 6100

60 TTTTGTGAT GCTCGTCAGG GGGGCGGAGC CTATGGAAAA ACGCCAGCAA 6150

CGCGGCCTTT TTACGGTTCC TGGCCTTTTG CTGGCCTTTT GCTCACATGT 6200

65 TCCTTCCTGC GTTATCCCCT GATTCTGTGG ATAACCGTAT TACCGCCTTT 6250

GAGTGAGCTG ATACCGCTCG CCGCAGCCGA ACGACCGAGC GCAGCGAGTC 6300
AGTGAGCGAG GAAGCGGAAG AGCGCCCAAT ACGCAAACCG CCTCTCCCCG 6350
5 CGCGTTGGCC GATTCATTAA TCCAGCTGGC ACGACAGGTT TCCCGACTGG 6400
AAAGCGGGCA GTGAGCGCAA CGCAATTAAT GTGAGTTACC TCACTCATTAA 6450
10 GGCACCCAG GCTTTACACT TTATGCTTCC GGCTCGTATG TTGTGTGGAA 6500
15 TTGTGAGCGG ATAACAATTT CACACAGGAA ACAGCTATGA CCATGATTAC 6550
GAATTAA 6557
20

(2) INFORMATION FOR SEQ ID NO:4:

25 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 7305 bases
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

30 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

TTCGAGCTCG CCCGACATTG ATTATTGACT AGTTATTAAT AGTAATCAAT 50
35 TACGGGGTCA TTAGTTCATA GCCCATATAT GGAGTTCCGC GTTACATAAC 100
TTACGGTAAA TGGCCCGCCT GGCTGACCGC CCAACGACCC CCGCCCATTG 150
40 ACGTCAATAA TGACGTATGT TCCCATAGTA ACGCCAATAG GGACTTTCCA 200
45 TTGACGTCAA TGGGTGGAGT ATTTACGGTA AACTGCCCCAC TTGGCAGTAC 250
ATCAAGTGTA TCATATGCCA AGTACGCCCC CTATTGACGT CAATGACGGT 300
50 AAATGGCCCG CCTGGCATTG TGCCAGTAC ATGACCTTAT GGGACTTTCC 350
TACTTGGCAG TACATCTACG TATTAGTCAT CGCTATTACC ATGGTGATGC 400
GGTTTTGGCA GTACATCAAT GGGCGTGGAT AGCGGTTTGA CTCACGGGGA 450
60 TTTCCAAGTC TCCACCCCAT TGACGTCAAT GGGAGTTTGT TTTGGCACCA 500
AAATCAACGG GACTTTCCAA AATGTCGTAA CAACTCCGCC CCATTGACGC 550
65 AAATGGGCGG TAGGCGTGTA CGGTGGGAGG TCTATATAAG CAGAGCTCGT 600

TTAGTGAACC GTCAGATCGC CTGGAGACGC CATCCACGCT GTTTTGACCT 650
CCATAGAAGA CACCGGGACC GATCCAGCCT CCGCGGCCGG GAACGGTGCA 700
5 TTGGAACGCG GATTCCCCGT GCCAAGAGTG ACGTAAGTAC CGCCTATAGA 750
GTCTATAGGC CCACCCCCTT GGCTTCGTTA GAACGCGGCT ACAATTAATA 800
CATAACCTTA TGTATCATAC ACATACGATT TAGGTGACAC TATAGAATAA 850
15 CATCCACTTT GCCTTTCTCT CCACAGGTGT CCACTCCCAG GTCCAACTGC 900
ACCTCGGTTC TAAGCTTATC GATATGAAA AGCCTGAACT CACCGCGACG 950
20 TCTGTCGAGA AGTTTCTGAT CGAAAAGTTC GACAGCGTCT CCGACCTGAT 1000
GCAGCTCTCG GAGGGCGAAG AATCTCGTGC TTTCAGCTTC GATGTAGGAG 1050
GGCGTGGATA TGTCTGCGG GTAAATAGCT GCGCCGATGG TTTCTACAAA 1100
30 GATCGTTATG TTTATCGGCA CTTTGCATCG GCCGCGCTCC CGATTCCGGA 1150
AGTGCTTGAC ATTGGGGAAT TCAGCGAGAG CCTGACCTAT TGCATCTCCC 1200
35 GCCGTGCACA GGGTGTACG TTGCAACACC TGCCTGAAAC CGAACTGCCC 1250
GCTGTTCTGC AGCCGGTCGC GGAGGCCATG GATGCGATCG CTGCGGCCGA 1300
TCTTAGCCAG ACGAGCGGGT TCGGCCATT CGGACCGCAA GGAATCGGTC 1350
45 AATACACTAC ATGGCGTGAT TTCATATGCG CGATTGCTGA TCCCCATGTG 1400
TATCACTGGC AAAGTGTGAT GGACGACACC GTCAGTGCGT CCGTCGCGCA 1450
50 GGCTCTCGAT GAGCTGATGC TTTGGGCCGA GGAAGTCCGGC 1500
ACCTCGTGCA CGCGGATTTC GGCTCCAACA ATGTCCTGAC GGACAATGGC 1550
CGCATAACAG CGGTCATTGA CTGGAGCGAG GCGATGTTTC GGGATTCCCA 1600
60 ATACGAGGTC GCCAACATCT TCTTCTGGAG GCCGTGGTTG GCTTGTATGG 1650
AGCAGCAGAC GACTTTCGAG CGGAGGCATC CGGAGCTTGC AGGATCGCCG 1700
65 CGGCTCCGGG CGTATATGCT CCGCATTGGT CTTGACCAAC TCTATCAGAG 1750

CTTGGTTGAC GGCAATTTTCG ATGATGCAGC TTGGGCGCAG GGTCGATGCG 1800
5 ACGCAATCGT CCGATCCGGA GCCGGGACTG TCGGGCGTAC ACAAATCGCC 1850
CGCAGAAGCG CGGCCGTCTG GACCGATGGC TGTGTAGAAG TACTCGCCGA 1900
10 TAGTGGAAC CGACGCCCCA GCACTCGTCC GAGGGCAAAG GAATAGAGTA 1950
GATGCCGACC GAAGGATCCC CGGGGAATTC AATCGATGGC CGCCATGGCC 2000
15 CAACTTGTTT ATTGCAGCTT ATAATGGTTA CAAATAAAGC AATAGCATCA 2050
CAAATTTTAC AAATAAAGCA TTTTTCAC TGCATTCTAG TTGTGGTTTG 2100
20 TCCAACTCA TCAATGTATC TTATCATGTC TGGATCGATC GGGAATTAAT 2150
25 TCGGCGCAGC ACCATGGCCT GAAATAACCT CTGAAAGAGG AACTTGGTTA 2200
GGTACCTTCT GAGGCGGAAA GAACCAGCTG TGAATGTGT GTCAGTTAGG 2250
30 GTGTGGAAG TCCCAGGCT CCCAGCAGG CAGAAGTATG CAAAGCATGC 2300
ATCTCAATTA GTCAGCAACC AGGTGTGGAA AGTCCCCAGG CTCCCCAGCA 2350
35 GGCAGAAGTA TGCAAAGCAT GCATCTCAAT TAGTCAGCAA CCATAGTCCC 2400
40 GCCCCTAACT CCGCCCATCC CGCCCCTAAC TCCGCCAGT TCCGCCATT 2450
CTCCGCCCCA TGGCTGACTA ATTTTTTTTA TTTATGCAGA GGCCGAGGCC 2500
45 GCCTCGGCCT CTGAGCTATT CCAGAAGTAG TGAGGAGGCT TTTTGGAGG 2550
CCTAGGCTTT TGCAAAAAGC TAGCTTATCC GGCCGGGAAC GGTGCATTGG 2600
50 AACGCGGATT CCCCCTGCCA AGAGTCAGGT AAGTACCGCC TATAGAGTCT 2650
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GTCCCCGAGC TCCCTGTCCG CCTCTGTGGG CGATAGGGTC ACCATCACCT 2950
5 GCCGTGCCAG TCAGAGCGTC GATTACGATG GTGATAGCTA CATGAACTGG 3000
TATCAACAGA AACCAGGAAA AGCTCCGAAA CTACTGATTT ACGCGGCCTC 3050
10 GTACCTGGAG TCTGGAGTCC CTTCTCGCTT CTCTGGATCC GGTTCCTGGGA 3100
CGGATTTCAC TCTGACCATC AGCAGTCTGC AGCCGGAAGA CTTCGCAACT 3150
15 TATTACTGTC AGCAAAGTCA CGAGGATCCG TACACATTTG GACAGGGTAC 3200
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CGCCCTCCAA TCGGGTAACT CCCAGGAGAG TGTCACAGAG CAGGACAGCA 3400
30 AGGACAGCAC CTACAGCCTC AGCAGCACCC TGACGCTGAG CAAAGCAGAC 3450
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35 CTCGCCCCGTC ACAAAGAGCT TCAACAGGGG AGAGTGTTAA GCTTCGATGG 3550
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45 GTTGTGGTTT GTCCAAACTC ATCAATGTAT CTTATCATGT CTGGATCGAT 3700
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50 GAACTTGGTT AGGTACCTTC TGAGGCGGAA AGAACCAGCT GTGGAATGTG 3800
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5 TTTTTTGGAG GCCTAGGCTT TTGCAAAAAG CTGTTAACAG CTTGGCACTG 4150

GCCGTCGTTT TACAACGTCG TGA CTGGGAA AACCTGGCG TTACCCAACT 4200

10 TAATCGCCTT GCAGCACATC CCCCCTTCGC CAGCTGGCGT AATAGCGAAG 4250

AGGCCCCGAC CGATCGCCCT TCCCAACAGT TGCGTAGCCT GAATGGCGAA 4300

15 TGGCGCCTGA TCGGTATTT TCTCCTTACG CATCTGTGCG GTATTTCACA 4350

CCGCATACGT CAAAGCAACC ATAGTACGCG CCCTGTAGCG GCGCATTAAAG 4400

20 CGCGGCGGGT GTGGTGGTTA CGCGCAGCGT GACCGCTACA CTTGCCAGCG 4450

25 CCCTAGCGCC CGCTCCTTTC GCTTTCTTCC CTTCTTTTCT CGCCACGTTT 4500

GCCGGCTTTC CCCGTCAAGC TCTAAATCGG GGGCTCCCTT TAGGGTTCCG 4550

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5 ATTGAAAAAG GAAGAGTATG AGTATTCAAC ATTTCCGTGT CGCCCTTATT 5300

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10 GGTGAAAGTA AAAGATGCTG AAGATCAGTT GGGTGACGA GTGGGTTACA 5400

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15 GAACGTTTTT CAATGATGAG CACTTTTAAA GTTCTGCTAT GTGGCGCGGT 5500

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20 ATTCTCAGAA TGAATTGGT GAGTACTCAC CAGTCACAGA AAAGCATCTT 5600

25 ACGGATGGCA TGACAGTAAG AGAATTATGC AGTGCTGCCA TAACCATGAG 5650

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30 AGCTAACCGC TTTTTTGCAC AACATGGGGG ATCATGTAAC TCGCCTTGAT 5750

CGTTGGGAAC CGGAGCTGAA TGAAGCCATA CCAACGACG AGCGTGACAC 5800

35 CACGATGCCA GCAGCAATGG CAACAACGTT GCGCAAATA TTAAGTGGCG 5850

40 AACTACTTAC TCTAGCTTCC CGGCAACAAT TAATAGACTG GATGGAGGCG 5900

GATAAAGTTG CAGGACCACT TCTGCGCTCG GCCCTTCCGG CTGGCTGGTT 5950

45 TATTGCTGAT AAATCTGGAG CCGGTGAGCG TGGGTCTCGC GGTATCATTG 6000

CAGCACTGGG GCCAGATGGT AAGCCCTCCC GTATCGTAGT TATCTACACG 6050

50 ACGGGGAGTC AGGCAACTAT GGATGAACGA AATAGACAGA TCGCTGAGAT 6100

55 AGGTGCCTCA CTGATTAAGC ATTGGTAACT GTCAGACCAA GTTTACTCAT 6150

ATATACTTTA GATTGATTTA AACTTCATT TTTAATTTAA AAGGATCTAG 6200

60 GTGAAGATCC TTTTGTGATA TCTCATGACC AAAATCCCTT AACGTGAGTT 6250

TTCTGTTCCAC TGAGCGTCAG ACCCGTAGA AAAGATCAAA GGATCTTCTT 6300

65 GAGATCCTTT TTTTCTGCGC GTAATCTGCT GCTTGCAAAC AAAAAACCA 6350

CCGCTACCAG CGGTGGTTTG TTTGCCGGAT CAAGAGCTAC CAACTCTTTT 6400

5 TCCGAAGGTA ACTGGCTTCA GCAGAGCGCA GATACCAAAT ACTGTCCTTC 6450

TAGTGTAGCC GTAGTTAGGC CACCACTTCA AGAACTCTGT AGCACCGCCT 6500

10 ACATACCTCG CTCTGCTAAT CCTGTTACCA GTGGCTGCTG CCAGTGGCGA 6550

TAAGTCGTGT CTTACCGGGT TGGACTCAAG ACGATAGTTA CCGGATAAGG 6600

15 CGCAGCGGTC GGGCTGAACG GGGGGTTTCGT GCACACAGCC CAGCTTGGAG 6650

CGAACGACCT ACACCGAACT GAGATACCTA CAGCGTGAGC ATTGAGAAAG 6700

20 CGCCACGCTT CCCGAAGGGA GAAAGGCGGA CAGGTATCCG GTAAGCGGCA 6750

GGGTCGGAAC AGGAGAGCGC ACGAGGGAGC TTCCAGGGGG AAACGCCTGG 6800

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30 TTTGTGATGC TCGTCAGGGG GCGGAGCCT ATGGAAAAC GCCAGCAACG 6900

CGGCCTTTTT ACGGTTCCTG GCCTTTTGCT GGCCTTTTGC TCACATGTTT 6950

35 TTTCCTGCGT TATCCCCTGA TTCTGTGGAT AACCGTATTA CCGCCTTTGA 7000

GTGAGCTGAT ACCGCTCGCC GCAGCCGAAC GACCGAGCGC AGCGAGTCAG 7050

TGAGCGAGGA AGCGGAAGAG CGCCCAATAC GCAAACCGCC TCTCCCCGCG 7100

45 CGTTGGCCGA TTCATTAATC CAGCTGGCAC GACAGGTTTC CCGACTGGAA 7150

AGCGGGCAGT GAGCGCAACG CAATTAATGT GAGTTACCTC ACTCATTAGG 7200

50 CACCCCAGGC TTTACACTTT ATGCTTCCGG CTCGTATGTT GTGTGGAATT 7250

GTGAGCGGAT AACAAATTCA CACAGGAAAC AGCTATGACC ATGATTACGA 7300

55 ATTAA 7305

60

CLAIMS

1. A DNA construct comprising a transcriptional initiation site, a transcriptional termination site, a selectable gene, a product gene
5 provided 3' to the selectable gene, a transcriptional regulatory region regulating transcription of both the selectable gene and the product gene, the selectable gene being positioned within an intron having a splice donor site 5' of the intron, which splice donor site regulates expression of the product gene using the transcriptional
10 regulatory region.
2. The DNA construct of claim 1 wherein the splice donor site comprises an efficient splice donor sequence.
- 15 3. The DNA construct of claim 2 wherein the splice donor site comprises a consensus splice donor sequence.
4. The DNA construct of claim 2 wherein the splice donor site comprises the sequence GACGTAAGT.
- 20 5. The DNA construct of claim 1 wherein the selectable gene is an amplifiable gene.
6. The DNA construct of claim 5 wherein the amplifiable gene is DHFR.
- 25 7. The DNA construct of claim 1 wherein the transcriptional regulatory region comprises a promoter and an enhancer.
8. A vector comprising the DNA construct of claim 1.
- 30 9. The vector of claim 8 wherein the selectable gene of the DNA construct is an amplifiable gene.
10. The vector of claim 8 that is capable of replication in a eukaryotic
35 host.
11. A eukaryotic host cell comprising the vector of claim 10.
12. A eukaryotic host cell comprising the DNA construct of claim 5.
- 40 13. The host cell of claim 11 wherein the vector is introduced into the host cell by electroporation.
14. A eukaryotic host cell comprising the DNA construct of claim 1
45 integrated into a chromosome of the host cell.

15. The host cell of claim 14 that is a mammalian cell.
16. A method for producing a product of interest comprising culturing the
host cell of claim 11 so as to express the product gene and
5 recovering the product from the host cell culture.
17. The method of claim 16 further comprising recovering the product from
the culture medium.
- 10 18. The method of claim 16 wherein the selectable gene is an amplifiable
gene and the splice donor site comprises an efficient splice donor
sequence.
19. A method for producing a product of interest comprising culturing the
15 host cell of claim 12 so as to express the product gene in a
selective medium comprising an amplifying agent for sufficient time
to allow amplification to occur, and recovering the product.
20. A method for producing eukaryotic cells having multiple copies of a
20 product gene comprising transforming eukaryotic cells with the DNA
construct of claim 5, growing the cells in a selective medium
comprising an amplifying agent for a sufficient time for
amplification to occur, and selecting cells having multiple copies
of the product gene.
- 25 21. The method of claim 20 further comprising recovering from the
selected cells the product of interest.

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FIG. 1A

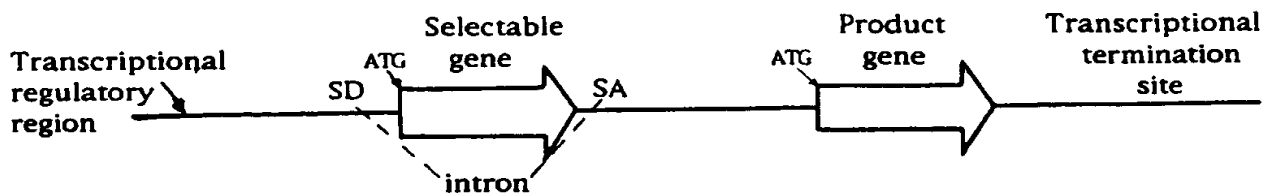


FIG. 1B

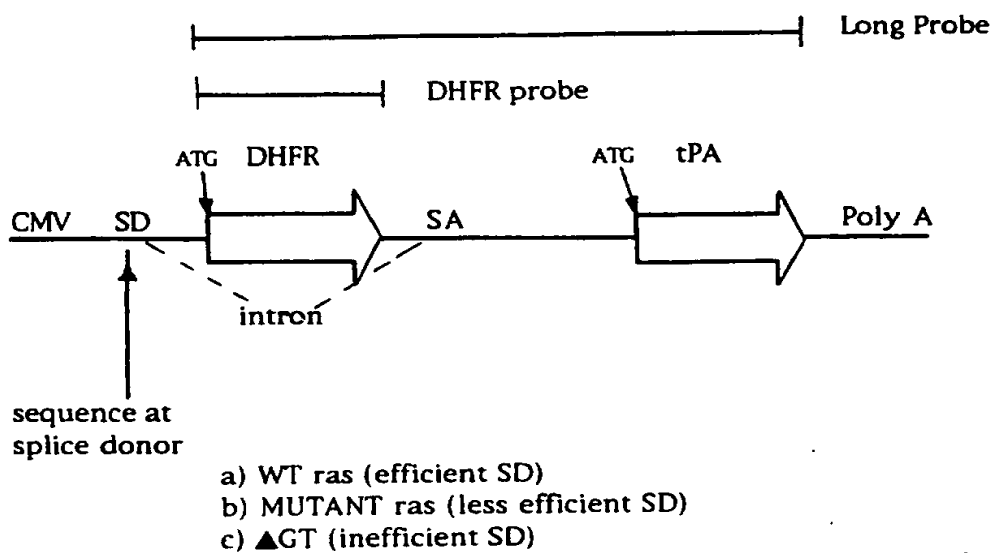
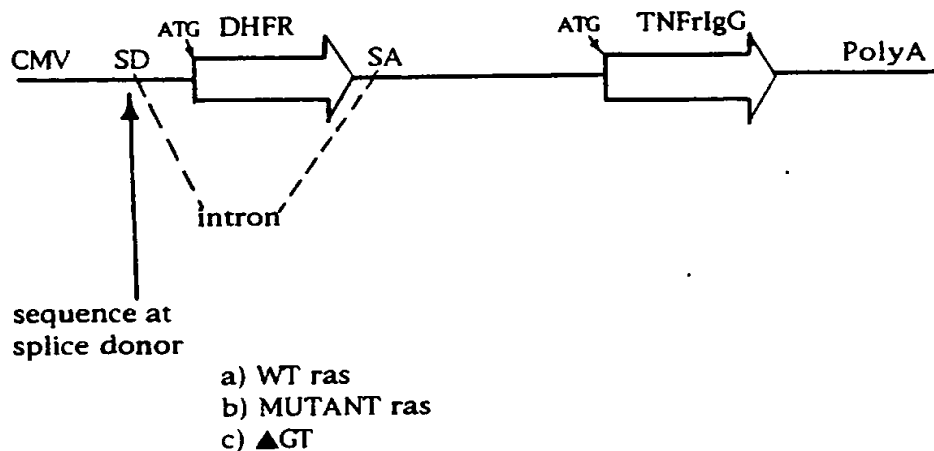


FIG. 1C



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FIG. 1D

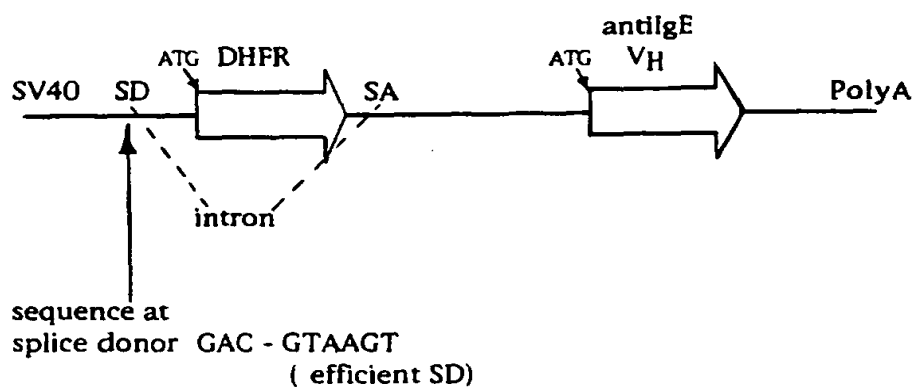
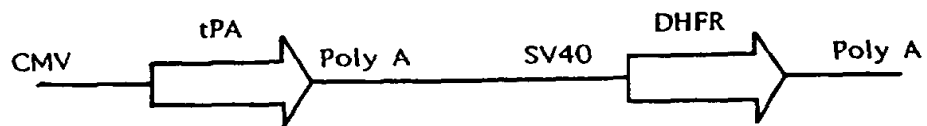


FIG. 2



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FIG. 3A

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1  TCGAGCTCG CCCGACATTG ATTATTGACT AGTTATTAAAT AGTAATCAAT TACGGGGTCA TTAGTTCATA GCCCATATAT GGAGTTCCGC GTTACATAAC
   AAGCTCGAGC GGCGTGTAAC TAATAACTGA TCAATAATTA TCATTAGTTA ATGCCCCAGT AATCAAGTAT CGGGTATATA CCTCAAGGCG CAATGTATTG
   taqI          rmaI   tru9I          bslI          aciI   maeIII
   maeI          maeI   asel/asnI/vspI
   speI          maeI   maeI          maeI          maeI          maeI
   acil          maeI          hinII/acyI          ahaII/bsaHI          maeIII
   bgli bstNI          sau96I          aciI          aciI          aatII          maeII          maeII
   haeIII/palI          asuI apyI[dcn+]          acil          aatII          maeII          maeII          maeII
   TCGGGTAA TGGCCGCCT GGCTGACCGC CCAACGACCC CCGCCCATTG ACGTCAATAA TGACGTATGT TCCCATAGTA ACGCCAATAG GGACTTTCCA
   AATGCCATT ACCGGCGGA CCGACTGGCG GGTGCTGGG GCGGGGTAAC TGCAGTTATT ACTGCATACA AGGTATCAT TCGGTTATC CCTGAAAGGT
   maeII          hinII/acyI          ahaII/bsaHI          aatII          maeII          maeII          maeII
   TCGAGTCAA TGGGTGGAGT ATTTACGGTA AACTGCCCCAC TTGGCAGTAC ATCAAGTGA TCATATGCCA AGTACGCCCC CTATTGACGT CAATGACGGT
   AACTGCAGT ACCCACCTCA TAAATGCCAT TTGACGGGTG AACCGTCAATG TAGTTCACAT AGTATACGGT TCATGCGGGG GATAACTGCA GTTACTGCCA
   scrFI          maeI          maeI          maeI          maeI          maeI          maeI
   maeI          maeI          maeI          maeI          maeI          maeI          maeI
   aciI          bgli          bgli          bgli          bgli          bgli          bgli
   bglI dsav          sau96I bstNI          haeIII/palI          bsrI nlaIII
   asuI apyI[dcn+]          TGGCCAGTAC TGCCCGATTA TGCCCGATTA TGCCCGATTA TGCCCGATTA TGCCCGATTA TGCCCGATTA
   TTTACCGGCG GGACCGTAAT ACGGGTCATG TACTGGATA CCCTGAAAGG ATGAACCGTC ATGTAGATGC ATAATCAGTA GCGATAATGG TACCACTACG

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FIG. 3B

[illegible]

FIG. 3C

tfII
 aciI
 thaI hinfI
 fnuDII/mvnI
 bstUI
 bsh1236I
 701 TTGGAACGGG GATTCCCGG GCCAAGAGTG CTGTAAGTAC CGCTATAGA GCGATAAGAG GATTTTATCC CGCTGCCAT CATGGTTGGA CCATTGAACT
 AACCTTGGC CTAAGGGCA CGGTTCTCAC GACATTCAATG CCGGATATCT CGGTATCTC CTAAATATAG GCGCAGGTA GTACCAAGCT GGTAACCTTGA
 fnu4HI
 bbvI
 nspBII
 aciI
 nlaIII
 taqI
 801 GCATCGTCCG CGTGTCCTCA AATATGGGA TTGGCAAGAA CGAGACCTA CCTGCGCCTC CGCTCAGAA CGGTTTCAAG TACTTCCAA GAATGACCAC
 CGTAGCAGG GCACAGGGT TTATACCCCT AACCGTTCTT GCCTCTGGAT GGGACGGAG CGGAGTCTT CCGCAAGTTC ATGAAGGTTT CTTACTGGTG
 pflMI
 bslI
 bsmAI
 bsaI
 bsrBI
 aflIII
 rsaI
 csp6I
 xmnI
 mnlI
 ddeI
 asp700
 scaI
 901 AACCTCTTCA GTGGAAGGTA AACAGAATCT GGTGATTATG GGTAGGAAA CCTGGTTCTC CATTCCTGAG AAGAATCGAC CTTTAAAGGA CAGAATTAAAT
 TTGGAGAAGT CACCTTCCAT TTGTCTTAGA CCACTAATAC CCATCCTTTT GGACCAAGAG GTAAGGACTC TTCTTAGCTG GAAATTTCTT GTCTTAATTA
 eco57I
 mboII
 earI/ksp632I
 mnlI
 tfII
 hinfI
 alwNI
 hphI
 bstNI
 apyI(dcm+)
 sexAI
 mboII
 taqI
 mseI
 tru9I
 mseI
 ahaIII/draI
 aseI/asnI/vspI
 1001 ATAGTTCTCA GTAGAGAACT CAAGAACCA CCACGAGGAG CTCATTTTCT TGCCAAAAGT TTGGATGATG CCTTAAGACT TATTGAACAA CCGGAATTGG
 TATCAAGAGT CATCTCTTGA GTTCTTGGT GGTGCTCCTC GAGTAAAGA ACGGTTTCA AACCTACTAC GGAATCTGA ATAACTTCTT GGCCTTAACC
 ddeI
 bslI
 mnlI
 bstXI
 fokI
 sfaNI
 mseI
 tru9I
 mspI
 hpaII
 bsaI

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FIG. 3D

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      haeIII/palI
      haeI
      nlaIII
      scrFI      mvaI      sau3AI
      mvaI      ecorII      mboI/ndeII(dam-)
      dsav      tfil      dsav      pleI      dpnI(dam+)
      bstNI      nlaIII      bstNI      ddeI      dpnII(dam-)
      apyI(dcm+)      hinfi      apyI(dcm+)      hinfi      maeIII      alwI(dam-)
      TGGTATAGTCG      GAGCGAGTTC      TGTATTACCAG      GAAGCCATGA      ATCAACCAGG      CCACCTTAGA      CTCTTTGTGA      CAAGGATCAT
      accI      nlaIII      mnlI      apyI(dcm+)      hinfi      apyI(dcm+)      hinfi      maeIII      alwI(dam-)
      TCTGTATTC      TCTGTATTC      TCTGTATTC      TCTGTATTC      TCTGTATTC      TCTGTATTC      TCTGTATTC      TCTGTATTC
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      GTTCATTCA TCTGTATTC ACCTATCAGC CTCCGTCAAG ACAATAGGTC CTTCGGTACT TACTGGTCC GGTTGAATCT GAGAACACT GTTCCTAGTA

      mnlI
      hinII/acyI      scrFI
      ahaII/bsaHI      mvaI
      scrFI      ecorII      dsav
      mvaI      ecoNI      dsav
      ecorII      sau96I
      dsav      avari
      bstNI      bslI      asuI      mnlI
      apyI(dcm+)      mnlI      bstNI
      bsaJI      hgaI      ddeI      apyI(dcm+)
      CCAGGAAAT      ATAAACCTCT      CCAGGAAAT      CCAGGAAAT      CCAGGAAAT      CCAGGAAAT      CCAGGAAAT      CCAGGAAAT
      1201 GCAGGAAAT GAAAGTGACA CGTTTTCCTC AGAATGAT      TTGGGAAAT      ATAAACCTCT      CCAGGAAAT      CCAGGAAAT      CCAGGAAAT      CCAGGAAAT
      CGTCCCTAAA CTTTCACTGT GCAAAAAGGG TCTTTAACTA AACCCCTTA TATTGGAGA GGTCTTATG GGTCGCGAGG AGAGACTCCA GGTCTCTCTT

      mnlI
      apoI      maeIII
      afluII
      maeIII
      sfanI      accI      mboII
      mboII
      scrFI      mvaI      sau3AI
      mvaI      ecorII      mboI/ndeII(dam-)
      dsav      tfil      dsav      pleI      dpnI(dam+)
      bstNI      nlaIII      bstNI      ddeI      dpnII(dam-)
      apyI(dcm+)      hinfi      apyI(dcm+)      hinfi      maeIII      alwI(dam-)
      TGGTATAGTCG      GAGCGAGTTC      TGTATTACCAG      GAAGCCATGA      ATCAACCAGG      CCACCTTAGA      CTCTTTGTGA      CAAGGATCAT
      1301 AAAGGCATCA AGTATAAGTT TGAAGTCTAC GAGAGAAAG ACTAACAGGA AGATGCTTC AAGTCTCTG CTCCCTCTC AAAGCTATGC ATTTTATAA
      TTCCCGTAGT TCATATTCAA ACTTCAGATG CTCTTCTTTC TGATTGTCTC TCTACGAAAG TTCAAGAGAC GAGGGAGGA TTTCGATACG TAAAAATATT

      fnu4HI
      aciI
      thal
      fnuDII/mvni      tru9I
      bstUI      msel
      bsh1236I      asei/asnl/vspl
      styI      bsaJI
      bsaJI
      CTTTGGCTGG CTTTAGACCC CTTTGGCTGG CTTTAGACCC CTTTGGCTGG CTTTAGACCC CTTTGGCTGG CTTTAGACCC
      1401 GACCATGGGA CTTTGGCTGG CTTTAGACCC CTTTGGCTGG CTTTAGACCC CTTTGGCTGG CTTTAGACCC CTTTAGACCC
      CTGGTACCCCT GAAACGACC GAATCTGGG GGAACCGAAG CAATCTTGG CCGATGTAA TTAGTATTG GAATACATAG TATGTATC TAAATCCACT

```


[illegible]

FIG. 3F

FIG. 3F

[illegible][illegible]

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FIG. 3H

[illegible]

FIG. 31

[illegible]

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FIG. 3K

```

          styI
        acII
      fnu4HI   sau96I
      bgII nlaIII
      sfII ncoI haeIII/palI
      haeIII/palI
      eaeI dsalI asuI
      cfrI bsaJI
3601 GATGGCGCC ATGCCCCAAC TTGTTTATTG CAGCTTATAA TGGTTACAAA TAAAGCAATA TTTACACAAA TTTACACAAAT AAAGCAATTT TTTCACTGCA
      CTACCGCGCG TACCGGGTTG AACAAATAAC GTCGAATATT ACCAATGTTT ATTCGTGTTT CGTAGTGTAA AAAGTGTTTA TTTCTGTAATA AAAGTGACGT
          aluI
          fnu4HI
          bbvI
          maeIII
          sfaNI apoI
          bsmI
3601 GATGGCGCC ATGCCCCAAC TTGTTTATTG CAGCTTATAA TGGTTACAAA TAAAGCAATA TTTACACAAA TTTACACAAAT AAAGCAATTT TTTCACTGCA
      CTACCGCGCG TACCGGGTTG AACAAATAAC GTCGAATATT ACCAATGTTT ATTCGTGTTT CGTAGTGTAA AAAGTGTTTA TTTCTGTAATA AAAGTGACGT
          aluI
          fnu4HI
          bbvI
          maeIII
          sfaNI apoI
          bsmI
          sau3AI
          mboI/ndeII[dam-]
          dpnI[dam+]
          dpnII[dam-]
          pvuI/bspCI
          mcrI
          taqI[dam-] tru9I
          clai/bsp106[dam-]
          sau3AI mseI
          mboI/ndeII[dam-]
          dpnI[dam+] xmnI
          dpnII[dam-]
          dpnII[dam-] aseI/asnI/vspI bsaJI
          nlaIII alwI[dam-] asp700 hhaI/cfoI nlaIII
          mnlI
3701 TTCTAGTTGT GGTTTGTCCA AACTCATCAA TGTATCTTAT CATGTCTGGA TCGATCGGGA ATTAATTGGG CGCAGCACCA TGGCCTGAAA TAACCTCTGA
      AAGATCAACA CCAACACAGT TTGAGTAGTT ACATAGAATA GTACAGACCT AGCTAGCCCT TAATTAAGCC GCGTCGTGGT ACCGGACTTT ATTGGAGACT
          rmaI
          maeI
          nlaIII
          mnlI
          rsal
          csp6I
          nlaIV
          kpnI
          hgiCI
          banI
          asp718 mnlI
          acc65I ddeI aciI
          mnlI
          nlaIV
          csp6I
          mvaI
          ecoRII
          dsav
          bstNI
          apyI[dcm+]
          bsaJI
3801 AAGAGGAACT TGGTTAGGTA CCTTCTGAGG CGGAAAGAAC CAGCTGTGGA ATGTGTGTCA GTTAGGTGT GGAAGTCCC CAGGCTCCCC AGCAGGCAGA
      TTCTCCTTGA ACCAATCCAT GGAAGACTCC GCCTTTCTTG GTCGACACCT TACACACAGT CAATCCACA CTTTTCAGG GTCCGAGGG TCGTCCGTCT
          aluI
          pvuII
          nspBII
          mnlI
          nlaIV
          csp6I
          mvaI
          ecoRII
          dsav
          bstNI
          apyI[dcm+]
          bsaJI

```

SUBSTITUTE SHEET (RULE 26)

FIG. 3L

```
3901 AGTATGCAAA GATGCATCT CAATTAGTCA GCAACCAGGT GTGGAAGTC CCCAGGCTCC CCAGCAGGCA GAAGTATGCA AAGCATGCAT CTCATATTAGT
TCATACGTTT CGTACGTAGA GTTAATCAGT CGTTGGTCCA CACCTTTTCAG GGGTCCGAGG GGTGCTCCGT CTTCATACGT TTCGTACGTA GAGTTAATCA

4001 CAGCAACCAT AGTCCCGCCC CTAACCTCCG CCATCCCGCC CCTAACTCCG CCAGTTCGG CCCATTCTCC GCCCATGGC TGAATAATTT TTTTATTATTA
GTCGTTGGTA TCAGGGCGGG GATTGAGCG GATTGAGCG GGTAGGGCGG GGATCAAGGC GGGTCAAGGC GGGGTACCG CGGGTACCG ACTGATTAAA AAAAATAAAT

4101 TGCAGAGGCC GAGGCGCCT CGGCTCTGA GCTATTCCAG AAGTAGTGAG GAGGCTTTT TGGAGGCTA GGCTTTTGA AAAAGCTGT AACAGCTTGG
ACGCTCCGG CTCCGGCGA GCGGAGACT CGATAAGTC TCATCACTC CTCCGAAAA ACCTCCGAT CCGAAAACT TTTTCGACA TTGTCGAACC

4201 CACTGGCCGT GGTTTTACAA CGTCGTGACT GGGAAAAACC TGGCGTTACC CAACTTAATC GCCTGCAGC ACATCCCCC TTCGCCAGCT GCGCTAATAG
GTGACCGGCA GCAAAATGTT GCAGCACTGA CCTTTTGGG ACCGCAATGG GTTGAATTAG CGGAACGTGC GTTAGGGGGG AAGCGGTGCA CCGCATTATC
```


BNSDOCID: <WO__9604391A1_1_>

FIG. 3N

[illegible]

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FIG. 30

```

nlaIV
aciI
thai
fnuDII/mvnI
bstUI
bsh1236I
hinPI
hhaI/cfoI

5201 CTTTTCGGG AAATGTGCGC GGAACCCCTA TTTGTTTATT TTTCTAAATA CATTCAAATA TGTATCCGCT CATGAGACAA TAACCTGTAT AAATGTTCA
GAAAAGCCCC TTTACAGCGG CCTTGGGAT AAACAAATAA AAAGATTTAT GTAAAGTTTAT ACATAGCGGA GTACTCTGTT ATTGGGACTA TTTACGAAGT

        rcaI
        bspHI
        bsrBI bsmAI
        aciI nlaIII
        fnu4HI
        aciI
        hphI

5301 ATAATATTGA AAAAGGAAGA GTATGAGTAT TCAACATTC CGTGCGCC TTATTCCCTT TTTGCGGCA TTTTGCTTC CTGTTTTTGC TCACCCAGAA
TATTATAACT TTTTCTTCT CATACTCATA AGTTGTAAAG GCACAGCGG AATAAGGAA AAAACGCCGT AAAACGGAAG GACAAAAACG AGTGGGTCTT

        hgiAI/aspHI
        bsp1286
        sau3AI bsiHKAI
        mboI/ndeII[dam-]
        dpnI[dam+] bmyI
        dpnII[dam-]
        eco57I apaLI/snoI
        hphI sfaNI mboII[dam-] alw44I/snoI maeIII taqI alwI[dam-] aciI bstyI/xhoII

5401 ACCTGGTGA AAGTAAAGA TGCTGAAGT CAGTTGGTG CACGAGTGG TTACATCGAA CTGGATCTCA ACACGGTAA GATCCTTGAG AGTTTTGCGC
TGCGACCACT TTCACTTCT ACGACTTCTA GTCAACCCAC GTGCTCACCC AATGTAGCTT GACCTAGAGT TGTCGCCATT CTAGGAATC TCAAAAGCGG

        scrFI
        nciI
        mspI
        hpaII
        dsav
        hinII/acyI
        hgaI cauII
        ahaII/bsaHI bciI mcrI fnu4HI
        aciI

5501 CCGAAGAACG TTTTCCAATG ATGAGCACTT TTAAGTTCT GCTATGTGGC GCGGTATTAT CCGTGATGA CGCGGGCAA GAGCAACTCG GTCGCGCAT
GGCTTCTTGC AAAAGGTTAC TACTCGTGAA AATTTCAAGA CGATACACCG CGCCATAATA GGGCACTACT CGCGCCCGTT CTCGTTGAGC CAGCGGCGTA

        rsaI
        csp6I bsrI
        scaI hphI maeIII sfanI foki nlaIII
        ddeI

5601 ACACTATTCT CAGAAAGACT TGGTTGAGTA CTCACCAGTC ACAGAAAAGC ATCTTACGGA TGGCATGACA GTAAGAGAAT TATGCACTGC TGCCATAACC
TGCGATAAGA GTCTTACTGA ACCAACTCAT GAGTGGTCAG TGCTTTTTCG TAGAATGCTT ACCGTACTGT CATCTCTTA ATACGTCACG ACGGTATTGG
        fnu4HI
        bbvI
        nlaIII

```

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FIG. 3P

```

sau96I
avaII
sau3AI asuI
mboI/ndeII(dam-)
dpmI(dam+)
dpmII(dam-)
pvuI/bspCI
mcrI mnlI
aluI aciI
nlaIII maeIII
sau3AI maeIII
mboI/ndeII(dam-)
dpmI(dam+)
dpmII(dam-)
nlaIII alwI(dam-)
5701 ATGAGTGATA ACACGCGG CAACCTACTT CTGACACAGA TCGGAGGACC GAAGGAGCTA ACCGCTTTT TGCACAACAT GGGGATCAT GTAACTCGCC
TACTCACTAT TGTGAGCCG GTTGAATGAA GACTGTGCT AGCCTCCTGG CTTCCTCGAT TGGCGAATAA ACGTCTGTA CCCCTAGTA CATTGAGCGG
haeIII/palI
eaeI
cfrI
fnu4HI
aciI
mcrI mnlI
aluI aciI
nlaIII maeIII
sau3AI maeIII
mboI/ndeII(dam-)
dpmI(dam+)
dpmII(dam-)
nlaIII alwI(dam-)
5801 TTGATCGTTG GGAACCGGAG CTGAATGAAG CCATACAAA CGACGAGCGT GACACACAGA TGCCAGCAGC AATGGCAACA ACCTGCGCA AACTATTAC
AACTAGCAAC CCTTGCCCTC GACTTACTTC GGTATGTTT GCTGCTCGCA CTGTGTGCT ACCTGCTGCTG TACCGTTGT TGAACGCGT TTGATAATTG
hinPI
hhaI/cfoI
mstI
aviII/fspI
maeII
psp1406I
tru9I
mseI
fnu4HI
sfaNI
bbvI
sau96I
haeIII/palI
hinPI asuI mspI
hhaI/cfoI hpalI
5901 TGGCGAACTA CTTACTCTAG CTTCCCGGCA ACAATTAATA GACTGGATGG AGCGGATAA AGTTGCAGGA CCACCTCTGC GCTCGCCCT TCCGGCTGGC
ACCGCTTGAT GAATGAGATC GAAGGCGCGT TGTTAATTAT CTGACCTACC TCCGCTATT TCAACGCTCT GGTGAAGACG CGAGCCGGA AGGCCGACCG
bgII
sau96I
haeIII/palI
hinPI asuI mspI
hhaI/cfoI hpalI
6001 TGGTTTATTG CTGATAAATC TGGAGCGGT GAGCGTGGT CTGCGGTAT CATTGCAGCA CTGGGCGCAG ATGGTAAGCC CTCCCGTATC GTAGTTATCT
ACCAATAAC GACTATTAG ACCTCGCCA CTGCGACCCA GAGCGCCATA GTACGCTGT GACCCCGTC TACCATTCCG GAGGCGATAG CATCAATAGA
mspI
hpaII
cfr10I
nlaIV hphI
gsuI/bpmI
thaI
fnuDII/mvni
bstuI
sau96I
haeIII/palI
nlaIV
bsaI bsh1236I
bbvI bsrI asuI
mnlI
6101 TGGTCTGCC CTGAGTCGGT TGATACCTAC TTGCTTATC TGTCTAGCA CTCTATCCAC GGAGTACTA ATTGTAACC ATTGACAGTC TGGTTCAAT
pleI
hinfi
eam1105I
fokI
nlaIV
sau3AI
mboI/ndeII(dam-)
dpmI(dam+)
dpmII(dam-)
hgiCI
tru9I
mseI
maeIII

```

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FIG. 3Q

```

        rnaI      sau3AI
sau3AI hphI mboI/ndeII[dam-]
mboI/ndeII[dam-]
dpmI[dam+] dpmI[dam+]
dpmII[dam-] dpmII[dam-]
        tru9I bstYI/xhoII alwI[dam-] nlaIII maeII
        mseI      tru9I mseI alwI[dam-] bstYI/xhoII rcaI tru9I
        ahaIII/draI mseI ahaIII/draI maeI mboII[dam-] bspHI mseI
6201 CTCATATATA CTTTAGATTG ATTTAAAGGA TCTAGGTGAA GATCCTTTTT GATAATCTCA TGACCAAAAT CCCTTAACGT
GAGTATATAT GAAATCTAAC TAAATTTTGA AGTAAATAATT AAATTTTCCT AGATCCACTT CTAGGAAAAA CTATTAGAGT ACTGGTTTTA GGGAAATTGA

        sau3AI
mboI/ndeII[dam-]
dpmI[dam+] sau3AI
dpmII[dam-] mboI/ndeII[dam-] thaI fnuDII/mvnI
        bstYI/xhoII dpmI[dam+] bstUI
sau3AI alwI[dam-] dpmII[dam-] bsh1236I
mboI/ndeII[dam-] alwI[dam-] hinPI fnu4HI
dpmI[dam+] mboII[dam-] bstYI/xhoII hhaI/cfoI bbvI
dpmII[dam-]
        sau3AI
mboI/ndeII[dam-]
dpmI[dam+]
dpmII[dam-]
alwI[dam-]
        aciI      nspBII      hpaII      aluI      bsrI      eco57I      hhaI/cfoI      hinPI
6401 AACACCGCT ACCAGCGGTG GTTTGTTTGC CGGATCAAGA GCTACCAACT CTTTTTCCGA AGTAACTGG CTTACAGCAGA GCGCAGATAC CAAATACTGT
TTGGTGGCGA TGGTCGCCAC CAAACAACG GCCTAGTTCT CGATGGTTGA GAAAAAGGCT TCCATTGACC GAAATCGTCT CGCCTCTATG GTTTATGACA

        rnaI      haeIII/palI
        maeI      haeI      bslI      haeI
6501 CTTTCTAGTG TAGCCGAGT TAGGCCACCA CTTCAAGAAC TCTGTAGCAC CGCCTACATA CCTCGCTCTG CTAATCTGT TACCAGTGGC TGCTGCCAGT
GGAAGATCAC ATCGCATCA ATCCGGTGTG GAAGTTCTTG AGACATCGTG GCGGATGTAT GGAGCGAGAC GATTAGGACA ATGGTCACCG ACGACGGTCA

```

BNSDOCID: <WO__9604391A1_1_>

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FIG. 3S

```

          thal
          fnuDII/mvnI
          bstUI
          bsh1236I
          hinPI
          hhAI/cfoI
          thal
          fnuDII/mvnI
          bstUI haeIII/palI aluI
          bsh1236I tru9I pvuII
          bslI eaeI tfil ael/asnI/vspI
          aciI cfrI hinfi msel nspBII bsrI
          7101 GTCAGTGAGC GAGGAAGCGG AAGAGCGCCC AATACGCAA CCGCCTCTCC CCGCGCGTGG GCCGATTTCAT TAATCCAGCT GGCACGACAG GTTCCCGAC
          CAGTCACTCG CTCCTTCGCC TTCTCGCGGG TTATGCGTTT GCGCGGAGAGG GCGCGGCAAC CGGCTAAGTA ATTAGGTGCA CCGTGTCTGTC CAAAGGGCTG
          sapI hinPI
          mboII hhAI/cfoI
          earI/ksp632I
          mnlI aciI haeII
          7201 TGGAAAGCGG GCAGTGAGCG CAACGCAATT AATGTGAGTT ACCTCACTCA TTACACTCAA TGGAGTGAGT AATCCGTGGG GTCCGAAATG TGAATAACGA AGGCCGAGCA TACAACACAC
          ACCTTTTCGCC CGTCACTCGC GTTGGGTTAA TTACACTCAA TGGAGTGAGT AATCCGTGGG GTCCGAAATG TGAATAACGA AGGCCGAGCA TACAACACAC
          hinPI msel tru9I
          hhAI/cfoI ael/asnI/vspI mnlI
          aciI bsrBI
          7301 GAATTGTGAG CGGATAACAA TTTCACACAG GAAACAGCTA TGACCATGAT TACGAATTAA
          CTTAACACTC GCCTATTGTT AAAGTGTC CTTTGTCGAT ACTGGTACTA ATGCTTAATT
          aciI
          bsrBI
          7301 GAATTGTGAG CGGATAACAA TTTCACACAG GAAACAGCTA TGACCATGAT TACGAATTAA
          CTTAACACTC GCCTATTGTT AAAGTGTC CTTTGTCGAT ACTGGTACTA ATGCTTAATT
          tru9I
          msel
          ael/asnI/vspI
          xmnI
          asp700
          7301 GAATTGTGAG CGGATAACAA TTTCACACAG GAAACAGCTA TGACCATGAT TACGAATTAA
          CTTAACACTC GCCTATTGTT AAAGTGTC CTTTGTCGAT ACTGGTACTA ATGCTTAATT

```

>length: 7360

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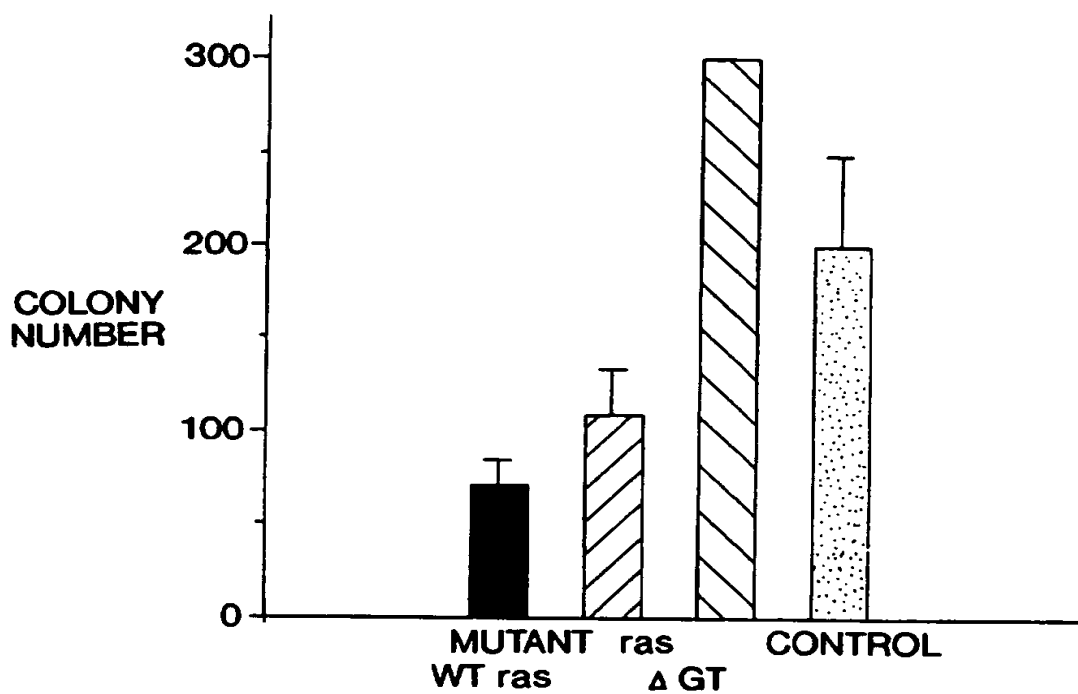


FIG. 4

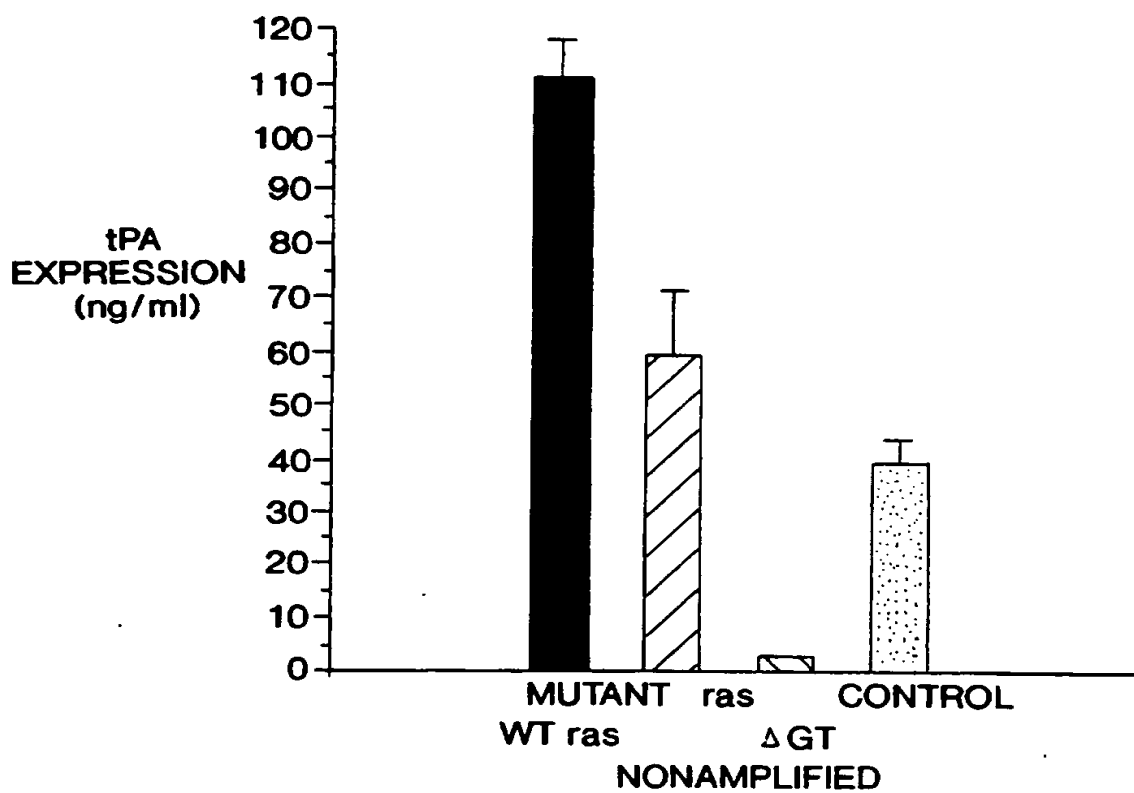


FIG. 5A

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FIG. 10A

```

1  TTCCAGCTCG CCCGACATG ATTATTGACT AGTTATTAAAT AGTAATCAAT TACGGGGTCA TTAGTTTCATA GCCCATATAT GGAGTTCCGC GTTACATAAC
   AAGCTGAGC GGGCTGTAAC TAATAACTGA TCAATAATTA TCATTAGTTA ATGCCCCAGT AATCAAGTAT CGGTATATA CCTCAAGCGG CAATGTATTG
   tagl          rmaI   tru9I          bslI          aciI maeIII
   maeI   mseI
   speI   aseI/asnI/vspI
201 TTGACGTCAA TGGGTGGAGT ATTTACGGTA AACTGCCCCAC TTGGCAGTAC ATCAAGTGA TCATATGCCA AGTACGCCCC CTATTGACGT CAATGACGGT
   AACTGCAGT ACCCACCTCA TAAATGCCAT TTGACGGGTG AACCGTCATG TAGTTCACAT AGTATACGCT TCATGCGGGG GATAACTGCA GTTACTGCCA
   maeII          hinII/acyI          maeII          hinII/acyI          ahaII/bsaHI          aatII
   ahaII/bsaHI          bglI          csp6I          ndeI          csp6I          aatII
201 TTGACGTCAA TGGGTGGAGT ATTTACGGTA AACTGCCCCAC TTGGCAGTAC ATCAAGTGA TCATATGCCA AGTACGCCCC CTATTGACGT CAATGACGGT
   AACTGCAGT ACCCACCTCA TAAATGCCAT TTGACGGGTG AACCGTCATG TAGTTCACAT AGTATACGCT TCATGCGGGG GATAACTGCA GTTACTGCCA
   maeII          hinII/acyI          maeII          hinII/acyI          ahaII/bsaHI          aatII
   ahaII/bsaHI          bglI          csp6I          ndeI          csp6I          aatII
301 AAATGGCCCG CCTGGCATTA TGCCGAGTAC ATGACCTTAT GGGACTTTCC TACTTGGCAG TACATCTACG TATTAGTCAT CGCTATTACC ATGGTGATGC
   TTTACCGGGC GGACCGTAAT ACGGTGCATG TACTTGAATA CCCTGAAAGG ATGAACCGTC ATGTAGATGC ATAATCAGTA GCGATAATGG TACCACCTAGC
   scrFI          mvaI          ecoRII          nlaIII
   aciI          bglI dsav          ncoI          dsal hphI aciI
   sau96I bstNI          maeII          snbI          bsaJI sfanI
   haeIII/palI          rsaI          csp6I          bsaI          csp6I          bsaI
   asuI apyI(dcm+)          bsrI nlaIII          csp6I          bsaI          csp6I          bsaI

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FIG. 9R

```

        thal
        fnuDII/mvnI
        bstUI
        bsh1236I
        hinPI
        hhai/cfoI
        thal
        fnuDII/mvnI
        bstUI
        bsh1236I haeIII/palI
        bsII eaeI tfII aseI/asnI/vspI
        acII cfrI hinI mseI nspBII
        6301 AGTGAGCGAG GAAGCGGAAG AGCGCCCAAT ACGCAAAACCG CCTCTCCCG CGGCTGGCC GATTCAATAA TCCAGCTGGC AGCAGAGGTT TCCCGACTGG
        TCACTCGCTC CTTCGCCCTC TCGCGGTTA TCGGTTTGGC GGAGAGGGGC GCGCAACCGG CTAAGTAATT AGGTGACCG TGCTGTCCAA AGGGCTGACC
        bsrI
        sapI hinPI
        mboII hhai/cfoI
        earI/ksp632I
        mnII acII haeII
        6401 AAAGCGGCA GTGAGCGCAA CGCAATTAAT GTGAGTTACC TCACTCAATA GGCACCCAG GCTTTACACT TTATGCTTCC GGTGCTATG TTGTGTGAA
        TTTGCCCCGT CACTCGCGT GCGTTAATTA CACTCAATGG AGTGAGTAAT CCGTGGGTC CGAAATGTA AATACGAAGG CCGAGCATAC AACACACCTT
        bsrI
        acII
        hinPI
        hhai/cfoI aseI/asnI/vspI
        tru9I mseI
        nlaIV bstNI
        hgiCI apyI[dcn+]
        banI bsaJI
        6501 TTGTGAGCGG ATAACAATTT CACACAGGAA ACAGCTATGA CCATGATTAC GAATTAA
        AACACTCGCC TATTGTTAAA GTGTGCTCTT TGTCGATACT GGTACTAATG CTTAATT
        acII
        bsrBI
        aluI
        nlaIII
        asp700
        xnnI
        aseI/asnI/vspI
        tru9I
        mseI
        6557

```

>length: 6557

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FIG. 9Q

```

                    hinPI          hhaI/cfoI          hhaII          aciI          fnu4HI
                    mspI          hpaII          bslI          bsaWI          aciI
5901 AGCGAAGCAGC CTACACCGAA CTGAGATACC TACAGGCTGA GCATTGAGAA AGCGCCACGC TTCCCGAAGG GAGAAGGCG GACAGGTATC CGGTAAGCGG
TCGCTTGCTG GATGTGGCTT GACTCTATGG ATGTGCACT CGTAACTCTT TCGCGGTGGG AAGGCTTCC CTCTTTCCGC CTGTCCATAG GCCATTCCGC

                    scrFI          mvaI          ecorII          mvaI          ecorII          dsaV          bstNI          dsaV
                    hinPI          mnlI          hhaI/cfoI          aluI          apyI[dcM+]          apyI[dcM+]          bstNI
6001 CAGGTCGGA ACAGGAGAGC GCACGAGGA GCTTCCAGG GGAACGCCT GGTATCTTTA TAGTCTGTC GGGTTTCGCC ACCTTGACT TGAGCGTCA
GTCCAGCCT TGTCTCTCG CGTCTCCCT CGAGGTCCC CTTTGCGGA CCATAGAAAT ATCAGGACAG CCCAAGCGG TGGAGACTGA ACTCGCAGCT

                    nlaIV          aciI          nlaIII          nspI          haeIII/palI          haeI          aflIII
                    sfaNI          nlaIV          aciI          nspI          haeIII/palI          haeI          aflIII
6101 TTTTGTGAT GCTGTCAGG GGGCGGAGC CTATCGAAA ACAGCAGCAA CGCGGCTTT TACGGTTCC TGGCCTTTTG CTGGCCTTTT GCTCAGATGT
AAAAACACTA CGAGCAGTCC CCCCCTCTCG GATACCTTTT TGGGCTCGTT GCGCCGAAA AATGCCAAGG ACCGGAAC GACCGAANA CGAGTGATCA

                    tfiI          hinFI          fnu4HI          bbsI          pleI          hinPI          hhaI/cfoI
                    bsrBI          aciI          fnu4HI          mcrI          hhaI/cfoI
6201 TCTTCTGCTG GTTATCCCCT GATTCGTGG ATAACCGTAT TACCGCTTT GAGTGAGCTG ATACCGCTCG CCGAGCGCA ACAGCGAGC GCAGCGAGTC
AGAAAGGACG CAATAGGGA CTAAGACACC TATTGGCATA ATGGCGAAA CTCACCTGAC TATGGCGAGC GCGCTCGCT TGCTGGCTCG CGTGGCTCAG

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FIG. 9P

```

sau3AI
mboI/ndeII[dam-]
dpnI[dam+] sau3AI          thAI
dpnII[dam-] mboI/ndeII[dam-]
bstYI/xhoII dpnI[dam+] fnuDII/mvnI
sau3AI alwI[dam-] dpnII[dam-] bstUI
mboI/ndeII[dam-] alwI[dam-] bsh1236I
dpnI[dam+] mboII[dam-] hinPI fnu4HI
dpnII[dam-] bstYI/xhoII hhaI/cfoI bbvI
5501 TTTTCGTTCC ACTGAGCGTC AGACCCCGTA GAAAGATCTTC TTGAGATCCT TTTTCTCTGC GCGTAATCTG CTGCTTGCAA ACAAAAAAAC
AAAAGCAAGG TGACTCGCAG TCTGGGGCAT CTTTCTCTAGT TTCCTAGAAG AACTCTAGGA AAAAAGAGAG CCGATTAGAC GACGAACGTT TGTTTTITG

sau3AI
mboI/ndeII[dam-]
dpnI[dam+]
dpnII[dam-]
alwI[dam-]
acII
nspBII
bsII haeI
maeI
hmaI
bsII haeI
haeIII/palI
scrFI
ncII
mspI
hpaII
dsav
cauII
pleI
hinfi
cauII
hinfI
pleI
bsaWI
hpaII
mspI
nspBII
fnu4HI
bbvI
hinPI mcrI
hhaI/cfoI
5601 CACCGCTACC AGCGGTGTTT TGTTCGCCG ATCAAGAGCT ACCAACTCTT TTTCCGAAGG TAACGTGCTT CAGCAGAGCG CAGATACCAA ATACTGTCTCT
GTGGCGATGG TCGCCACCAA ACAACGGCC TAGTCTCTGA TGGTTGAGAA AAAGGCTTCC ATTGACCGAA GTCTCTCTGC GTCTATGCTT TATGACAGGA

maeIII eco57I hhaI/cfoI
bsrI hinPI
bsrI fnu4HI
bbvI
fnu4HI
5701 TCTAGTTAG CCGTAGTTAG GCCACCACTT CAAGAATCTT GTAGCACCGC CTACATACCT CGCTCTGCTA ATCTGTGTAC CAGTGGCTGC TGCCAGTGGC
AGATCACATC GGCATCAATC CGGTGGTGAA GTTCTTGAGA CATCTGGCG GATGTATGA GCGAGACGAT TAGGACAATG GTCACCGAGC ACGGTCAACG

acII
nspBII
bsII haeI
maeI
hmaI
bsII haeI
haeIII/palI
scrFI
ncII
mspI
hpaII
dsav
cauII
pleI
hinfi
cauII
hinfI
pleI
bsaWI
hpaII
mspI
nspBII
fnu4HI
bbvI
hinPI mcrI
hhaI/cfoI
5801 GATAAGTCGT GTCTTACCG GTTGACTCA AGACGATAGT TACCGGATAA GGCGCAGCGG TCGGGCTGAA CGGGGGGTTC GTGCACACAG CCCAGCTTGG
CTATTACGCA CAGAAATGGC CAACCTGAGT TCTGTCTATCA ATGGCCTATT CCGCGTGGCC AGCCCGACTT GCGCCCCAAG CACGTGTGTC GGTTCGAACC

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FIG. 90

[illegible]

FIG. 9N

FIG. 9N

4601

4701

4801

4901

FIG. 9M

hinPI
 hhaI/cfoI
 thaI
 fnuDII/mvnl
 bstUI
 nspBII bsh1236I
 aciI hgaI
 nspBII aciI hgaI
 4101 ACAATCTGCT CTGATGCCG ATAGTTAAGC CAACTCGCT ATCGCTACGT GACTGGGTCA TGGCTGGCC CGACACCCG CCAACACCCG CTGACGGCCG
 TGTTAGACGA GACTACGGG TATCAATTG GTTAGGCGA TAGCGATGCA CTGACCCAGT ACCGACGGG GGTGTGGC GACTGGCCG
 mspI hpaII
 scrFI
 nciI
 dsav sfaNI
 cauII foki
 drdI
 4201 CTGACGGGCT TGTCTGCTCC CGGCATCCG TTACAGACAA GCTGTGACCG TCTCCGGGAG CTGCATGTGT CAGAGGTTTT CACCGTCATC ACCGAAACGC
 GACTGCCGA ACAGACGAGG GCGTAGGCG AATGTCTGTT CGACACTGGC AGAGGCCCTC GACGTACACA GTCTCCAAA GTGGCAGTAG TGGCTTTGCG
 mspI hpaII
 scrFI
 nciI
 dsav sfaNI
 cauII foki
 drdI
 4301 GCGAGGCAGT ATTCTTGAAG ACGAAGGGC CTCGTGATAC GCCTATTTTT ATAGTTAAT GTCATGATAA TAATGGTTTC TTAGAGTCA GGTGGCAGCTT
 CGCTCCGCTA TAAGAACTTC TGCTTTCCCG GAGCACTATG CGGATAAAAA TATCCAATTA CAGTACTATT ATTACCAAAG AATCTGCAGT CCACCGTGAA
 mspI hpaII
 scrFI
 nciI
 dsav sfaNI
 cauII foki
 drdI
 4401 TTCCGGGAAA TGTGCGGGA ACCCCTATTT GTTTATTTTT CTAAATACAT TCAATATGT ATCCGCTCAT GAGACAATAA CCCTGATAAA TGCTTCAATA
 AAGCCCCCTT ACACGGCCT TGGGGATAA CAATAAAAA GATTTATGTA AGTTTATACA TAGGCGAGTA CTCTGTTATT GGGACTATTT ACGAAGTTAT
 mspI hpaII
 scrFI
 nciI
 dsav sfaNI
 cauII foki
 drdI
 4501 ATATTGAAAA AGGAAGAGTA TGAGTATTCA ACATTTCCGT GTCGCCCTTA TTCCCTTTTT TGCGGCATTT TGCCTTCCCTG TTTTGTCTCA CCCAGAAACG
 TATAACTTTT TCCTTCTCAT ACTCATAAGT TGTAAAGGCA CAGCGGGAAT AAGGAAAAA ACGCCGTAAA ACGGAAGGAC AAAAAGCAGT GGGTCTTTGC

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[illegible]

FIG. 9K

[illegible]

FIG. 9J

taqI{dam-}
clai/bsp106{dam-}
sau3AI
mboI/ndeII{dam-}
dpmI{dam+}
dpmII{dam-}
lwi{dam-}
GAT
CTA

2801 AAAGCAATAG CATCACAAAT TTACACAAATA AAGCATTTTT TTCACCTGCAT TCTAGTTGTG GTTCTGCCAA ACTCATCAAT GTATCTTATC ATGCTCGGAT
TTTCGTTATC GTAGTGTTTA AAGTGTTTTAT TTCGTAAAAA AAGTGACGTA AGATCAACAC CAAACAGGTT TCAGTAGTTA CATAGAAATG TACAGACCTA

[illegible]

3001	TGTGTGTCAG	TTAGGGTGTG	GAAAGTCCCC	AGGCTCCCCA	GCAGGCAGAA	GTATGCCAAG	CATGCATCTC	AATTAGTCAG	CAACCCAGTG	TGGAAGTCC
	ACACACAGTC	AATCCACAC	CTTTCAGGG	TCCGAGGGGT	CGTCCGTCTT	CATACGTTTC	GTACGTAGAG	TTAATCAGTC	GTGTGTCAC	ACCTTTCAGG

[illegible]

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FIG. 9H

[illegible]

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FIG. 9G

```

scrFI      hinPI      nlaIV
mvaI      narI      kasi
ecorIII   hinII/acyI hgiCI
ecoNI     hphI      haeII
dsav      mspI      bspI
bstNI     hpaII     bmyI
bsII      cfrI01    fnu4HI
apyI(dcm+) bsaWI tthIII/aspI
fnu4HI     bslI ageI maeIII ddeI hhaI/cfoI nspBII alw44I/snoI cauII dsav
bbvI      bslI agcI maeIII ddeI hhaI/cfoI nspBII alw44I/snoI cauII dsav
1801 GGTGCTCTGG TCAAGGACTA CTTCCCGGAA CCGGTGACGG TGTCGTGGAA CTCAGGGCGC CTGACCAGCG GCGTGCACAC CTTCCCGGCT GTCTACAGT
CGGACGGACC AGTTCTTGAT GAAGGGGCTT GGCACCTGCC ACAGCACCTT GAGTCCGCGG GACTGGTCCG CGCAGGTGTG GAAGGGCCGA CAGGATGTCA

fnu4HI     nlaIV
ddeI pleI fnu4HI hgiCI
mnII hinfi mnII bbvI bspI
eco8II     mnII fnu4HI maeIII bspI286 rmaI bspI286
bsu36I/mstII/sauI ddeI bbvI hphI bmyI maeI aluI bmyI
1901 CCTCAGGACT CTACTCCCTC AGCAGGTGG TGACTGTGCC CTCTAGCAGC TTGGGCACCC AGACCTACAT CTGCAACGTG ANTCAACAGC CCAGCAACAC
GGAGTCCTGA GATGAGGGAG TCGTCCGACC ACTGACACCG GAGATCGTCG AACCCGTGGG TCTGGATGTA GACGTGCAC TTAGTGTTCG GGTCTGTGTG

eaml105I
sau96I
scrFI      mvaI      avaiI
ecoRII     dsav      bstNI
bstNI      bsaJI nlaIV
bsaJI      apyI(dcm+) mboII mboII
apyI(dcm+) bbsI mnII
2001 CAAGGTGGAC AAGAAAGTTG AGCCCAAAATC TTGTGACAAA ACTCACACAT GCCCACCCTG CCCAGCACCT GAATCTCTGG GGGACCCGTC AGTCTTCTCTC
GTTCCACCTG TTCTTTCAAC TCGGGTTTAG AACACTGTTT TGAGTGTGTA CGGGTGGCAC GGGTCTGTGA CTTGAGGACC CCCCTGGCAG TCAGAAAGGAG

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FIG. 9F

scrFI mvaI ecorII dsav bstNI apyI[dcn+] hinPI hhaI/cfoI nlaIV nlaI kasi hinII/acyI hgiCI haeII bani ahaII/bsaHI
 1601 ACCTGCAGAT GAACAGCCTG CGTGCCTGAGG ACACTGCCGT CTATTATTGT GCTCGAGGCA GCCACTATTTC CGGCGCCTGG CACTTCGCCG TGTGGGGTCA
 TGGACGCTCA CTTCGCGGAC GCACGACTCC TGTGACGGCA GATAATAACA CGAGCTCCGT CGGTGATAAA GCCGGGACC GTGANGCGGC ACACCCCACT
 scfI pstI bsgI bspMI ddeI drdI mnlI xhoI paeR7I auaI hgiAI/aspHI bsp1286 bsiHKAI fnu4HI bmyI taqI bbvI
 sau96I haeIII/palI sau96I nlaIV hgiJII bsp1286 bmyI bni scrFI mvaI ecorII dsav bstNI hgiAI/aspHI bsp1286
 1701 AGGAACCTG GTCACCGTCT CCTCGGCTC CACCAAGGCG CCATCGGTCT TCCCGCTGGC ACCCTCTCC AAGAGCACCT CTGGGGGCAC AGCGGCCCTG
 TCCTTGGGAC CAGTGGCAGA GGAGCGGAG GTGTTTCCG GGTAGCCAGA AGGGGAGCCG TGGAGGAGG TTCTCGTGA GACCCCGCTG TCCTCGGAC

FIG. 9E

FIG. 9E

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FIG. 9D

[illegible]

FIG. 9C

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```

      haeIII/palI
      haeI
scrFI      scrFI      mvaI      mvaI      mvaI      mvaI      mvaI      mvaI      mvaI      mvaI
      ecorII      ecorII      ecorII      ecorII      ecorII      ecorII      ecorII      ecorII
dsav      tfII      dsav      bstNI      nlaIII      bstNI      ddel      pleI
      apyI(dcm+)      hinfi      apyI(dcm+)      hinfi      apyI(dcm+)      hinfi      apyI(dcm+)      hinfi
801 ACAACCGGAA TTGCGAAGTA AAGTAGACAT GGTTTGATA GTGCGAGGCA GTTCTGTTTA CCAGGAAGCC ATGAATCAAC CAGGCCACCT TAGACTCTTT
TGTGGCCTT AACCGTTTCA TTCACTCTGA CCAACCTAT CAGCCTCCGT CAAGACAAAT GGTCTCTGGG TACTTAGTGG GTCCGGTGGG ATCTGAGAAA

      accI      nlaIII      mnlI
      mspI      hpaII      bsaBI
      mboI/ndeII(dam-)
      dpnI(dam+)
      dpnII(dam-)
      maeIII      alwI(dam-)      apoI      maeIII      mnlI
901 GTGACACGGA TCATCCAGGA ATTTGAAAGT GACACGTTTT TCCAGAAAT TGATTTGGG AAATATAAAC CTCTCCAGA ATACCAGGC GTCTCTCTG
CACTGTTCT AGTACGTCCT TAAACTTTCA CTGTGCAAAA AGGGTCTTTA ACTAAACCCC TTATATTTG GAGAGGGTCT TATGGGTCCG CAGGAGAGAC

      scrFI      mvaI      ecorII      dsav      bstNI      apyI(dcm+)      mnlI      bsaJI      bslI      ddel
      sau96I      avaiI
      asuI      mnlI      sfaNI      accI      mboII      mboII      mnlI      aluI
1001 AGGTCCAGGA GGAAGAAGGC ATCAAGTATA AGTTTGAAGT CTACGAGAAG AAAGACTAAC AGGAGATGC TTTCAGATTG TCTGTCTCCC TCCTAAGCT
TCCAGGTCCT CCTTTTCCG TAGTTCATAT TCAAACTTCA GATGCTCTTC TTTCGATTG TCCTTCTACG AAAGTTCAAG AGACGAGGG AGGATTTGGA

      styI      bsaJI
      sau3AI
      mboI/ndeII(dam-)
      dpnI(dam+)
      dpnII(dam-)
      alwI(dam-)
      bstYI/xhoII
      ppu101      nsii/avaIII      bsaJI
      aluI      tru9I      mseI
      fnu4HI      bbvI      aseI/asnI/vspI
1101 ATGCATTTT ATAAGACCAT GGAACCTTTT GTGGCTTTAG ATCCCTTGG CTTCGTTAGA AGCAGCTAC AATTAATACA TAACCTTATG TATCATACAC
TAGGTAAAAA TATTCTGGTA CCTGAAAC GACCGAATC TAGGGGAACC GAAGCAATCT TCGCTCGATG TTAATTATGT ATTGGAATAC ATAGTATGTG

```

BNSDOCID: <WO___9604391A1_I_>

FIG. 9A

[illegible]

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FIG. 8

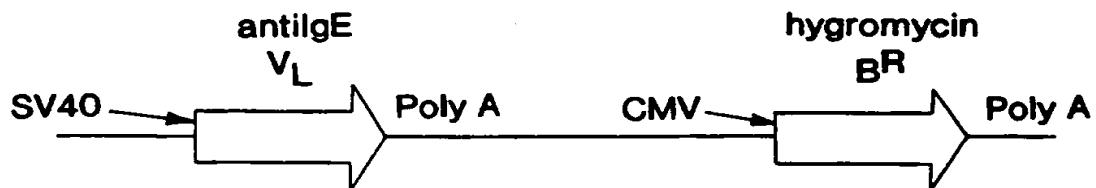
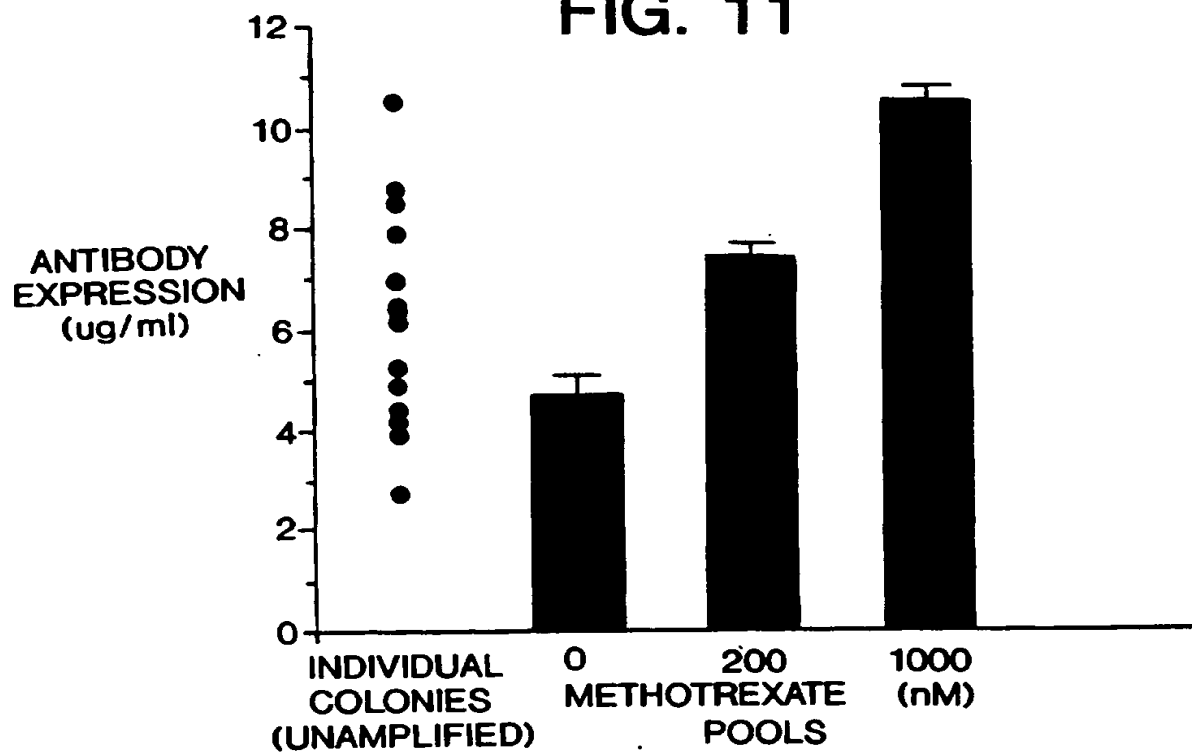


FIG. 11



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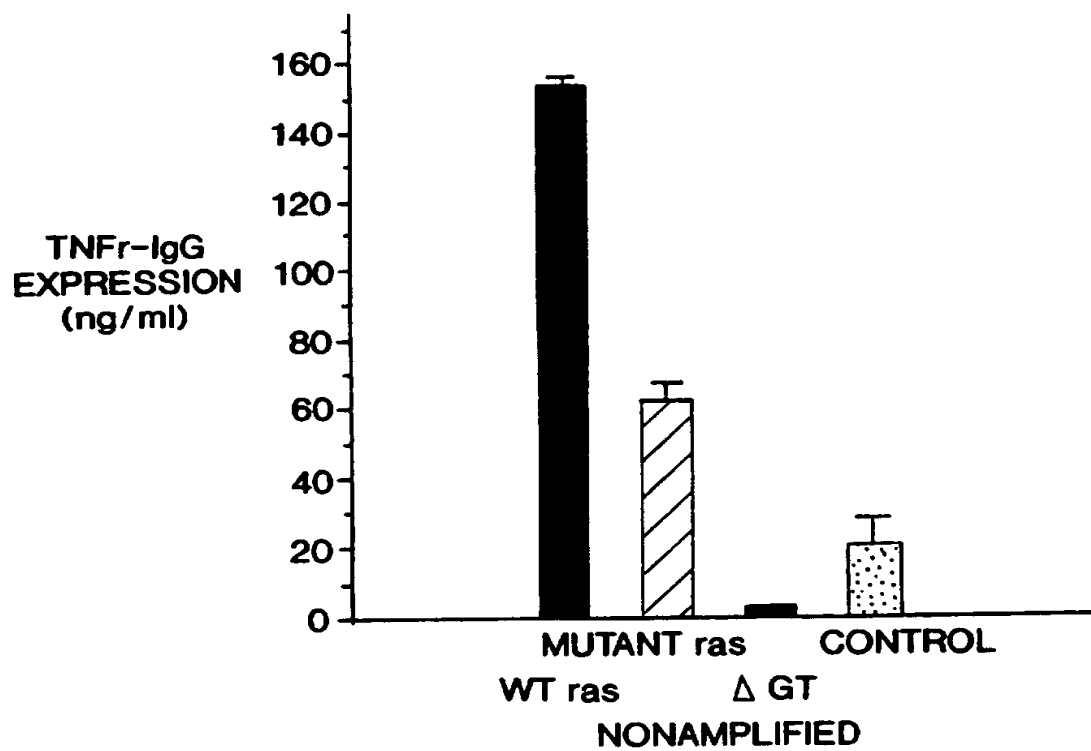


FIG. 7A

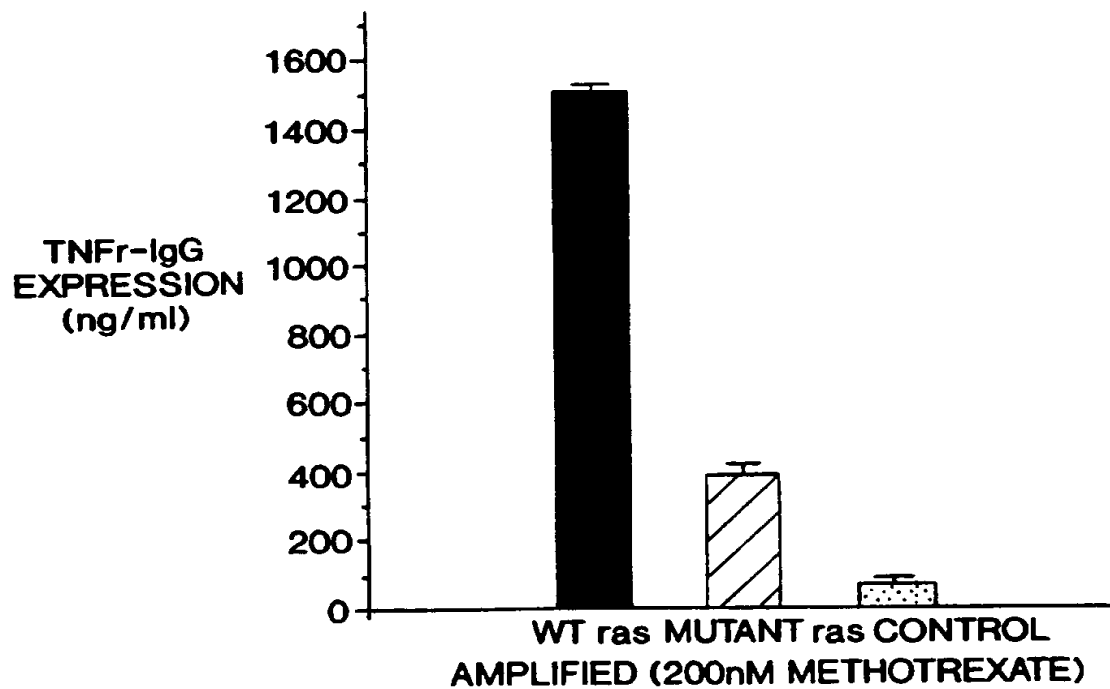


FIG. 7B

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FIG. 6R

```

haeIII/palI
haeI
scrFI
mvaI bslI
ecorII
dsav
nlaIII
nspi
haeIII/palI nspHI
bstNI
apyI[dcM+]
haeI
aflIII
6501 CCTGGCCCTT TGCTGCACAT GTTCTTTCTT GCGTTATCCC CTGATTCTGT GGATAACCGT ATTACCGCCT TTGAGTGAGC TGATACCGCT
GGACCCGAA ACACCGGAA AACGAGTGTA CAAGAAGGA CGCAATAGG GACTAAGACA CCTATTGGCA TAATGGCGA AACTCACTCG ACTATGCGCA
bsrBI
aciI
alul
6601 CCGCGCAGCC GAACGACCGA GCGCAGCGAG TCAGTGAGCG AGGAAGCGGA AGAGCGCCCA ATACGCAAC CGCTCTCCC CGCGGTGG CCGATTCAAT
GCGCGTCGG CTGTGGCT AGTCAGTC AGTCAGTC CCTTTCGCC TCCTTCGCC TCTCGGGT TATCGGTTG GCGGAGAGG GCGCGCAAC GGCTAAGTAA
thaI
fnuDII/mvni
bstUI
bsh1236I
hinPI
hhaI/cfoI
thaI
fnuDII/mvni
bstUI haeIII/palI tru9I
bsh1236I
bslI eaeI tfil ael/asnI/vspI
aciI mnlI acil cfrI hinFI msel
6601 CCGCGCAGCC GAACGACCGA GCGCAGCGAG TCAGTGAGCG AGGAAGCGGA AGAGCGCCCA ATACGCAAC CGCTCTCCC CGCGGTGG CCGATTCAAT
GCGCGTCGG CTGTGGCT AGTCAGTC AGTCAGTC CCTTTCGCC TCCTTCGCC TCTCGGGT TATCGGTTG GCGGAGAGG GCGCGCAAC GGCTAAGTAA
scrFI
mvaI
ecorII
dsav
nlaIV bstNI
hgiCI apyI[dcM+]
banI bsaJI
6701 AATCCAGCTG GCACGACAGG TTTCCCGACT GGAAGCGGG CACTGAGCG CACTGAGCTA ATGTGAGTTA CCTCACTCAT TAGGCACCCC AGGCTTTACA
TTAGGTCGAC CGTGTGTCC AAAGGGCTGA CCTTTCGCC GTCAGTCGG GTCACTCAAT TACTCTCAAT GGAGTGAGTA ATCCGTGGG TCCGAAATGT
tru9I
msel
ael/asnI/vspI
xmnI
6801 CTTTATGCTT CCGGCTCGTA TGTGTGTGG AATTGTGAGC GGATAACAAT TTCACACAGG AACAGCTAT GACCATGATT ACGAATTAA
GAAATACGAA GGCCGAGCAT ACAACACACC TTAACACTCG CCTATTGTTA AAGTGTGTC TTTGTGATA CTGGTACTAA TGCTTAATT

```

>length: 6889

FIG. 6Q

6101 TAATCCTGTT ACCAGTGGCT GCTGCCAGTG GCGATAAGTC GTGCTTTACC GGGTTGGACT CAAGACGATA GTTACCGGAT AAGGCGCAGC GGTCCGGGCTG
ATTAGGACAA TGGTCACCGA CGACGGTCAC CGCTATTTCAG CACAGAATGG CCAACCTGA GTTCTGCTAT CAATGGCCTA TTCCGGCTCG CCAGCCCGAC

6201 AACGGGGGT TCGTGCACAC AGCCAGCTT GGAGCGAAGC ACCTACACCG AACTGAGATA CTPACAGCGT GAGCATTGAG AAAGCGCCAC GCTTCCCGAA
TTGCCCCCA AGCAGGTGTG TCGGGTCGAA CCTCGCTTGC TGGATGTGGC TTGACTCTAT GGATGTGCA CTGTAATCTC TTTCGGGTG CGAAGGGCTT

6301 GGGAGAAAG CGACAGGTA TCCGGTAAGC GGCAGGGTGG GAACAGGAGA GCGCAGGAGG GAGCTTCCAG GGGGAAACGC CTGGTATCTT TATAGTCTCG
CCCTCTTCC GCGTGTCCAT AGGCCATTGG CCGTCCAGC CTTGTCTCTCT CCGGTGCTCC CTCGAAGGTC CCCCTTGGC GACCATAGAA ATATCAGGAC

6401 TCGGGTTTCG CCACCTCTGA CTTGAGCGTC GATTTTGTG ATGCTCGTCA GGGGGGCGGA GCCTATGGAA AAACGCCAGC AACCGGCCT TTTTACGGTT
AGCCCAAAGC GGTGGAGACT GAACTCGCAG CTAAACAC TACGAGCAGT CCCCCCGCT CGATACCTT TTTTCGGTGG TTGCGCCGGA AAAATGCCAA

6501 GGGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG

6601 GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG

6701 GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG

6801 GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG

6901 GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG

7001 GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG

7101 GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG

7201 GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG

7301 GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG

7401 GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG

7501 GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG

7601 GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG

7701 GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG

7801 GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG

7901 GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG

8001 GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG

8101 GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG

8201 GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG

8301 GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG

8401 GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG

8501 GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG

8601 GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG

8701 GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG

8801 GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG

8901 GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG

9001 GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG

9101 GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG

9201 GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG

9301 GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG

9401 GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG

9501 GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG

9601 GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG

9701 GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG

9801 GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG

9901 GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG

10001 GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG GCGGCTGCTG

FIG. 6P

5701 AAGCATTGGT AACGTGCAGA CCAAGTTTAC TCATATATAC TTAGATTGA TTAAAACTT CAITTTTAAT TTAAGAAGAT CTAGGTGAAG ATCCTTTTGG
 TTCGTAACCA TTGACAGTCT GGTTCAAATG AGTATATATG AAATCTNACT AAATTTTGAA GTAAAAATTA AATTTTCCTA GATCCACTTC TAGGAAAAAC
 maeIII
 tru9I ahaIII/draI maeI alwI/dam-] bstyI/xhoII mboII/dam-]
 mseI tru9I bstyI/xhoII bstyI/xhoII bstyI/xhoII
 ahaIII/draI mseI mseI alwI/dam-] mboII/dam-]
 sau3AI
 mboI/ndeII(dam-]
 dpnI(dam+] sau3AI
 dpnII(dam-] mboI/ndeII(dam-]
 bstyI/xhoII dpnI(dam+]
 sau3AI alwI/dam-] dpnII(dam-]
 mboI/ndeII(dam-] alwI(dam-]
 dpnI(dam+] mboII(dam-]
 dpnII(dam-] bstyI/xhoII
 5801 ATAATCTCAT GACCAAAATC CCTTAACGTG AGTTTTCGTT CCACTGAGCG TCAGACCCCG TAGAAAAGAT CAAAGGATCT TCTTGAGATC CTTTTTTTCT
 TATTAGAGTA CTGGTTTTAG GGAATTGCAC TCAAAAGCAA GGTGACTCGC AGTCTGGGCG ATCTTTTCTA GTTCTCTAGA AGAAGCTCTAG GAAAAAAGA
 nlaIII maeII
 rcaI tru9I
 bspHI mseI
 hgaI
 ddeI
 sau3AI
 mboI/ndeII(dam-]
 dpnI(dam+]
 dpnII(dam-]
 alwI(dam-]
 mspi
 hpaII aluI
 maeIII eco57I
 5901 CGCGGTAAATC TGCTGCTTGC AAACAAAAAA ACCACCGCTA CCAGCGGTGG TTTGTTTGGC GGATCAAGAG CTACCAACTC TTTTCCGAA GGTAAGTGGC
 CGCGCATTAG ACGACGAACG TTTGTTTTTT TGGTGGCGAT GGTCCGCCAC GGTCCGCCAC AAACAAACCG CCTAGTTCTC GATGTTGAG AAAAAGGCTT CCATTGACCG
 hinPI fnu4HI
 hhaI/cfoI bbvI
 bsh1236I
 bspHI
 rcaI
 nlaIII
 maeII
 tru9I
 mseI
 hgaI
 ddeI
 sau3AI
 mboI/ndeII(dam-]
 dpnI(dam+]
 dpnII(dam-]
 alwI(dam-]
 mspi
 hpaII aluI
 maeIII eco57I
 6001 TTCAGCAGAG CGCAGATACC AAATACGTGC CTTCTAGTGT AGCCGTAGTT AGGCCACCAC TTCAAGAACT CTGTAGACC GCCTACATAC CTCGCTCTGC
 AAGTCGTCTC GCGTCTATGG TTTATGACAG GAAGATCACA TCGGCATCAA TCCGGTGGTG AGTTCTTGA GACATCGTGG CGGATGTATG GAGCGAGAGC
 hinPI
 hhaI/cfoI
 bspHI
 rcaI
 nlaIII
 maeII
 tru9I
 mseI
 hgaI
 ddeI
 sau3AI
 mboI/ndeII(dam-]
 dpnI(dam+]
 dpnII(dam-]
 alwI(dam-]
 mspi
 hpaII aluI
 maeIII eco57I

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FIG. 60

sau96I
 avaiI
 sau3AI asuI
 mboI/ndeII(dam-)
 dpnI(dam+)
 dpnII(dam-)
 pvuI/bspCI
 mcrI mnlI aluI aciI
 haeIII/palI
 eaeI
 cfrI
 fnu4HI
 aciI
 nlaIII
 bbvI
 fnu4HI
 maeIII
 nlaIII
 sau3AI
 mboI/ndeII(dam-)
 dpnI(dam+)
 dpnII(dam-)
 nlaIII alwI(dam-)
 GCACAAACATG GGGGATCATG TAACTGCGCT
 ATTCTCTTAA TAGCTACGA CGGTATTGGT ACTCACTATT GTGACGCCGG TTGAATGAAG ACTGTTGCTA GCCTCCTGSC TTCTCTGATT GGCGAAAAAA
 5201 TAAGAGAATT ATGCAGTGCT GCCATAACCA TGAGTGATAA CACTGCGGCC AACTTACTTC TGACAAACGAT CGGAGGACCG AAGGAGCTAA CCGCTTTT
 5301 GCACAAACATG GGGGATCATG TAACTGCGCT TGATCGTTGG GAACCGGAGC TGAATGAAGC CATAACCAAC GACGAGCGTG ACACCAAGAT GCCAGCAGCA
 CGTGTGTGAC CCCTAGTAC ATTGAGCGGA ACTAGCAACC CTTGGCCTCG ACTTACTTCG GTATGGTTTG CTGCTCGCAC TGTTGTGCTA CGGTGCTCGT
 fnu4HI
 maeIII
 sfaNI
 bbvI
 mspI
 hpaII
 scrFI
 aluI nciI
 rmaI dsav
 maeI cauII
 TTAATTTATC TGACCTACCT CCGCTTATTT CAACGTCCTG
 5401 ATGGCAACAA CGTTGCGCAA ACTATTAACT GCGGAACCTAC TTAATTTATC TTCCCGGCAA CAATTATAG ACTGGATGGA GGCGGATAAA GTTGAGGAC
 TACCGTTGTT GCAACGCGTT TGATAATTGA CCGCTTGATG AATGAGATCG AAGGCGCGTT GTTAATTTATC TGACCTACCT CCGCTTATTT CAACGTCCTG
 bglI
 sau96I
 haeIII/palI
 hinPI asuI mspI
 hhaI/cfoI hpaII
 GGTATTATTC CCGCTGGCTT CCGCTGGCTT CCGCTGGCTT CCGCTGGCTT CCGCTGGCTT CCGCTGGCTT CCGCTGGCTT CCGCTGGCTT
 5501 CACTTCTGCG CTCGCGCCCTT CCGCTGGCTT CCGCTGGCTT CCGCTGGCTT CCGCTGGCTT CCGCTGGCTT CCGCTGGCTT CCGCTGGCTT
 GTGAAGACGC GAGCGGGAA GGCGGACCGA CCAATAACG ACTATTTAGA CCTCGGCCAC CTCGCGGCCAC TCGCACCCAG AGCGCCATAG TAACGTCGTG ACCCGGTCT
 ddeI
 sau3AI nlaIV
 mboI/ndeII(dam-) mnlI
 dpnI(dam+) hgiCI
 dpnII(dam-) banI
 tru9I
 mseI
 haeIII/palI
 sau96I
 nlaIV
 fnu4HI
 bbvI
 bsrI
 asuI
 5601 TGSTAAAGCCC TCCCGTATCG TAGTTATCTA CACGACGGGG ACTCAGGCAA CTATGGATGA ACAGAAATAGA CAGATCGCTG AGATAGTGC CTCACGTATT
 ACCATTGCGG AGGCGATAGC ATCAATAGAT GTGCTGCCCC TCAGTCCGTT GATACCTACT TGCTTTATCT GTCTAGGCAC TCTATCCAGC GAGTGACTAA

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FIG. 6N

```

nlaIV
aciI
thai
fnuDII/mvnI
bstUI
bsh1236I
hinPI
hhaI/cfoI
rcaI
bspHI
bsrBI
aciI nlaIII
4701 AATAATGGTT TCTTAGACGT CAGGTGGCAC TTTTCGGGGA AATGTGCGG GAACCCCTAT TTGTTTATTT TTCTAAATAC ATTCAAATAT GTATCCGCTC
TTATTACCAA AGAATCTGCA GTCCACCGTG AAAAGCCCCT TTACACGCGC CTTGGGGATA AACAAATAAA AAGATTTATG TAAGTTTATA CATAGCGCAG

bsmAI
sspI
mboII
earI/ksp632I
4801 ATGAGACAAT AACCCGTGATA AATGCTTCAA AATATATTGAA AAAGGAAGAG TATGAGTATT CAACATTTCG GTGTGCCCCC TATTCCTCTC
TACTCTGTTA TTGGGACTAT TTACGAAAGTT ATTATAACTT TTTCCTTCTC ATACTCATAA GTTGTAAGG CACAGCGGA ATAAGGGAAA AACCCCGTA

hgiAI/aspHI
bsp1286
sau3AI
mboI/ndeII[dam-]
dpmI[dam+] bmyI
dpmII[dam-]
eco57I
apaLI/snoI
sfaNI mboII[dam-] alw44I/snoI maeIII taqI
4901 TTGCTTCC TGTTTTGCT CACCCAGAAA CGCTGGTGAA AGTAAAGAT GCTGAAGATC AGTGGGTGC ACGAGTGGT TACATCGAAC TGGATCTCAA
AAACGGAAGG ACAAACGA GTGGTCTTT GCGACCACIT TCATTTTCTA CGACTTCTAG TCAACCCACG TGCTACCCA ATGTAGCTTG ACCTAGAGTT

hphI
hphI
maeII
psp1406I
xmnI
asp700
mboII
4901 TTTGCTTCC TGTTTTGCT CACCCAGAAA CGCTGGTGAA AGTAAAGAT GCTGAAGATC AGTGGGTGC ACGAGTGGT TACATCGAAC TGGATCTCAA
AAACGGAAGG ACAAACGA GTGGTCTTT GCGACCACIT TCATTTTCTA CGACTTCTAG TCAACCCACG TGCTACCCA ATGTAGCTTG ACCTAGAGTT

aciI
sau3AI
mboI/ndeII[dam-]
dpmI[dam+]
dpmII[dam-]
alwI[dam-]
bstYI/xhoII
bsrI dpmII[dam-]
5001 CAGCGGTAAG ATCCTTGAGA GTTTTCGCC CGAAGAACGT TTTCCATGA TGAGCACTTT TAAAGTTCTG CTATGTGGCG CGGTATTATC CGGTGATGAC
GTCGCCATTC TAGGAACTCT CAAAGCGGG GCTTCTTGCA AAAGGTTACT ACTCGTAAA ATTCAAGAC GATACACCGC GCCATAATAG GGCACACTG

scrFI
nciI
mspi
hpaII
dsav
5101 GCCGGCAAG AGCAACTCGG TCGCCCGCAT CACTATTCTC AGAATGACTT GGTGAGTAC TCACCAGTCA CAGAAAAGCA TCTTACGGAT GGCATGACAG
CGCCCCGTC TCGTTGAGCC ACGGCGGTAT GTGATAAGAG TCTTACTGAA CCAACTCATG AGTGTGCTGT GTCTTTTCTG AGAATGCCTA CCGTACTGTC

hpaII
dsav
5101 GCCGGCAAG AGCAACTCGG TCGCCCGCAT CACTATTCTC AGAATGACTT GGTGAGTAC TCACCAGTCA CAGAAAAGCA TCTTACGGAT GGCATGACAG
CGCCCCGTC TCGTTGAGCC ACGGCGGTAT GTGATAAGAG TCTTACTGAA CCAACTCATG AGTGTGCTGT GTCTTTTCTG AGAATGCCTA CCGTACTGTC

```

FIG. 6M

FIG. 6M

4201	ATCGCCCTGA	TAGACGGTTT	TTGCCCTTT	GAGGTGGAG	TCCACGTTT	TTAATAGTG	ACTCTGTT	CAAACCTGAA	CAACACTCAA	CCCTATCTCG
	TAGCGGGA	CT	CTGCAACCT	AGGTGCAAG	AATTATCACC	AGGTGCACTT	GGGATAGAGC			

	thai									
	fnuDI1/mvni					fnuDI1/mvni				
	tru9I		apoI		tru9I		apoI		tru9I	
	msei		msei		msei		msei		msei	
4301	GGCTATTCTT	TTGATTTTATA	AGGGATTTTG	CCGATTTCGG	CCTATTGGT	AAAAATGAG	CTGATTTTAC	AAAAATTTAA	CGCGAATTTT	AACAAATAT
	CCGATAAGAA	AACATAATAT	TCCCTAAAAC	GGCTAAAGCC	GGATAAGCC	TTTTTTACTC	GACTAAATTC	TTTTTAAATT	GGCGCTAAAA	TTGTTTTATA

	hgiAI/aspHI	bsp1286	bsiHKA1	bmyI	ddeI	aciI	tru9I	aciI	maeIII	fnu4HI	hinPI
maeII				apaLI/snoI	rsal	fnu4HI			maeII bsrI	nlaiII hhaI/cfoI	
psp1406I				alw44I/snoI	csp6I	sfaNI	mseI		bsaAI tth11I/aspI	bbvI	
4401	TAACGTTTAC	AATTTTATGG	TGCACCTCTCA	GTACAATCTG	CTCTGATGCC	GCATAGTTAA	GCCAACTCCG	CTATCGCTAC	GTGACTGGGT	CATGCGTGGC	
	ATTGCAAATG	TTAAAATACC	ACGTGAGAGT	CATGTTAGAC	GAGACTACGG	CGTATCAATT	CGGTTGAGGC	GATAGCGATG	CAC TGACCCCA	GTACCGACGC	

[illegible][illegible]

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FIG. 6L

hinPI
 hhaI/cfoI
 nlaIV
 nari
 kasi
 hinII/acyI
 hgiCI
 haeII
 bani
 sfaNI
 ahaII/bsaHI
 bglI
 acII
 fnu4HI
 acII
 thal
 fnuDII/mvni
 bstUI
 hinPI
 hhaI/cfoI
 hinPI
 thal
 fnuDII/mvni
 bstUI
 scfi
 bsh1236I
 rsaI
 hhaI/cfoI
 fnu4HI
 tru9I
 bsh1236I
 csp6I
 bslI
 acII
 mseI
 hhaI/cfoI
 mspI
 hpaII
 naeI
 maeII
 cfr10I
 sau3AI
 mboI/ndeII[dam-]
 sau96I
 haeIII/palI
 asuI
 mnlI
 dpnI[dam+]
 dpnII[dam-]
 pvuI/bspCI
 merI
 mboII
 aciI
 pvuI/bspCI
 earI/ksp632I
 mcrI
 aluI
 pvuII
 nspBII
 foki
 CATCCCCCT TCGCCAGCTG GCGTAATAGC GCGAGGCGCC GCACCGATCG CCTTCCCAA CAGTTGCGTA GCCTGAATGG CGAATGGGC CTGATCGCGT
 GTAGGGGGA AGCGTCGAC CGCATTATCG CTTCTCCGG CGTGGTAGC GCGAAGGCTT GTCAACGCAT CGGACTTACC GCTTACCOCG GACTACGCCA
 3801
 fnu4HI
 hinPI
 hhaI/cfoI
 thal
 fnuDII/mvni
 bstUI
 sfaNI
 aciI
 maeII
 acII
 maeII
 csp6I
 bslI
 acII
 rsaI
 hhaI/cfoI
 fnu4HI
 tru9I
 bsh1236I
 csp6I
 bslI
 acII
 mseI
 hhaI/cfoI
 mspI
 hpaII
 naeI
 maeII
 cfr10I
 3901
 ATTCTCTCT TACGCATCTG TCGGGTATT CACACCGCAT ACGTCAAGC AACCATAGTA CGGCCCTGT AGCGCGCAT TAAGCGCGC GGGTGTGGT
 TAAAGAGGA ATCGGTAGAC ACGCCATAA GTGTGGCGTA TGCACTTCG TTGCTATCAT GCGCGGACA TCGCGCGTA ATTGCGCGC CCCACACCAC
 fnu4HI
 hinPI
 hhaI/cfoI
 thal
 fnuDII/mvni
 bstUI
 maeII
 bbvI
 maeII
 acII
 TACACTTGC TACACTTGC AGCGCCCTAG CGCCGCTCC TTTCGCTTC TTCTCGCCAC GTTCGCGGC TTTCCCGTC
 CAATGCGCGT CGCACTGGC ATGTGAACGG TCGCGGATC GCGCGGAGG AAGCGAAAG AAGGGAAGGA AAGAGCGGTG CAAGCGCGC AAAGGGCAG
 4001
 nlaIV
 hgiJII
 bsp1286
 bmyI
 banII
 aluI
 AAGCTCTAAA TCGGGGGCTC CCTTTAGGGT TCCGATTTAG TGCTTTACG CACCTCGACC CCAAAAAACT TGAITTTGGT GATGTTTAC GTAGTGGCC
 TTCGAGATT AGCCCCCGAG GGAATCCCA AGGCTAAATC ACGAATGCC GTGGAGCTGG GGTTTTGA ACTAAACCA CTACCAAGTG CATCACCOCG
 4101
 maeII
 haeIII/palI
 draII
 sau96I
 bsaI
 asuI
 hphI

FIG. 6K

nlaIV
 scrFI
 mvaI
 ecoRII
 dsav
 bstNI
 apyI[dcn+]
 bsaJI
 ppulOI
 nsiI/avaIII
 nlaIII
 sphI
 nspi sfanI
 nspHI
 nlaIV
 scrFI
 mvaI
 ecoRII
 dsav
 bstNI
 apyI[dcn+]
 bsaJI
 ppulOI
 nsiI/avaIII
 nlaIII
 sphI
 nspi sfanI
 nspHI
 nlaIV
 scrFI
 mvaI
 ecoRII
 dsav
 bstNI
 apyI[dcn+]
 bsaJI
 3401 GAAAGTCCCC AGGCTCCCCA GCAGGCAGAA GTATGCAAG CATGCATCTC AATTAGTCAG CAACCCAGGTG TGGAAAGTCC CCAGGCTCCC CAGCAGGCAG
 CTTTCAGGG TCCGAGGGGT CGTCCGTCTT CATACGTGTTT GTACGTAGAG TTAATCAGTC GTTGGTCCAC ACCTTCAGG GGTCCGAGGG GTCGTCCGTC

sfanI
 ppulOI
 nsiI/avaIII
 nlaIII
 sphI
 nspi
 nspHI
 3501 AAGTATGCAA AGCATGCATC TCAATTAGTC AGCAACCATA GTCCCGCCCC TAACTCGCC CATCCGCCCT CTAACTCGCC CCAGTTCGGC CCATTCTCCG
 TTCATACGTT TCGTACGTAG AGTTAATCAG TCGTGGTAT CAGGGCGGG ATTGAGGGG GTAGGGGGG GATTGAGGG GGTCAAGGG GGTAAAGAGG

nlaIII
 styI
 ncoI
 dsaI
 bsaJI
 fnu4HI
 sfiI mnlI
 haeIII/palI
 bsaJI bglI
 haeIII/palI bsaJI mnlI aluI
 mnlI mnlI aciI haeIII/palI
 mnlI mnlI
 3601 CCCCATGGCT GACTAATTTT TTTTATTTAT GCAGAGGCCG AGGCCGCTC GGCCTCTGAG CTATTCCAGA AGTAGTGAGG AGGCTTTTTT GGAGGCCTAG
 GGGGTACCGA CTGATTAAAA AAATAAATA CGTCTCCGC TCCGGCGGAG CCGGAGACTC GATAAGTCT TCATCACTCC TCCGAAAAAA CCTCCGGATC

nlaIV
 scrFI
 mvaI
 ecoRII
 dsav
 bstNI
 apyI[dcn+]
 bsaJI
 ppulOI
 nsiI/avaIII
 nlaIII
 sphI
 nspi sfanI
 nspHI
 nlaIV
 scrFI
 mvaI
 ecoRII
 dsav
 bstNI
 apyI[dcn+]
 bsaJI
 fnu4HI
 sfiI mnlI
 haeIII/palI
 bsaJI bglI
 haeIII/palI bsaJI mnlI aluI
 mnlI mnlI aciI haeIII/palI
 mnlI mnlI
 3701 GCTTTTGCAA AAAGCTGTTA ACAGCTTGGC ACTGGCCGTC GTTTTACAAC GTCGTGACTG GGAACACCT GGCCTTACCC AACTTAATCG CCTTGACAGCA
 CGAAACGTT TTTGACAAT TGTCGAACCG TGACCGGCG CAAATGTTG CAGCACTGAC CCTTTTGGGA CCGCAATGGG TTGAATTAGC GGAACGTCGT

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[illegible]

FIG. 61

[illegible]

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FIG. 6H

```

          eam1105I
          sau96I
          scrFI
          mvaI      avaII
          ecoRII
          dsav
          bstNI    asuI      mboII mboII
          bsaJI    mnlI      bpuAI earI/ksp632I    styI
          apyI(dcm+) bbsI mnlI      bsaJI
          2401 CTGTGTGACAC ACCTCCCCCA TGCCACGCGT GCCCAGCACC TGAACCTCCTG GGAGGACCGT CAGTCTTCCT CTTCCTCCCA AAACCCAAGG ATACCTTTAT
              GAACACTGTG TGGAGGGGGT ACGGGTGCCA CGGGTGTGG ACTTGAGGAC CCTCCTGGCA GTCAGAGGA GAAGGGGGGT TTTGGGTTCC TATGGGAATA

          sau96I
          nlaIV
          avaII
          asuI      maeII
          mspI      pmlI
          hpaII     eco72I
          scrFI     mnlI bsaAI
          nciI      ddel maeIII
          dsav      eco81I bbrPI
          cauII     bsu36I/mstII/sauI      maeII
          2501 GATTTCCCGG ACCCTGAGG TCACGTGCGT GGTGGTGGAC GTGAGCCACG AAGACCCCGA GGTCAGTTC AAGTGGTACG TGGACGGCGT GGAGGTGCAT
              CTAAAGGGCC TGGGGACTCC AGTGCACGCA CCACCACCTG CACTCGGTGC TTCTGGGGCT CCAGGTCAAG TTCACCATGC ACCTGCCGCA CCTCCACGTA

          acil
          thal
          fnuDII/mvnI
          bstUI
          bsh1236I
          sacII/sstII
          nspBII
          kspI
          dsal
          bsaJI
          acil
          fnu4HI mnlI      maeII
          2601 AATGCCAAGA CAAAGCCCGG GGAGGAGCAG TTCAACAGCA CGTTCCGTGT GGTCCGTC CTCACCGTC CTACCGTCG TGCACCAAGGA CTGGCTGAAC GGCAAGGAGT
              TTACGGTTCT GTTTCGGCGC CCTCTCGTC AAGTTGTCGT GCAAGGCACA CCAGTCGCAG GAGTGGCAGG ACGTGGTCTT GACCGACTTG CCGTTCTCTCA

          scrFI
          mvaI bsrI
          ecoRII
          dsav
          bstNI
          mnlI      ecoNI    bstNI
          hgaI    hphI    bsII    apyI(dcm+)
          rsaI
          csp6I
          rsaI
          csp6I

```

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FIG. 6G

hgiAI/aspHI bsp1286 bsiHRAI bmyI apaLI/snoI alw44I/snoI
 hgiAI/aspHI scrFI mvaI ecorII dsav bstNI apyI[dcM+] draIII
 bsp1286 bsiHRAI bmyI apaLI/snoI alw44I/snoI
 sau96I auaII bmyI mnlI dsav bstNI apyI[dcM+] draIII
 asuI apaLI/snoI bstNI apyI[dcM+] draIII
 mnlI nlaIV alw44I/snoI apyI[dcM+] draIII
 mnlI bsvI muni bsrI
 alwNI fnu4HI
 2001 CGGCATTATT GGAGTGAAA CCTTTTCCAG TGCTTCAATT GCAGCCTCTG CCTCAATGGG ACGGTGCACC TCTCTGCCA GGAGAAACAG AACACCGTGT
 GCCGTAATAA CTCACCTTTT GGAAAAGGTC ACGAAGTTAA CGTCGGAGAC GGAGTTACCC TGGCAGGTGG AGAGGACGGT CCTCTTTGTC TTGTGGCACA
 hgiAI/aspHI bsp1286 bsiHRAI bmyI
 gsui/bpmI scrFI mvaI apaLI/snoI ecorII dsav
 bstNI alw44I/snoI apyI[dcM+]
 2101 GCACCTGCCA TGCAGGTTTC TTCTTAAGAG AAAACGAGTG TGCTCTCTGT AGTAACTGTA AGAAAAGCCT GGAGTGCACG AAGTTGTGCC TACCCACAGAT
 CGTGGACGGT ACGTCCAAAG AAAGATTCTC TTTTGCTCAC ACAGAGGACA TCATTGACAT TCTTTTCGGA CTCACGTGC TTCAACACGG ATGGGGTCTA
 bspMI nlaIII ddeI bsmAI maeIII
 aluI sstI sacI hgiJII
 hgiAI/aspHI ecl136II
 bsp1286 bsiHRAI bmyI banII
 nlaIV pleI hgiCI maeIII hphI nspI dsai bani
 nspI bsaJI bmyI
 2201 TGAGAAATGTT AAGGCACATG AGGACTCAGG CACCACAGAC AAGAGAGTTG AGCTCAAAAC CCCACTTGGT GACACAATC ACACATGCC ACCGTGCCA
 ACTCTTACAA TTCCCGTGAC TCCTGAGTCC GTGGTGCTG TTCTCTCAAC TCGAGTTTG GGGTGAACCA CTGTGTTGAG TGTGTACGG TGCCACGGGT
 bsp1286 nlaIV hgiCI hgiJII
 dsai bmyI bsp1286 bmyI
 bsp1286 bani bmyI bsaJI banII
 maeIII mnlI maeIII mnlI nlaIII bani
 2301 GAGCCCAAT CTGTGTGAC ACCTCCCCCG TGCCACGGT GCCCAGACCC CAATCTTGT GACACACCTC CCCCATGCC ACCGTGCCA GAGCCCAAT
 CTCGGGTTTA GAACACTGTG TGGAGGGGGC ACGGTGCCA CGGTCTCG GTTGAACA CTGTGTGGAG GGGTACGG TGCCACGGGT CTCGGGTTTA

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FIG. 6F

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scrFI      nciI      mspI      hpaII      dsav      cauII
xmaI/pspAI
smaI
scrFI      nciI      dsav      cauII      bslI
sau96I
haeIII/palI
asuI
scrFI
mvaI bsaJI
ecorII
dsav      bstNI bsaJI      scfI      pstI      bsgI
nlaIV      fnu4HI      aciI      hphI      eco57I      alwNI
          bsrBI      hphI      eco57I      ddeI      mnlI
1801 CCTACTTGTA CAATGACTGT CCAGGCCCGG GGCAGGATAC GAGTGTGAGA CGGCTCCTT CACCGCTTCA GAAACCACC TCAGACACTG
GGATGAACAT GTTACTGACA GGTCCGGGCC CCGTCTATG CCTGACGTCC CTCACACTCT CGCCAGGAA GTGCGGAAGT CTTTGGTGG AGTCTGTGAC

          fnu4HI      aluI      pvuII      nspBII      ddeI      mnlI      bbvI
          scrFI      mboII      earI/ksp632I      sau3AI      mboI/ndeII[dam-]      dpnI[dam+]      dpnII[dam-]      bstYI/xhoII      bglII
          nciI      mspI      hpaII      dsav      cauII      sau96I      avalI      asuI      draIII      bboII      mboII      bsrI cfr10I
          mspI      hpaII      rsaI      csp6I      fnu4HI      bboII      mboII      bsrI cfr10I
1901 CCTCAGCTGC TCCAAATGCC GAAAGAAAT GAGTCAAGTG GAGATCTCTT CTTCACAGT GGACCGGGAC ACCGTGTGTG GCTGCAGGAA GAACCACTAC
GGAGTCGACG AGGTTTACGG CTTTCCTTTA CCAGTCCAC CTCTAGAGAA GAACGTGTCA CTTGCCCTCTG TGCCACACAC CGACGTCTT CTTGGTCATG

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FIG. 6E

haeIII/palI
 eaeI
 cfrI
 mspI
 hpaII
 scrFI
 nciI
 ecorI dsav
 taqI apoI cauII
 clai/bsp106 bsaJI aluI
 mnlI
 bsaJI
 hincII/hindII
 apyI[dcm+] mnlI
 gsuI/bpmI bsaJI
 maeIII
 foki
 scfI
 CACTATAGAA TAACATCCAC TTTCCTTTC TCTCCACAGG TGTCACTCCA GGTCAACTGC ACCTCGGTTT CATCGATTGA ATTCCCGGC CATAGCTGTC
 GTGATATCTT ATTGTAGTG AACGGAAAG AGAGGTGTC ACAGTGAGGT CCAGTTGACG TGGAGCCAAAG ATAGCTAACT TAAGGGCCG GTATCGACAG
 1501
 gsuI/bpmI
 scrFI
 mvaI
 ecorII
 dsav
 bstNI
 apyI[dcm+] mnlI
 bsaJI
 hincII/hindII
 maeIII
 foki
 scfI
 CACTATAGAA TAACATCCAC TTTCCTTTC TCTCCACAGG TGTCACTCCA GGTCAACTGC ACCTCGGTTT CATCGATTGA ATTCCCGGC CATAGCTGTC
 GTGATATCTT ATTGTAGTG AACGGAAAG AGAGGTGTC ACAGTGAGGT CCAGTTGACG TGGAGCCAAAG ATAGCTAACT TAAGGGCCG GTATCGACAG
 1601
 mnlI
 haeIII/palI
 sau96I
 asuI
 nlaIII
 TGGCATGGG CTCTCCACCG TGCCTGACCT GCTGCTGCCG CTGCTGCTCC TGGAGCTGTT GGTGGGAATA TACCCCTCAG GGGTTATTGG ACTGGTCCCT
 ACCGTACCCG GAGAGGTGGC ACGGACTGGA CGACGACGGC GACCACGAGG ACCTCGACAA CCACCCCTAT ATGGGAGTC CCCAATAACC TGACCAGGGA
 1701
 rmaI
 maeI
 styI
 bsaJI
 blnI
 avrII
 mboII
 earI/ksp632I
 styI
 bsaJI
 mnlI
 taqI
 rsaI
 csp6I
 nlaIV
 CACCTAGGG ACAGGGAGAA GAGAGATAGT GTGTGTCCCC AAGGAAATA TATCCACCTT CAAATTAATT CGATTTCCTG TACCAAGTGC CACAAGGAA
 GTGGATCCCC TGTCCTCTT CTCTCTATCA CACACAGGGG TTCTTTTAT ATAGTGGA GTTTTATTA GCTAAACGAC ATGTTTCAAG GTGTTTCCTT

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[illegible]

FIG. 6C

tfilI
 acilI
 thaI hinfI
 fnuDII/mvnI
 bstUI
 bsh1236I
 701 TTGGAACGG GATTCCCGT GCCAAGAGTG CTGTAAGTAC CGCCTATAGA GCGATAAGAG GATTATATCC CGCTGCCAT CATGTTTGA CCATTGAAC
 AACCTTGGC CTAAGGGCA CGTTCTCAG GACATTCTAC CGGTCTCAG CCGGATATCT CGCTATTCTC CTAAATAGG GCGACGGTA GTACCAAGCT GGTAACTTGA
 fnu4HI
 bbvI
 nspBII
 acilI
 nlaIII
 taqI
 thaI
 fnuDII/mvnI
 bstUI
 bsh1236I
 mlui
 bsrBI
 aflIII
 rsaI
 acilI
 xmnI
 csp6I
 mnli
 ddelI
 asp700
 scaI
 801 GCATCGTGC CGTGTCCTCA AATATGGGA TTGGCAAGAA CGGAGACCTA CCCTGCCCTC CGCTCAGAA CGCTTCAAG TACTTCCAA GAATGACCAC
 CGTAGCAGG GCACAGGTT TTATACCOCT AACCGTTCTT GCCTCTGGAT GCGACGGGAG GCGAGTCTT GCGCAAGTTC ATGAAGTTT CTTACTGGTG
 scrFI
 mvaI
 ecorII
 dsav
 bstNI
 apyI(dcm+)
 sexAI
 tfilI
 hinfI
 mboII
 taqI
 mseI
 tru9I
 mseI
 aseI/asnI/vspI
 eco57I
 mboII
 earI/ksp632I
 mnli
 alwNI
 hphI
 901 AACCTCTTCA GTGGAAGTA AACAGATCT GGTGATTATG GGTAGAAAA CCTGGTTCTC CATTCCTGAG AAGATCGAC CTTTAAAGGA CAGATTAAT
 TTGGAGAGT CACCTTCCAT TTGTCTTAGA CCACTAATAC CCATCTTTT GGACCAAGAG GTAAGGACTC TTCTTAGCTG GAAATTTCT GTCTTAATTA
 aluI
 sstI
 sacI
 hgiJII
 hgiAI/aspHI
 ecl136II
 bsp1286
 bsiHKA
 bmyI
 banII
 bslI
 mnli
 bstXI
 fokI
 sfanI
 mseI
 tru9I
 mspI
 hpaII
 bsaWI
 1001 ATAGTTCTCA GTAGAGACT CAAGAACCA CCACGAGGAG CTCATTTCTT TGCCAAAGT TTGGATGATG CCTTAAGACT TATTGAACAA CCGGAATTGG
 TATCAAGAGT CATCTCTTGA GTTCTTGGT GGTGCTCCTC GAGTAAAGA ACGGTTTTCA AACCTACTAC GGAATCTTGA ATAACTTGT GGCCTTAACC

FIG. 6B

FIG. 6B

```

401  GGTTTGGCA  GTACATCAAT  GGGGGTGGAT  AGCGGTTTGA  CTCACGGGGA  TTTCCAAGTC  TCCACCCCAT  TGAGTCAAT  GGGAGTTTGT  TTTGGACCA  CCAAAACCGT  CATGTAGTTA  CCGCACCTA  TCGCCAAACT  GAGTGCCCT  AAAGGTTTCA  AGGTGGGTA  ACTGCAGTTA  CCTCAACA  AAACCGTGT

      rsal      csp6I      pleI      hinfI      acil      hgaI      maeII      acil      hgaI      csp6I      mnlI      banII
      nlaIV      hnlI/acyI      ahaII/bsaHI      aatII      bsmAI      rsal      csp6I      mnlI      banII
      sstI      sacI      hgiJII      hgiAI/asphI      eel36II      bsp1286      bsiHKAI      bmyI
      aluI      sstI      sacI      hgiJII      hgiAI/asphI      eel36II      bsp1286      bsiHKAI      bmyI
      aluI      sstI      sacI      hgiJII      hgiAI/asphI      eel36II      bsp1286      bsiHKAI      bmyI

501  AAATCAACGG  GACTTTCCAA  AATGTCGTAA  CAATCCGCC  CCATTGACGC  AAATGGGCGG  TAGGCGTGT  CGGTGGGAGG  TCTATATAAG  CAGAGTCGT  TTAGTTGCC  CTGAAGGTT  TTACAGCATT  GTTGAGGCGG  GGTAACTGCG  TTTACCCGCC  ATCCGCACAT  GCCACCTTCC  AGATATATTC  GTCTCGAGCA

      maeII      acil      hgaI      csp6I      mnlI      banII
      rsal      csp6I      mnlI      banII
      haeIII/palI      mcrI      eagI/xmaIII/ecI XI      eaeI      cfrI      fnu4HI      aciI      thal      fnuDII/mvnI
      sacII/stII      nspBII      kspI      scrFI      dsal      nciI      bglI      bslI      mspI      sau3AI      mnlI      bstUI      mboI/ndeII[dam-]      hpaII      dpnI[dam+]      bsaJI      dsav      dpnII[dam-]      bsh1236I      alwI[dam-]      aciI      cauII
      sau96I      avaII      asuI      nlaIV      scrFI      nciI      mspI      hpaII      mboI      bpuAI      bbsI      cauII      mnlI      gttttgacct      ccatagaaga      caccgggacc      gatccagcct      ccggcgccgc      gaaagggtgca
      esp3I      scrFI      mvaI      bsmAI      ecorII      dsav      bstNI      hinII/acyI      apyI(dcm+)      sau3AI      gsuI/bpmI      mboI/ndeII[dam-]      dpnI[dam+]      hgaI      fokI      dpnII[dam-]      ahaII/bsaHI      gttttgacct      ctggagacgc      catccacgct      gattcttct      gtaggtgcca      caaaactgga      ggtatcttct      gtggccctgg      ctaggtcgga      gccgcggcc      ctggcacgt

601  TTAGTGAACC  GTCAGATCGC  CTGGAGACGC  CATCCACGCT  GTTTTGACCT  CCATAGAAGA  CACCGGACC  GATCCAGCCT  CCGGCGCCG  GAAAGGTGCA  AATCATTGG  CAGTCTAGCG  GACTCTGCG  GTAGTGCGA  CAAACTGGA  GGTATCTTCT  GTGGCCCTGG  CTAGGTGCGA  GCGCGCGCC  CTGGCACGT

```

BNSDOCID: <WO__9604391A1_I >

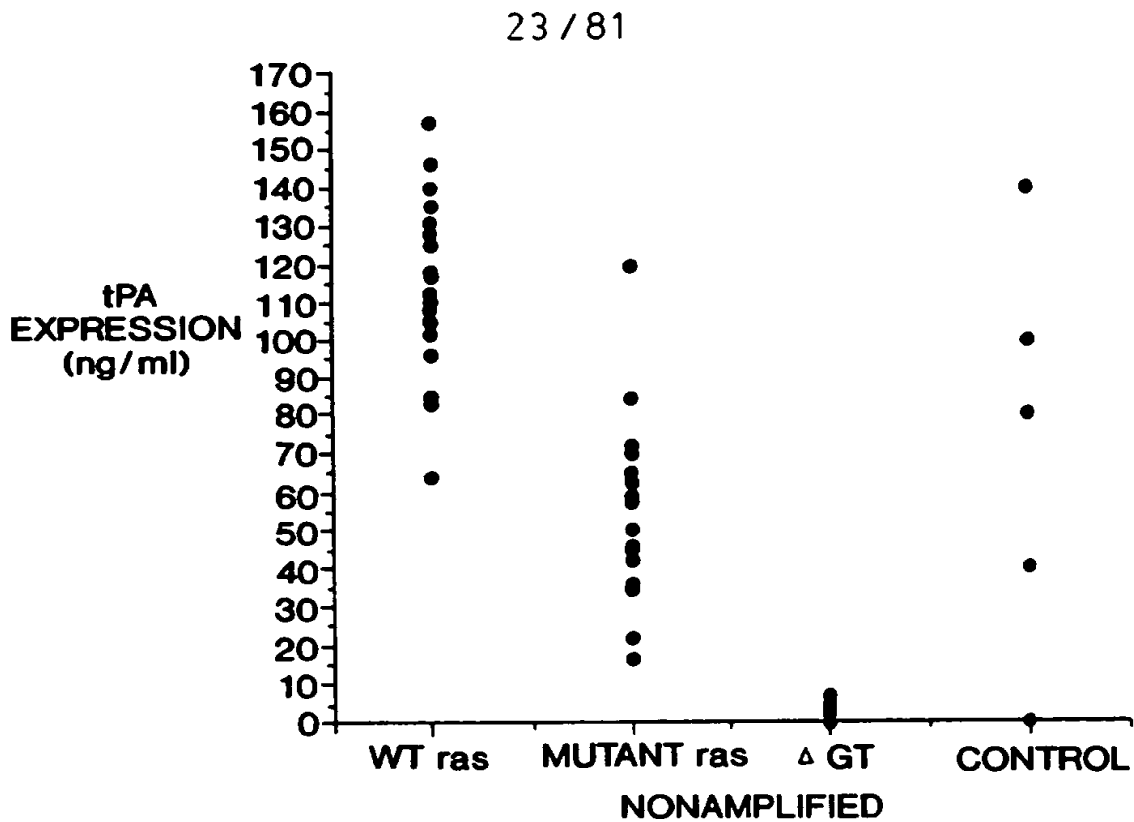
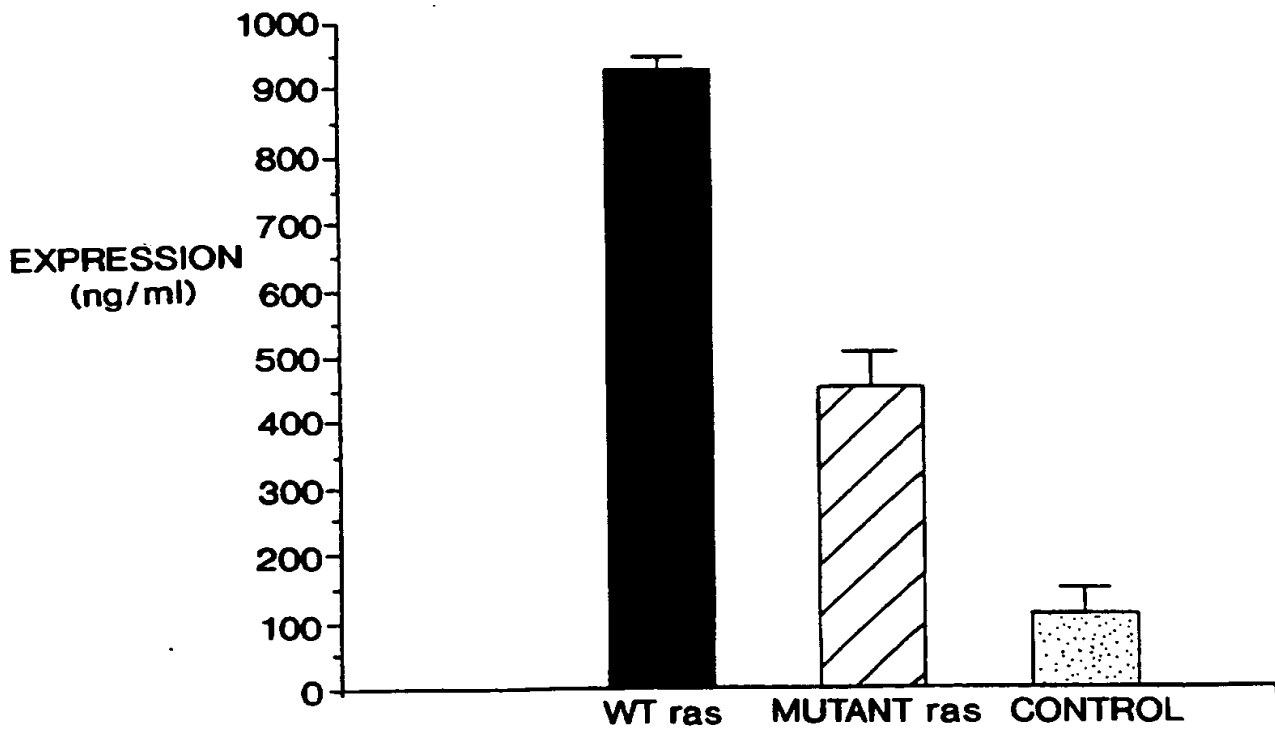


FIG. 5B



AMPLIFIED (200nM METHOTREXATE)

FIG. 5C

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FIG. 10B

[illegible]

[illegible]

FIG. 10D

hinPI mspI hhaI/cfoI hgiAI/aspHI
 hhaI/cfoI hpaI bspI286
 thai hpaI bsiHKAI
 aciI mroI bmyI
 haeIII/palI bspMI
 mcrI fnuDII/mvnI bspEI
 eagi/xmaIII/ecI XI bsaWI
 eaeI bstUI tfiI
 cfrI bsh1236I hinfi
 sfaNI fnu4HI bslI accIII
 mboI/ndeII(dam-) mspI
 dprII(dam+) apol
 dprII(dam-) ecorI
 1101 GATCCTTATG TTTATCGGCA CTTTGCATCG GCGCGCTCC CGATTCCGGA AGTGCTTGAC ATTGGGAAT TCAGCGAGAG CTGACCTAT TGCATCTCCC
 CTAGCAATAC AAATAGCCGT GAAACGTAGC CGCGCGGAGG GCTAAGGCT TCACGAACCTG TAACCCCTTA AGTCGCTCTC GGACTGGATA ACGTAGAGGG

mcrI eagi/xmaIII/ecI XI
 eaeI fnu4HI
 styI
 thai ncoI
 fnuDII/mvnI
 fnu4HI bstUI dsal
 bbvI mcrI haeIII/palI dprII(dam-) sau3AI
 scfI mspI bsh1236I bsaJI sfaNI fnu4HI mboI/ndeII(dam-) dprII(dam+)

nspBI pstI hpaI mnlI nlaIII pvuI/bspCI haeIII/palI
 aciI bsgI cfr10I aciI haeI foki mcrI bbvI cfrI dprII(dam-)

sau96I rsrII/cspI
 aveII
 asuI
 sau96I haeIII/palI aciI tfiI hinfi
 haeIII/palI asuI cpoI
 ddeI bsrBI maeIII
 aciI bsrBI maeIII
 1201 GCCGTGCACA GGGTGTACG TTGCAACACC TGCCTGAAC CGAATGCCC GCTGTCTCG AGCGGTGCG GGAGGCCATG GATCGCATG CTGCGGCGGA,
 CGGCACGTGT CCCACAGTGC AACGTTGTGG ACGGACTTGT GCTTGACGGG CGACAAGAGC GTCGCGTAC CTACGCTAGC GACGCCGCT

sau3AI
 thai fnuDII/mvnI nlaIII
 bstUI mboI/ndeII(dam-)
 bsh1236I dprII(dam-)
 hinPI dprII(dam-)
 ndeI hhaI/cfoI alwI(dam-)
 nlaIII
 1301 TCTTAGCCAG ACGAGCGGT TCGGCCCAT TCGGCCGCA GGAATCGGTC AATACACTAC ATGGCGTAT TCCATATCGG CGATTGCTGA TCCCATGTG
 AGAATCGGTC TGCTCGCCCA AGCCGGGTAA GCCTGGCGTT CCTTAGCCAG TTATGTGATG TACCGCACTA AAGTATACGC GCTAACGACT AGGGGTACAC

hinPI draIII
 hhaI/cfoI nlaIV
 thai hgiCI
 fnuDII/mvnI bsaJI
 haeIII/palI bsaJI
 bstUI sau96I mspI
 bsh1236I aluI asuI mnlI bslI hpaII
 tthlIII/aspl drdI
 1401 TATCACTGGC AAACGTGTGAT GGACGACACC GTCACTGCGT CCGTCCGCA GGTCTCGAT GAGCTGATG TTTGGGCCGA GGACTGCCG GAACTCCGCG
 ATAGTGACCG TTTGACACTA CCTGCTGTGG CAGTCACGCA GGCAGCGCGT CCGAGAGCTA CTCGACTAGC AAACCCGCT CCGACGCGG CTTCAGGCGG

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acil
thai
fnuDII/mvni
hgiAI/aspHI
bsp1286
bsiHKAI
bmyI bstUI
apaLI/snoI
aiw44I/snoI
mniI bsh1236I
1501 ACCTCGTCGA CGCGGATTTC
TGGAGCAGCT CGCGCTAAAG

11501 ACCTCGTCA CGCGGATTC GGCTCAACA ATGTCCTGAC GGACAATGGC CGCATACAG CGGTCAATTGA CTGGAGCGAG CGCATGTTCC GCGATATCCCA
TGGAGCACGT GCGCCCTAAG CCGAGTTGT TACAGGACTG CCTGTTACCG GCGTATTGTC GCCATGTAAC T GACTTCGCTC CGCTACAAGC CCCTAAGGGT

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fnu4HI
thaI
fnuDII/mvnI
bstUI
bsh1236I
sacII/sstII
nspBII
kspI
dsaI
bsaJI
aciI
fnu4HI
sau3AI aciI
mboI/ndeII(dam-)
dpnI(dam+)
dpnII(dam-)
alwI(dam-)
mspI
hpaII
mroI
bspMII
bspEI
bsaWI
rsal aciI foki
fnu4HI csp6I bsrBI sfaNI aluI
bbvI maeII taqI mnlI accII
fnu4HI haeIII/palI
mboII mnlI bsaJI
mboII gsuI/bpmI
mnlI

1601 ATACGAGGTC GCCAACATCT TCTTCTGGAG GCGGTGGTTG GCTTGATATG AGCAGGACATC CGGAGGCTTGC AGGATCGCCG
TATGCTCCAG CGGTTGTAGA AGAAGACCTC CGGCACCAAC CGAACATACC TCGTCGTCTG CATGAAGCTC GCCTCCGTAG GCCTCGAACG TCCTAGCGGC

Restriction Enzyme	Recognition Sequence	Enzyme
scrFI	CGGCTCCGGG	scrFI
nciI	CGGCTCCGGG	nciI
mspI	CGGCTCCGGG	mspI
hpaII	CGGCTCCGGG	hpaII
dsaV	CGGCTCCGGG	dsaV
cauII	CGGCTCCGGG	cauII
nlaiV	CGGCTCCGGG	nlaiV
1701	CGGCTCCGGG	1701

FIG. 10F

```

nlaIV
mspI      haeIII/palI
hpaII     mcrI
bslI      eagI/xmaIII/ecI XI
mroI      eaeI
bspMII    cfrI
bspEI(dam-) fnu4HI
bsaWI     aciI
accIII(dam-) thal
sau3AI    fnuDII/mvmI
mboI/ndeII(dam-) bstUI
dpmI(dam+) bsh1236I sau96I
dpmII(dam-) hinPI      avaiI
alwI(dam-)  rsaI       csp6I
              aciI      asuI
              aciI      asuI
1801 ACGCAATCGT CGGATCCGGA GCCGGGACTG TCGGGCGTAC ACAATCGCC CGCAGAAGCG CGGCCGTCTG GACCGATGGC TGTGTAGAAG TACTGCCGA
TCCGTTAGCA GGCTAGGCCT CGGCCCTGAC AGCCCGCATG TGTTAGCGG CGGTCTTGC GCGGCAGAC CTGGCTACCG ACACATCTTC ATGAGCGGCT

scrFI
nciI
mspI
hpaII
dsav
xmaI/pspAI
smaI
scrFI
nciI
dsav
cauII
bsaJI
avaI
bsaJI
sau3AI
mboI/ndeII(dam-)
dpmI(dam+)
dpmII(dam-)
alwI(dam-)
nlaIV cauII
bstVI/xhoII
bamHI bsaJI ecoRI
alwI(dam-) apoI
mcrI
bslI
sfaNI
mnII
bsaJI
hinII/acyI
hgaI
ahaII/bsaHI
1901 TAGTGGAAAC CGACGCCCCA GCACCTCGTCC GAGGGCAAAG GAATAGAGTA GATGCCGACC GAAGATCCC CGGGGAATTC AATCGATGGC CGCCATGGCC
ATCACCTTTG GCTCGGGGT CGTGAGCAGG CTCCCGTTTC CTTATCTCAT CTACGGCTGG CTTCTCTAGG GCGGCTTAAG TTAGCTACCG GCGGTACCGG

          aciI haeIII/palI
          fnu4HI asuI
          bglI nlaIII
          sfiI styI
          eaeI ncoI
          cfrI dsal
          taqI haeIII/palI
          clal/bsp106 bsaJI
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FIG. 10G

```

2001 CAACCTGTTT ATTCAGCTT ATAATGGTTA CAATAAAGC AATAGCATCA CAATTCAC AAATAAGCA TTTTTCAC TGCATTCAG TTGCGGTTG
    aluI fnu4HI bbvI maeIII sfanI apoI rnaI bsmI maeI
    GTTGAACAAA TAACGTCGAA TATTACCAAT GTTATTTCG TTATCGTAGT GTTTAAGTG TTTATTTCGT AAAAAAAGTG ACGTAAGATC AACACCAAC

    sau3AI mboI/ndeII(dam-) dpnI(dam+) dpnII(dam-) pvuI/bspCI mcrI
    taqI(dam-) tru9I claiI/bsp106(dam-) sau3AI mseI fnu4HI styI haeIII/palI haeI
    mboI/ndeII(dam-) dpnI(dam+) xmnI hinPI dsai bbvI ncoI
    dpnII(dam-) asei/asnI/vspI bsaJI nlaIII alwI(dam-) asp700 hhaI/cfoI nlaIII mnlI
    2101 TCCAAACTCA TCAATGTATC TTATCATGTC TGGATCGATC GGGAAATTAAT TCGGCGCAGC ACCATGGCCT GAAATAACCT CTGAAAGAGG AACTTGTTA
    AGGTTTGAGT AGTTACATAG AATAGTACAG ACCTAGCTAG CCCTTAATTA AGCCGCTCG TGGTACCAGA CTTTATTGGA GACTTCTCC TTGAACCAAT
    rsaI csp6I nlaIV kpnI hgiCI banI aluI pvuII nspBII
    asp718 mnlI acc65I ddeI aciI
    2201 GGTACCTTCT GAGCGGAAA GAACCAAGCTG TGGAAATGCT GTGAGTAAG GTGCGGAAAG TCCCCAGGT CCCCAGCAG CAGAAGTATG CAAAGCATGC
    CCATGGAAGA CTCGCGCTTT CTTGGTCGAC ACCTTACACA CAGTCAATCC CACACCTTTC AGGGGTCCGA GGGGTCTGCC GTCTTCATAC GTTTCGTACG
    scrFI mvaI ecorII dsav bstNI apyI(dcm+) bsaJI nlaIV
    2301 ATCTCAATTA GTCAGCAACC AGGTGTGGA AGTCCCCAGG CTCGCCAGCA GGCAGAAGTA TGCAGAGCAT GCATCTCAAT TAGTCAGCAA CCATAGTCCC
    TAGAGTTAAT CAGTCGTTGG TCCACACCTT TCAGGGGTCC GAGGGGTCTG CCGTCTTCAT ACGTTTCGTA CGTAGAGTTA ATCAGTCGTT GGTATCAGGG
    scrFI mvaI ecorII dsav bstNI apyI(dcm+) bsaJI nlaIV sphI nspI sfanI nspHI aciI
    sexAI

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FIG. 10H

fnu4HI
 bglI
 sfiI
 haeIII/palI
 mnlI
 haeIII/palI
 mnlI bsaJI aciI
 TTTATGCAGA GGCCGAGGCC
 AAATACGTCT CCGGCTCCGG
 nlaIII
 styI
 ncoI
 bslI dsal
 aciI bsaJI
 CTCCGCCCA TGGCTGACTA ATTTTTTTAA
 TTTATGCAGA GGCCGAGGCC
 TAAAAAAAAT AAATACGTCT CCGGCTCCGG
 scrFI
 nciI
 mspI
 hpaII
 dsav
 haeIII/palI
 mcrI
 eagI/xmaII/eclXI
 eaeI
 cfrI
 mspI caulI
 hpaII
 aluI aluI
 hpaII
 tagCTTATCC GGCCGGGAAC GGTCATTTGG
 ATCGAATAGG CCGGCCCTTG CCACGTAACC
 styI
 bsaJI
 blnI
 haeIII/palI
 stuI rmaI
 haeI maeI
 mnlI avrII
 haeIII/palI
 ddeI
 mnlI
 bsaJI mnlI aluI
 CTGAGCTATT CCAGAAAGTAG TGAGGAGGCT TTTTGGAGG CCTAGGCTTT TGCAAAAAGC TAGCTTATCC GGCCGGGAAC GGTCATTTGG
 GACTCGATTA GGTCTTCATC ACTCCTCCGA AAAAACCTCC GGATCCGAAA AGTTTTCG ATCGAATAGG CCGGCCCTTG CCACGTAACC
 tfiI
 hinFI
 aciI
 thaI
 fnuDII/mvnI
 bstUI
 bsh1236I
 AACGGCGATT CCGCGTGCCA AGAGTCAGGT AAGTACCGCC TATAGAGTCT ATAGGCCAC CCCCTTGGCT TCGTTAGAAC CCGGCTACAA TTAATACATA
 TTGCGCCTAA GGGGCACGGT TCTCAGTCCA TTCTAGGCGG ATATCTCAGA TATCCGGGTG GGGGAACCGA AGCAATCTTG CCGCGATGTT AATTATGAT

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FIG. 10I

```

sau3AI      sau96I
mboI/ndeII(dam-)  avaiI
dpnI(dam+)      asuI
dpnII(dam-)     scrFI
alwI(dam-)      mvaI
taqI(dam-)      ecoRII
claiI/bsp106(dam-)  dsav
sau3AI          bstNI
mboI/ndeII(dam-)  apyI(dcm+)
dpnI(dam+)      bslI bsaJI
dpnII(dam-)     foki
alwI(dam-)      bslI bsaJI
2701 ACCTTTTGGG TCGATCCTAC TGACACTGAC ATCCACTTTT TCTTTTCTC CACAGGTGTC CACTCCGAGG TCCAACTGCA CCTCGGTTCC CGAAGCTAGC
TGGAAACCT AGCTAGGATG ACTGTGACTG TAGGTGAAA AGAAAAAGAG GTGTCCACAG GTGAGGGTCC AGGTTGACGT GGAGCCAAGC GCTTCGATCG

nlaIII
styI
pflMI
ncol
sfanI      ecorI      rsaI
fnu4HI taqI apoI      gsuI/bpmI
bbvI claiI/bsp106    bsaJI      rmaI
nlaIII foki      maeI      pvuII tth111I/aspI
2801 TTGGGGTCCA TCGATTGAAT TCCACCATGG GATGGTCATG TATCATCCTT TTTCTAGTAG CAACTGCAAC TGGAGTACAT TCAGATATCC AGCTGACCCA
AACCCGACGT AGCTAACCTA AGTGGGTACC CTACCAGTAC ATAGTAGGAA AAAGATCATC GTTGACGTTG ACCTCATGTA AGTCTATAGG TCGACTGGGT

aluI
sstI
sacI
hgiJII
hgiAI/aspHI
ec1136II
bsp1286
bsiHKA1
bmyI
banII      mnlI      taqI
avaI      aciI      hphI      hmaIII      bspMI      bsrI      hphI      aluI      nlaIII      bsrI
2901 GTCCCCGAGC TCCCTGTCCG CCTCTGTCCG CGATAGGGTC ACCATCACCT GCCGTGCCAG TCAGAGCGTC GATTACGATG GTGATAGCTA CATGAAGCTGG
CAGGGGCTCG AGGACAGGC GGAGACACCC GCTATGCCAG TGGTAGTGGA CGGCACGGTC AGTCTCCGAG CTAATGCTAC CACTATCGAT GTACTTGACC

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FIG. 10J

[illegible]

FIG. 10K

sstI
 sacI
 hgiII
 hgiAI/aspHI
 eci136II
 bsp1286
 bsiHKA1
 bmyI
 haeIII/palI
 sau96I aluI
 asuI banII
 hphI
 eco0109I/draII
 maeIII alwNI ddeI
 accI
 cgcCTGCGAA
 gTCACCCATC AGGGCTGAG
 GCGGACGCTT CAGTGGGTAG TCCCGGACTC

ddeI
 celII/espI
 bpu1102I
 hgaI
 ddeI fnu4HI
 scfI mnlI bbvI
 CTACAGCCTC AGCAGCACCC
 TGACGCTGAG CAACGAGAC
 TACGAGAAC ACAAGTCTA
 GTCCTGCTG ATGCTCTTG
 TGTTCAGAT GCGGACGCTT
 CAGTGGGTAG TCCCGGACTC

sau96I
 nlaIII
 aciI haeIII/palI
 fnu4HI asuI
 bglI styI
 sfII ncoI
 aluI
 hindIII eaeI dsal
 tru9I cfrI bsaJI
 mseI taqI haeIII/palI
 maeIII aluI
 TCAACAGGGG AGAGTGTAA
 GCTTCGATGG CCGCATGGC
 CCAACTGTGT TATTGCAGCT
 TATAATGGTT ACAATAAAG
 GAGCGGCAG TGTTTCTCGA
 AGTTGTCCCC TCTCACAATT
 CGAAGCTACC GCGGTACCG
 GGTTGAACAA ATAACGTCA
 ATATTACCA TGTATTATTC

sau3AI
 mboI/ndeII(dam-)
 dpnI(dam+)
 dpnII(dam-)
 pvuI/bspCI
 mcrI
 taqI(dam-)
 claI/bsp106(dam-)
 sau3AI
 mboI/ndeII(dam-)
 dpnI(dam+)
 dpnII(dam-)
 nlaIII alwI(dam-)
 imal
 bsmI maeI
 ctgcATTCTA GTTGTGGTTT
 GTCCAACTC ATCAATGTAT
 CTTATCATGT CTGGATCGAT
 TAAATAAAGT GACCTAAGAT
 CAACACCAG CAGGTTTGG
 TAGTTACATA GAATAGTACA
 GACCTAGCTA

sfanI apoI
 CAATAGCATC ACAATTCTCA
 CAAATAAAGC ATTTTTTCA
 CTGATTCTA GTTGTGGTTT
 GTCCAACTC ATCAATGTAT
 CTTATCATGT CTGGATCGAT
 GTTATCGTAG TGTTTAAAGT
 GTTTATTTCG TAAATAAAGT
 GACCTAAGAT CAACACCAG
 CAGGTTTGG TAGTTACATA
 GAATAGTACA GACCTAGCTA

FIG. 10L

[illegible]

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[illegible]

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FIG. 100

```

scrFI      thai      fnuDII/mvnI
nciI       bstUI
mspI       bsh1236I
hpaII fnu4HI hinPI
dsav aluI nspHI hhaI/cfoI
cauII bbvI nlaIII mnlI hphI hphI hphI
5001 TCCGGGAGCT GCATGTGTCA GAGGTTTCA CGGTCATCAC CGAACCGCG GAGGCAGTAT TCTTGAAGAC GAAAGGGCCT CGTGATACGC CTATTTTAT
AGGCCCTCGA CGTACACAGT CTCCAAAAGT GGCAGTAGTG GCTTTGGCG CTCCGTCATA AGAAGCTTCTG CTTTCCCGGA GCACTATGCG GATAAAATA
nlaIV
aciI
thai
fnuDII/mvnI
bstUI
bsh1236I
hinPI
hhaI/cfoI
maeII
hinII/acyI
ahaII/bsaHI
dclI aatII
5101 AGGTTAAGT CATGATAATA ATGGTTTCTT AGAGTTCAGG TGGCACTTTT CGGGGAAATG TCGCGGAAC CCTATTGT TTAATTTTCT AAATACATTC
TCCAATTACA GTACTATTAT TACCAAGAA TCTGCAGTCC ACCGTGAAA GCCCCTTTAC AGCGCCTTG GGGATAACA AATAAAAGA TTTATGTAG
rcaI
bspHI
bsrBI bsmAI
aciI nlaIII
5201 AAATATGTAT CCGTTCATGA GACAATAACC CTGATAAATG CTCAATAAT ATTGAAAAG GAAGAGTATG AGTATTCAC ATTTCCGTGT CGCCCTTATT
TTTATACATA GCGGAGTACT CTGTATTGG GACTATTAC GAAGTTATTA TAACTTTTC CTCTCATAC TCATAAGTTG TAAAGGCACA GCGGGAATAA
hgiAI/aspHI
bsp1286
sau3AI
mboI/ndeII(dam-)
dpmI(dam+) bmyI
dpmII(dam-)
hpaI
hphI
hphI
hphI
5301 CCCTTTTTG CGGCAFTTTG CCTTCCTGTT TTTGCTCACC CAGAACGCT GGTGAAAGTA AAAGATGCTG AAGATCAGTT GGTGCACGA GTGGTTACA
GGGAAAAAAC GCCGTAAAC GCGGTAAAC GGAAGGACAA AAACGAGTGG GTCTTTGCGA CCACCTTCAT TTTCTACGAC TTCTAGTCAA CCCACGTGCT CACCCAATGT

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FIG. 10P

sau3AI mboI/ndeII(dam-) sau3AI mboI/ndeII(dam-) acilI
 dpnI(,am+) dpnI(dam+) dpnI(dam+) thaI
 i-styI/xhoII dpnII(dam-) dpnII(dam+) fnuDII/mvnI
 bsrI nspBII alwI(dam-) alwI(dam-) bstOI bstOI
 taqI alwI(dam-) aciI bstyI/xhoII bsh1236I
 5401 TCGAACCTGGA TCTCAACAGC GGTAAAGTCC TTGAGAGTTT TCGCCCGGAA GAAGCTTTTC CAATGATGAG CACTTTTAAA GTTCTGCTAT GTGGCGCGGT
 AGCTTGACCT AGAGTTGTGC CCAITCTAGG AACTCTCAA AGCGGGGCTT CTTCGAAAAG GTTACTACTC GTGAAAATTT CAAGACGATA CACCGCGCCA
 scrFI
 nciI
 mspI
 hpaII
 dsav
 cauII
 hinII/acyI
 hgaI
 ahaII/bsaHI bcgI mcrI fnu4HI aciI
 5501 ATTATCCCGT GATGACGCCG GGCAAGACCA ACTCGGTGCG CGCATACACT ATTCTCAGAA TGACTTGGTT GAGTACTCAC CAGTCACAGA AAAGCATCTT
 TAATAGGCA CTACTGCGC CGTTCTCGT TGAGCCAGCG GCGTATGTGA TAAGAGTCTT ACTGAACCAA CTCATGAGTG GTGAGTGTCT TTTCGTAGAA
 hinII/acyI
 hgaI
 ahaII/bsaHI bcgI mcrI fnu4HI aciI
 5601 ACGGATGGCA TGACAGTAAG AGAATTATGC AGTGTGCGCA TAACCATGAG TGATAACACT GCGGCCAACT TACTTCTGAC AAGCATCGGA GGACCGAAGG
 TGCCTACCGT ACTGTCAATC TCTTAATACG TCACGACGGT ATTGTACTC ACTATTGTGA CGCCGGTTGA ATGAAGACTG TTGCTAGCCT CCTGGCTTCC
 foki nlaIII fnu4HI bbvI nlaIII
 sau3AI maeIII
 mboI/ndeII(dam-) sau3AI nlaIV mspI
 dpnI(dam+) mboI/ndeII(dam-) dpnI(dam+) hpaII
 alwI(dam-) dpnII(dam-) bsaWI aluI
 5701 AGCTAACCGC TTTTTCGAC AACATGGGGG ATCATGTAAC TCGCCTTGAT CGTTGGGAC CGGAGCTGAA TGAAGCCATA CCAAGGACG AGCGTGACAC
 TCGATTGGCG AAAAACGCTG TTGTACCCCT TAGTACATTC AGCGGAACCTA GCAACCCCTTG GCCTCGACTT ACTTCGGTAT GGTTCGCTGC TCGCACTGTG
 maeIII

FIG. 10Q

[illegible]

FIG. 10R

sau3AI
 mboI/ndeII(dam-)
 dpnI(dam+)
 dpnII(dam-)
 bstYI/xhoII
 sau3AI alwI(dam-)
 mboI/ndeII(dam-)
 dpnI(dam+)
 dpnII(dam-)
 ddeI hgaI
 sau3AI
 mboI/ndeII(dam-)
 dpnI(dam+)
 dpnII(dam-)
 alwI(dam-)
 mspI
 hpaII
 acII
 nspBII
 acII
 nspBII
 hpaII
 aluI
 mspI
 haeIII/palI
 haeI
 bslI
 haeI
 scfI
 acII
 6201 GTGAAGATCC TTTTIGATAA TCTCATGACC AAAATCCCTT AACGTGAGTT TTCGTTCCAC TGAGCGTCAG ACCCGTAGA AAAGATCATAA GGATCTTCTT
 CACTTCTAGG AAAAATACTATT AGAGTACTGG TTTTAGGGAA TTGCACTCAA AAGCAAGGTG ACTCGCAGTC TGGGGCATCT TTTCTAGTTT CCTAGAAGAA
 sau3AI
 fnuDII/mvnI
 mboI/ndeII(dam-)
 dpnI(dam+)
 bstUI
 bsh1236I
 dpnII(dam-)
 hinPI
 fnu4HI
 bstYI/xhoII
 hhaI/cfoI
 bbvI
 6301 GAGATCCTTT TTTTCTGCGC GTAATCTGCT GCTTGCAAC AAAAATAACCA CCGTACCAG CGGTGGTTTG TTTGCGGAT TTTGCGGAT CAAGAGCTAC CAACCTCTTT
 CTCTAGGAAA AAAAGACGCG CATTAGACGA CGAACGTTTG TTTTITTTGGT GCGATGCTC GCCACCAAC AAACGGCCTA GTTCTCGATG GTTGAGAAAA
 bsrI
 hinPI
 hhaI/cfoI
 maeIII
 eco57I
 hhaI/cfoI
 maeI
 bslI
 haeI
 scfI
 acII
 6401 TCCGAAGGTA ACTGGCTTCA GCAGAGCGCA GATACCAAT ACTGTCTTC TAGTGTAGCC CACCACITCA AGAATCTGT AGCACCCTT
 AGGCTTCCAT TGACCGAAGT CGTCTCGCT CTATGTTTA TGACAGGAG ATCATATCG CATCAATCG GTGGTGAAGT TCTTGAGACA TCGTGCGGA
 mnlI
 maeIII
 bbvI
 bsrI
 fnu4HI
 alwNI
 bbvI
 fnu4HI
 pleI
 hinfi
 maeIII
 hhaI/cfoI
 6501 ACATACCTCG CTCGTCTAAT CTTGTTACCA GTGGCTGCTG CCAGTGSCGA TAAGTCGTGT CTTACCGGT TGGACTCAAG ACATAGTTA CCGATAAGG
 TGTATGGAGC GAGACGATTA GGACAATGGT CACCGACGAC GGTACCGCT ATTCAGCACA GAATGGCCCA ACCTGAGTTC TGCTATCAAT GGCCTATTCC
 mcrI
 acII
 nspBII
 fnu4HI
 bbvI
 hgiAI/aspHI
 bsp1286
 bslHKA1
 bmyI
 apaLI/snoI
 alw44I/snoI
 aluI
 ddeI
 scfI
 hinPI
 hhaI/cfoI
 haeII
 6601 CGCAGCGGTC GGGCTGAACG GGGGTTTCTG GCACACAGCC CAGCTTGGAG CGAACGACCT ACACCGAAT GAGATACCTA CAGCGTGCAG ATTGAGAAAG
 GCGTCGCCAG CCGACTGC CCCCCAAGCA CGTGTGTGCG GTCGAACCTC GCTTGTGGA TGTGGCTGA CTCTATGGAT GTGCACTCG TAACTCTTC

SUBSTITUTE SHEET (RULE 26)

FIG. 10S

FIG. 103

```

6701 CGCCACGCTT CCCGAAGGGA GAAAGCGGGA CAGGTATCCG GTAAGCGGCA GGGTCGGAAC AGGAGAGCGC ACGAGGGAGC TTCACGGGGG AAACGCCCTGG
      acil          fnu4HI          hinPI mnlI          hhaI/cfoI aluI          apyI(dcm+)
      mspI          hpaII          bslI          bsaWI          bsaJI          dsav          bstNI          bsh1236I
      hpaII          bslI          bsaWI          bsaJI          dsav          bstNI          bsh1236I
      acil          fnu4HI          hinPI mnlI          hhaI/cfoI aluI          apyI(dcm+)
      GCGGTGCGAA GGGCTTCCCT CTTTCCGCCT GTCCATAGGC CATTGCGCGT CCCAGCCTTG TCCTCTCGCG TGCCTCCCTCG AAGTCCCCC TTTGCGGACC
      scrFI          mvaI          ecorII          mvaI          ecorII          dsav          bstNI          bsaJI          apyI(dcm+)
      fnu4HI          acil          thai          fnuDII/r          bstUI          bsh1236I

6801 TATCTTTTATA GTCCTGTCCG GTTTGCGCCAC CTTGACTTGG AGCTGCGATT TTTGTGATGC TCGTCAGGGG GCGGAGCCT ATGGAANAAC GCCAGCAACG
      mnII          drdI          hgaI          taqI          sfanI          nlaIV          aciI          bsh1236I
      ATAGAAATAT CAGGACAGCC CAAGCGGTG GAGACTGAAC TGCAGCTAA AACACTACG AGCACTCCC CGCCTCGGA TACCTTTTGG CGTCTGTTGC
      haeIII/palI          scrFI          mvaI          ecorII          dsav          bstNI          bslI          apyI(dcm+)          haeIII/palI          nlaIV          haeI          haeIII/palI          nspHI          afIII          hinfI          aciI

6901 CGGCCTTTT ACGGTTCCCTG GCCTTTGCT GGCCTTTTGC TCACATGTTG TTTCTCTGCGT TATCCCTGTA TTCTGTGGAT AACCGTATTA CCGCCTTTGA
      bslI          haeIII/palI          nlaIV          haeI          haeIII/palI          nspHI          afIII          hinfI          aciI
      GCCGGAANA TGCCAAGGAC CGGAAACGA CCGGAACGA AGTGATCAAG AAAGGACGCA ATAGGGGACT AAGACACCTA TTGGCATAAT GCGGGAACCT

```

FIG. 10T

FIG. 10T

7101 CGTTGGCCGA TTCATTATATC CAGCTGTC
GCAACCGGCT AAGTAATTAG GTCGAC

scrFI
mvaI
ecoRII
dsav
bstNI
apyI|dd
bsaJI

	apyl(dcm+)	mspI	aciI	bsrBI	alul	nleIII	xmnI			
	bsaJI	hpaII					asp700			
7201	CACCCCAAGC	TTTACACTTT	ATGCTTCCGG	CTCGTATGTT	GTGTGGAATT	GTGAGCGGAT	AACAATTTCACACAGGAAC	AGCTATGACC	ATGATTACGA	
	GTGGGGTCCG	AAATGTGAAA	TACGAAGGCC	GAGCATACAA	CACACCTTAA	CACTCGCCTA	TTGTTAAAGT	GTGTCTTTTG	TGATACTGG	TACTAATGCT

tru9I
mseI
aseI/asnI/vspI
7301 ATTAA
TAATT

```
>length: 7305
```

INTERNATIONAL SEARCH REPORT

International Application No
PCT/US 95/09576

A. CLASSIFICATION OF SUBJECT MATTER

IPC 6 C12N15/64 C12N15/67 C12N15/85 C12N9/72 C12N5/10

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	DNA CLONING, VOLUME III, EDITED BY D.M. GLOVER, 1987 IRL PRESS, OXFORD, GB;, pages 189-212, A.M.C. BROWN AND M.R.D. SCOTT 'Retroviral vectors'	1-3,7,8
Y	see page 192, line 7 - page 196, line 5; figures 2,3 --- -/--	5,6, 9-12, 16-21

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

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Date of the actual completion of the international search

23 November 1995

Date of mailing of the international search report

08.12.95

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Hornig, H

INTERNATIONAL SEARCH REPORT

onal Application No
PCT/US 95/09576

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	CELL, vol. 37, no. 3, July 1984 CELL PRESS, CAMBRIDGE, MA, US;, pages 1053-1062, C.L. CEPKO ET AL. 'Construction and applications of a highly transmissible murine retrovirus shuttle vector' cited in the application	1-3,7,8
Y	pZIP-Neo SV(B)1 see figure 1	5,6, 9-12, 16-21
Y	--- MOL. CELL. BIOL., vol. 5, no. 3, March 1985 ASM WASHINGTON, DC, US, pages 431-437, A.D. MILLER ET AL. 'Generation of helper-free amphotrophic retroviruses that transduce a dominant-acting, methotrexate-resistant dihydrofolate reductase gene' see page 432, right column, line 5 - page 436, right column, line 7; figure 1	5,6, 9-12, 16-21
Y	WO,A,94 05784 (US) 17 March 1994 see the whole document	5,6, 9-12, 16-21
Y	--- EP,A,0 215 548 (ZYMOGENETICS INC ;UNIV WASHINGTON (US)) 25 March 1987 see the whole document	5,6, 9-12, 16-21
A	--- WO,A,92 17566 (GENENTECH INC) 15 October 1992 cited in the application see the whole document	1-21
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A	--- EP,A,0 260 148 (GENENTECH INC) 16 March 1988 see the whole document	1-21
A	--- EP,A,0 160 457 (GENENTECH INC) 6 November 1985 cited in the application see the whole document	1-21
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INTERNATIONAL SEARCH REPORT

International Application No
PCT/US 95/09576

C (Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	<p>PROC. NATL. ACAD. SCI., vol. 86, February 1989 NATL. ACAD SCI., WASHINGTON, DC, US;, pages 1041-1045, M. VIVAUD ET AL. 'A 5' splice-region G-C mutation in exon 1 of the human beta-globin gene inhibits pre-mRNA splicing: A mechanism for beta+-thalassemia' see the whole document -----</p>	1-4

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/US 95/09576

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		EP-A- 0385558	05-09-90
		HK-A- 8395	27-01-95
		JP-A- 60243023	03-12-85
		JP-A- 6040942	15-02-94
		NO-B- 174934	25-04-94
		SG-A- 3994	10-06-94
		US-A- 4965199	23-10-90

